

Culturable bacteria community development in postglacial soils of Ecology Glacier, King George Island, Antarctica

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Abstract Glacier forelands are excellent sites in which to study microbial succession because conditions change rapidly in the emerging soil. Development of the bacterial community was studied along two transects on lateral moraines of Ecology Glacier, King George Island, by culture-dependent and culture-independent approaches (denaturing gradient gel electrophoresis). Environmental conditions such as cryoturbation and soil composition affected both abundance and phylogenetic diversity of bacterial communities. Microbiocenosis structure along transect 1 (severe cryoturbation) differed markedly from that along transect 2 (minor cryoturbation). Soil physical

and chemical factors changed along the chronosequence (time since exposure) and influenced the taxonomic diversity of cultivated bacteria, particularly along transect 2. *Arthrobacter* spp. played a pioneer role and were present in all soil samples, but were most abundant along transect 1. Cultivated bacteria isolated from transect 2 were taxonomically more diverse than those cultivated from transect 1; those from transect 1 tended to express a broader range of enzyme and assimilation activities. Our data suggest that cryoturbation is a major factor in controlling bacterial community development in postglacial soils, shed light on microbial succession in glacier forelands, and add a new parameter to models that describe succession phenomena.

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Introduction

Glacial retreat is one of the most conspicuous signs of warming in polar regions. The resulting ice-free terrain offers an excellent opportunity for ecological research (Schmalenberger and Noll 2010). Deglaciated terrain presents a chronosequence of soil formation, primary succession, and ecosystem development. Distance from a retreating glacier reflects soil age, with vegetation-free sandy deposits at the front of the glacier contrasting with more differentiated soils further away (Schmidt et al. 2008). Antarctic soils that only recently became ice-free are considered to be nutrient limited (Sigler et al. 2002; Darmody et al. 2005), despite the influence of local melt-water streams, organic and inorganic inputs from marine aerosols, guano deposits in penguin and other seabird

rookeries, slow-growing cryptogamic plants, and marine algae deposited on the shore and dispersed by wind (Orchard and Corderoy 1983; Tibbles and Harris 1996; Beyer et al. 1999; Tockner et al. 2002; Grzesiak et al. 2009). Therefore, an urgent need exists to understand nutrient- and carbon-cycling processes in these recent and emerging environments.

Microorganisms are important in nutrient cycling in postglacial environments (Tscherko et al. 2003b) but are negatively affected by long periods of snow cover, low temperatures, and accompanying low water availability (Bardgett et al. 2007). Other studies, however, indicate that C and N pools increase in the first 20 years of succession, hinting at C and N fixation in these soils (Nemergut et al. 2007). The prevailing view is that photoautotrophs such as algae, mosses, lichens, and higher plants start primary succession on newly exposed surfaces by fixing carbon and promoting the accumulation of organic material necessary for development of heterotrophic communities (Walker and del Moral 2003). However, Tscherko et al. (2003a) show that heterotrophic microbial communities occur in newly exposed glacial terrain before photoautotrophic communities establish. Heterotrophic microbes clearly play an important role in the progress of primary succession in nutrient-limited soils in polar regions.

The main focus of the research described here is deglaciated soils in the vicinity of Ecology Glacier (King George Island, South Shetland Islands, Antarctica), whose retreat between 1978 and 2007 equates to 16.4 % (0.58 km²) of its 1978 surface area (Pudełko, Potocki, Angiel unpublished). Ecology Glacier moraines are influenced by cryoturbation and solifluction (an erosion process) (Blume et al. 2002a). Cryoturbation mainly occurs during the summer months when there is no snow on the soil surface, and the process is primarily based on the freeze–thaw cycle of water in the soil. Rock and mineral fragments break during these cycles (Blume et al. 2002b). The intensity of cryoturbation is evident in the fine silt and clay content of the soils (Seppelt 2002). During solifluction, thawed soil and rock move slowly downhill. Moreover, solifluction and cryoturbation hamper the establishment of permanent extended vegetation cover (Bölter and Blume 2002). High silt contents are also unfavorable for roots because intensive cryoturbation prevents stabilization of the soil fabric. Thus, silty to loamy soils affected by active cryoturbation lack plant cover (Beyer et al. 2002). Conditions in these soils may also change over short distances, and in many cases over meter scales. Mosses or lichens may develop in places where cryoturbation is absent, especially on sandy soils with gravel (Beyer et al. 2002). Frost impact is then reduced by a thick plant cover (Blume et al. 2002b).

Plant communities in ice-free areas around Ecology Glacier have been studied extensively (Olech 2002; Smykla et al. 2007; Parnikoza et al. 2008; Victoria et al. 2009), but little is known about bacterial succession, especially when there is ongoing cryoturbation. We hypothesize that varying soil conditions here have a large discriminating effect on the nature of the microbial community, leading to the establishment of dominant and robust ‘pioneer’ microbes. We thus investigated microbial community development on Ecology Glacier moraines, focusing especially on culturable heterotrophic bacteria in terms of their abundance, diversity, and morphophysiology.

Materials and methods

Sites and sampling

Surface soil samples (~0.5 kg) were collected in the vicinity of Ecology Glacier into sterile plastic bags using sterile plastic spatulas, in the 2008/2009 austral summer. One sample was also taken from Ecology Glacier itself as ice cores of the glacier’s surface with an auger (20 cm long, 2.5 cm diameter) (Table 1); pooled cores totaled ~0.5 kg and were allowed to melt in a refrigerator (~4 °C) before being processed for microbiological analyses. A duplicate set of cores was not thawed, but was stored at –20 °C in order to provide material for further research in Poland.

Soil at different distances from Ecology Glacier was collected in areas with less (transect 1, termed TR1) and more (transect 2, termed TR2) stable ground (Table 1): TR1, closer to the island’s rock massifs, lacks vegetation (samples collected here are termed 0, and A1 to E1); TR2, hosts plant communities of varying complexity (samples collected here are termed A2 to F2). Time since exposure of the ground along these transects was estimated by reference to maps (Birkenmajer 2002; Pudełko 2003, 2008); samples with the same letter names (e.g., A1 and A2) from both transects were exposed around the same time. Consecutive letters were given to samples collected at increasing distances from the glacier. Three ‘zones’ were identified: Zone I, exposed by glacial retreat in the last 5 years (samples 0, A1, A2); Zone II, exposed since 1978 and comprising mainly ground moraines almost devoid of vegetation cover, particularly along TR1, while terrain along TR2 hosts mosses (samples B1, B2, C1, C2); Zone III, comprising neoglacial moraines with more mature vegetation and the commonly occurring *Usnea*, indicative of an age of dozens to several 100 years (samples D1, D2, E1, E2, F2). Samples for TR1 in this zone were gathered from spots of ground that lacked vegetation, indicating cryoturbation influence.

Table 1 Characteristics of sampling points and soil composition

Sampling point (zone)	Site description	Age (years of exposure)	Coordinates	Vegetation	Distance from glacier (m)	Composition (%)		
						Stone/gravel	Sand	Silt/clay
G	Ecology Glacier surface	–	62°10'419"S; 58°28'501"W	None	–	0	0	0
0(I)	Moraine squeezed from beneath the ice front	0	62°10'246"S; 58°28'202"W	None	0	16.5	62.4	21.1
A1(I)	Contemporary moraine (old glacial river bank)	2	62°10'150"S; 58°28'221"W	None	171	18.9	64.6	16.6
A2(I)	Contemporary ground moraine; high shore of lagoon	5	62°10'168"S; 58°28'048"W	None	191	8.2	68.6	24.8
B1(II)	Top of an old glacial river bank	18	62°10'066"S; 58°28'175"W	None	327	12.3	64.8	22.9
B2(II)	Sandy, flat ground moraine	12	62°10'086"S; 58°28'060"W	Scattered mosses	315	9.6	65.6	24.8
C1(II)	Moraine on solid rock	24	62°10'048"S; 58°28'199"W	None	360	10.8	68.6	20.6
C2(II)	Moraine ridge	29	62°09'984"S; 58°28'069"W	First plants—rare mosses	492	19.7	64.1	16.1
D1(III)	Neoglacial lateral moraine	53	62°09'96"S; 58°28'192"W	None	514	24.8	55.8	20.1
D2(III)	Valley between moraines; stream interim	36	62°09'960"S; 58°28'067"W	Traces of <i>Sanionia uncinata</i> (moss) and <i>Leptogium puberulum</i> (lichen)	536	60.3	35.2	4.5
E1(III)	Oldest neoglacial moraine	>100	62°09'901"S; 58°28'357"W	None	647	9.6	64.7	25.6
E2(III)	Oldest neoglacial moraine (near cliffs)	>100	62°09'892"S; 58°28'358"W	Covered with <i>Usnea antarctica</i> (lichen)	664	36.2	59.3	4.5
F2(III)	Valley between moraines	45	62°09'939"S; 058°28'058"W	Community of <i>Sanionia uncinata</i> (moss)	577	6.7	80	13.3

Samples from transect 1: 0, A1, B1, C1, D1, E1; samples from transect 2: A2, B2, C2, D2, E2, F2

Measurement of soil components

Granulometric composition of the soil was determined by sieving, according to Polish Standards (PN-R-04033: 1998). Soil matrix (stone/gravel, sand and silt/clay) fractions were separated by sieve (2 mm mesh) and 0.05-mm mesh. Each fraction was weighed to determine percentage contribution. The pH of 1 g (wet wt.) of soil suspended in 100 ml sterile distilled water was determined with a Hanna Instruments HI 9025 pH meter; wet weight was determined directly after sampling. Dry weight was determined after 24-h incubation at 65 °C in a dry box with circulating air (Dowgiałło 1975). Dried soil was powdered and used for elementary CHN analysis (Carlo Erba CHNS-O-meter, EA 1108). Phosphorus was determined in dried material by the colorimetric meta-vanadate method, and other elements

were determined by atomic absorption spectroscopy (Tatur et al. 1997). Ash content was obtained by weighing the residue after combustion at 450 °C. The mean and standard errors (SEs) in water content, dry and wet weights, were determined from three measurements.

Bacterial abundance

Bacterial counts were determined in 1–2 g wet wt. soil and melted glacier ice. Soil was suspended in 20 ml sterile saline (0.85 %) in 100-ml sterile flasks with glass beads and shaken gently on a universal shaker (type 327) (120 rpm, 20 min, 5 °C). Suspensions were then stored in the refrigerator (4 °C) for 10–20 min to allow larger particles to settle. A decimal dilution series of the supernatant to 10⁻⁵ was prepared in sterile 0.85 % saline.

Sub-samples for total counts (TCs) were fixed with buffered formalin to a final concentration of 1 %. TCs were determined by epifluorescence microscopy using 4',6-diamidino-2-phenylindole (DAPI) on black Nuclepore polycarbonate 0.2- μm -pore-size filters (Porter and Feig 1980), under a Nikon E-200 microscope with a 100 W Hg lamp and 100 \times CFI 60 oil immersion objective, with a digital DS Cooled Camera Head DS-5Mc-U1, and a filter block of wavelengths EX 330–380, DM 400, BA 420. Images of fields were analyzed in Nikon NIS Elements BR 2.30 and MultiScan v. 14.02 (computer scanning systems). A minimum of 400 cells in 20 fields per sample were counted automatically in the image analysis system. Cell volume and total bacterial biomass were evaluated through image analysis (Sieracki et al. 1985; Świątecki 1997). Cell volume (V) was converted to bacterial biomass in terms of carbon (C), where C (fg) = $120 \times V^{0.72}$ (Simon and Azam 1989). Average values of three measurements using three independently prepared filters were calculated.

Photoautotrophs, including cyanobacteria and photoautotrophic eukaryotes, were counted under blue 450–490 nm (B-2A Nikon filter) and green 510–560 nm (G-2A Nikon filter) light excitation (Putland and Rivkin 1999). Cyanobacteria were distinguished from photoautotrophic eukaryotes because of the former's gold-yellow autofluorescence (Rassoulzadegan and Sheldon 1986).

A cell morph type diversity analysis describing the frequency of morphological forms in each size class, based on cell shapes (cocci, rods, curved forms) in five classes of volumes (<0.1; 0.1–0.2; 0.2–0.5, 0.5–1.0; >1 μm^3), was conducted with a modified Shannon index (Gurienowich 1995; Nübel et al. 1999).

Denaturing gradient gel electrophoresis analysis

The bacterial community in each sample was compared through denaturing gradient gel electrophoresis (DGGE). DNA was extracted from 10 g of each soil sample using the MegaSoil DNA isolation Kit (MoBio, USA), according to the manufacturer's protocol. The concentration of the extracted DNA was determined by measuring absorbance at 260 nm, and purity was estimated from the A_{260}/A_{280} ratio. The V2–V5 regions in the 16S rRNA gene in the template DNA were amplified by polymerase chain reaction (PCR) with the primer 341f with a GC clamp 5'-CGC CCG CCG CGC CCC GCG CCC GTC CCG CCG CCC CCG CCC GCC TAC GGG AGG CAG CAG-3', complementary to position 341–357, and 907r (CCG TCA ATT CMT TTG AGT TT) complementary to positions 926–907 (*Escherichia coli* numbering) (Crump et al. 2003, 2007). PCRs were performed in an MJ MiniTM Gradient Thermal Cycler (BioRad Laboratories, USA). The PCR (50 μl)

contained 1 \times PCR buffer, 200 ng bovine serum albumin (Sigma-Aldrich Co.), 2.5 mM MgCl_2 , 200 μM of each deoxynucleoside triphosphate, 0.25 μM (each) primer, 1.5 U *Taq* polymerase (Sigma-Aldrich Co.) and approximately 1–2 ng μl^{-1} template DNA. PCR conditions were as follows: initial denaturation (94 $^\circ\text{C}$, 5 min), followed by 10 touchdown cycles (denaturation 94 $^\circ\text{C}/1$ min, annealing 65 $^\circ\text{C}$ (decrease 1 $^\circ\text{C}/\text{cycle}$) and elongation 72 $^\circ\text{C}/3$ min), 20 standard cycles (94 $^\circ\text{C}/1$ min, 55 $^\circ\text{C}/1$ min, 72 $^\circ\text{C}/5$ min); the final elongation step was 5 min. Amplification products were visualized in a 1 % (wt/vol) agarose gel stained with ethidium bromide. Amplified DNA was separated using the D-Code Universal Mutation Detection System (BioRad Laboratories, USA) DGGE with a 6 % (wt/vol) 16 \times 16 cm polyacrylamide gel (37:1 acrylamide:bisacrylamide) in 1 \times TAE buffer (40 mM tris, 20 mM acetic acid, 1 mM EDTA, pH 8.0) with a 35–70 % denaturant gradient (where 100 % denaturant was 7 M urea and 40 % formamide). Electrophoresis was at constant voltage (60 V) for 17 h at 60 $^\circ\text{C}$. Electrophoresis products were stained by gently agitating the gel for 30 min in 100 ml 1 \times TAE containing 5 μl of a 1:10,000 dilution of SYBR Gold nucleic acid stain (Molecular Probes Inc., PoortGebouw, The Netherlands). DGGE gel images were analyzed using the Quantity One software on a GelDoc gel documentation system (BioRad Laboratories, USA). Gel bands were identified using GelCompar software to create a presence-absence matrix (Crump and Hobbie 2005). Each band represents a bacterial OTU (operational taxonomic unit).

Diversity of cultivated bacteria

Colony-forming units (CFU) enumerated by in a decimal dilution series of the sample suspension by the spread plate method on soil extract agar (SEA) were considered copiotrophic bacteria (CBcop), while those on 100 \times diluted SEA were considered oligotrophic bacteria (CBoligo) (Ogram and Feng 1997). The composition of SEA, per liter, is bacto beef extract (3 g), bacto-tryptone (5 g), bacto-agar (15 g) (Fenchel and Hemmingsen 1974) in 1 l of soil extract, prepared by autoclaving 200 g of Antarctic soil in 1 l water (Klement et al. 1990; Zdanowski et al. 2005). Inoculated plates were incubated in darkness at 4 $^\circ\text{C}$ and inspected every second to third day for 30 days for colony development and growth; this also enabled observation of particular colony types, and the rate at which CFU grew, from different sampling points on the moraine. CFU numbers either did not increase or increased only minimally, after 30 days. In that respect, CFU numbers by 7 days of incubation (considered fast-growing isolates) were compared with those determined periodically up to the 30th day of incubation (considered slow-growing

isolates). CFU abundance was calculated on the basis of the number of colonies present at the end of the incubation period, with the SE being determined from three replicates.

After 30 days of incubation, several colony types that differed in terms of size, color, shape, and other colony characteristics were selected per sample. The number selected per sample varied depending on the diversity. Colonies of a particular type were counted and their number considered during morphophysiological tests and taxonomic identification. Pure isolates derived from streaking colonies for isolation were passed through morphophysiological tests, including Gram reaction, pigment production, ability to grow at 4, 22, and 32 °C on a nutrient-rich medium, and responses in the API 20NE and API ZYM test kits. A total of 46 morphological physiological and biochemical tests were performed on each isolate. The following tests were conducted: ability to grow at 4, 22, 32 °C; morphological analyses, which comprised counts of pigmented colonies (PIGM), two microscopy tests—Gram reaction (GRAM), morphology (ROD); the API 20NE system which allows nine biochemical tests: reduction in nitrate to nitrite (NO₃), and to nitrogen (N₂); indole production (TRP); fermentation of glucose (GLU_F); arginine dihydrolase (ADH); urease ((URE); β-glucosidase (ESC); gelatin hydrolysis (GEL); cytochrome oxidase (OX); and 12 tests for assimilation of carbohydrates as sole carbon sources; glucose (GLU); arabinose (ARA), mannose (MNE); mannitol (MAN); *N*-acetyl-glucosamine (NAG); maltose (MAL); gluconate (GNT); caprate (CAP); adipate (ADI); malate (MLT); citrate (CIT); phenyl-acetate (PAC); the API ZYM system tests for the activities of 18 constitutive enzymes: alkaline phosphatase (PHAL); esterase—C4 (EST4); esterase lipase—C8 (EST8); lipase—C 14 (LIP14); leucine arylamidase (LEU); valine arylamidase (VAL); cystine arylamidase (CYS); trypsin (TRY); chymotrypsin (CHY); acid phosphatase (PHAC); α-galactosidase (AGAL); β-galactosidase (BGAL); β-glucuronidase (BGLUR), α-glucosidase (AGLUC); β-glucosidase (BGLUC); *N*-acetyl-α-glucosaminidase (NACET); α-mannosidase (AMANN); α-fucosidase (AFUC).

A binomial test compared the frequency of positive responses among the pure cultures from each transect (McDonald 2009).

Cultivated bacteria were provisionally identified through the nucleotide sequence of a fragment of their 16S rRNA gene. Genomic DNA was extracted from a colony of each pure culture; the colony was suspended in 50 μl TE buffer (10 mM Tris–HCl, 1 mM EDTA) with 0.6 % Tween 20 and incubated in 80 °C for 20 min. Upon cooling, the suspension was emended with 0.6 μg of proteinase K/ml and incubated at 42 °C for 3 h. Genomic DNA was purified

with a Heliconius A&A Biotechnology DNA Clean-up kit. Amplification was performed by PCR (Zdanowski et al. 2004); two sets of primers were used (U515F 5'-GTGCC AGCMGCCGCGGTAA-3' and 1492R 5'-GGTTACCT TGTTACGACTT-3'; L63F 5'-CAGGCCTAACACAT GCAAGTC-3'; and P1387R 5'-GGGCGGTGTGTACA AGGC-3') (Wang and Qian 2009; Żmuda-Baranowska 2010). Nucleotide sequences were compared by BLAST with those in the NCBI database (Johnson et al. 2008) and using the RDP classifier online program (Cole et al. 2009). Sequences were deposited in GenBank under accession numbers JF728899–JF728980.

Statistics

Simple regression analysis between abundance of the three bacterial fractions (TC, CBcop, and CBoligo) and environmental factors was carried out in STATISTICA v. 9 (StatSoft). Multiple stepwise regression analysis was conducted for three main microbiological parameters (dependent variables): TC, CBcop, and CBoligo. Models were built for two systems: transect 1, transect 2. Analyses were carried out in STATISTICA version 9, (StatSoft).

A canonical correspondence analysis was conducted using a statistical package for Windows v. Canoco 4.5 (ter Braak and Šmilauer 2002). Microbiological data for each sample were analyzed in relation to the environmental background—'all other data'. To confirm taxonomic diversity in both transects, a DGGE analysis of 10 samples was performed. This allowed use of canonical correspondence analysis to determine differences, but also the impact of the factors determining diversity.

Results

Environmental factors and sampling sites characteristics

Sampling sites were situated along two transects extending 0 m ('0') to 672 m ('E2') from the edge of Ecology Glacier (Table 1). Soil from TR1 was silty, with sand and fine gravel. Soils from areas with vegetation along TR2 were mixed with plant debris. Ice from the surface of Ecology Glacier was a third sample.

The content of stones and gravel accounted for 6.7–60.3 % (mean 18.4 ± 3.9), sand from 35.2 to 68.7 % (mean 63.2 ± 2.6); dust + silt from 4.5 to 25.6 % (mean 18.6 ± 1.8). The structural components of the terrain between the zones differed slightly along TR1. Terrain furthest from the glacier along TR2 was visually distinguishable.

Total microbial abundance and diversity

Along TR1, with little vegetation, highest TC values were recorded along the edge of the glacier at sampling points 0 ($29.48 \pm 5.77 \times 10^7$) and A1 ($39.78 \pm 2.52 \times 10^7$), with the lowest at point B1 ($3.15 \pm 0.00 \times 10^7$) (Table 2). Along TR2, characterized by richer vegetation, TC values were significantly lower (lowest at A2, $0.41 \pm 0.14 \times 10^7$), yet they increased with distance from the glacier, reaching a maximum at sampling point F2 ($59.06 \pm 1.15 \times 10^7$).

Highest values of bacterial biomass (Cbact) in TR2 were at a point overgrown with vegetation, while along TR1 the highest values were recorded in the soil at the leading edge of the glacier (Table 2). A modified Shannon index calculated on the basis of cell morphotypes determined by light microscopy ranged from 0.610 to 1.105 along TR2 and correlated with changes in TC (Spearman's rank correlation, $p < 0.05$). No such correlation was observed in TR1.

Photoautotrophic microorganisms, including numerically dominant cyanobacteria, diatoms, and unidentified objects of $<50 \mu\text{m}$ in diameter exhibiting red fluorescence, were detected throughout both transects as the smallest TC fraction, with the TAC to TC ratio ranging from 0.05 % at A1 to 31.8 % at D2. Along transects, both the total number of photoautotrophs and cyanobacteria increased from very low values in the youngest Zone I and reached a maximum in the oldest Zone III (Table 2). Note that no cyanobacteria were found at point A1 (TR1). In sample '0', taken at the edge of the glacier front, relatively high numbers of mainly coccoid cells ranging in size from 0.5 to 4.95 μm (probably cyanobacteria) were detected. Filamentous forms dominated in the third zone, in contrast to the coccoid forms prevailing in Zone I. High numbers of diatoms were observed in the third zone, and many of them were covered by bacteria.

Denaturing gradient gel electrophoresis

Molecular analysis by DGGE showed significant differences in taxonomic structure of the soils (Fig. 1). Collectively, 76 OTUs were defined. From the soil samples collected along transects I and II, 40, and 64 OTUs were defined, respectively. In TR1, OTU numbers per sampling site were as follows: '0'-17, A1-28, B1-27, C1-19, D1-16, and E1-4. Along TR2, OTU numbers were as follows: A2-10, B2-20, C2-11, D2-30, E2-25, F2-43.

The microbial structure of the soil, soil age, and degree of vegetation cover correlated significantly (Fig. 2). In TR1, the greatest abundance (TC) and taxonomic uniqueness (number of OTUs) were detected in the youngest soil formations, located directly in the glacier foreland. The lowest diversity was determined in the neoglacial Zone III.

Within soils in the area of TR2, the highest taxonomic diversity occurred in the oldest soils (Zone III), characterized by the highest vegetation coverage, while the lowest was determined in an area exposed just a few years ago (Zone I).

Culturable bacteria (CB) abundance, diversity, and distribution

Along TR1, CB counts followed the same pattern of TC, with highest numbers in freshly exposed soil ('0') and lowest at point B1. In TR2, highest CFU counts were recorded at the oldest exposed sites (E2), and the lowest at points A2 and B2 (Table 2).

Most colonies developed in the later stage of incubation (between 7 and 30 days) and were defined as 'slow-growers', as opposed to the minority of CB that grew in the early stages of incubation (up to 7 days), referred to as 'fast-growers'. The proportion of 'fast-' to 'slow-growers' was relatively high along TR1's youngest Zone I (Table 3). In contrast, the highest such proportions in TR2 occurred in the area farthest from the glacier, in places covered with vegetation.

Significant differences occurred among the morpho-physiological characteristics of bacteria isolated along the two transects (Fig. 3). With the exception of twelve characteristics, most features were significantly more frequent in bacteria cultivated from TR1. Psychrophiles (those whose growth was inhibited above 20 °C) occurred along both transects, but were clearly more prevalent in TR2 (80 %). Note that psychrophiles accounted for only 60 % of the total number of strains isolated from the glacier surface. Almost 40 % of bacteria cultures from TR1 could reduce NO_3^- to NO_2^- , while less than 10 % of such strains from TR2 could do so. Up to 30 % of bacteria cultures prepared from TR1-expressed cytochrome oxidase activity (OX+). A high percentage of oxidase positive strains were isolated from the sample in direct contact with the glacier at that time.

The number of cultivated isolates that could assimilate single carbon sources varied between transects 1 and 2. The mean number of positive responses in assimilation of 12 different carbohydrates and organic acids, meaning those that could use these as sole sources, was statistically higher among bacteria cultivated from TR1. Relatively few of the strains isolated from the surface of the glacier assimilated these carbohydrates. The percentage of cultivated strains expressing glycoside hydrolase activities was highest among bacteria cultivated from TR1. Strains from Zones II and III were most likely to express glycosidases. A low percentage of strains possessing glycosidases was observed on the surface and in contact with the glacier. TR1 hosts bacterial communities characterized by a broader range in

Table 2 Differences in bacterial total count (TC), total photoautotrophic count (TAC), cyanobacteria count (CYAN), cultivated copiotrophic bacteria (CBcop), cultivated oligotrophic bacteria (CBoligo), CB/TC ratios, total bacterial carbon (TCB), morphotype diversity index (Shannon index), loss on ignition (OM_{LOI}), pH, carbon (C), nitrogen (N), C/N ratios, phosphorus (P), and mineral compounds content in samples used for microbiological analysis

Variables	Zone I					Zone II					Zone III				
	Surface of the glacier	A1	A2	B1	B2	C1	C2	D1	D2	E1	E2	F2			
TC ($\times 10^7$)	0.01 ± 0.00	29.48 ± 5.77	39.78 ± 2.52	0.41 ± 0.14	3.15 ± 0.00	0.88 ± 0.11	6.20 ± 0.27	0.61 ± 0.09	14.54 ± 0.75	0.85 ± 0.28	4.21 ± 0.51	5.06 ± 0.32	59.06 ± 1.15		
TAC ($\times 10^7$)	0.087	0.018	0.031	0.026	0.018	0.052	0.035	0.15	0.27	0.24	0.21	0.21	0.21		
CYAN ($\times 10^7$)	0.062	0.0	0.022	0.009	0.004	0.009	0.013	0.079	0.17	0.19	0.14	0.14	0.14		
CBcop ($\times 10^5$)	0.01 ± 0.00	4.56 ± 2.24	1.38 ± 0.54	0.51 ± 0.05	0.03 ± 0.02	0.16 ± 0.06	0.74 ± 0.43	0.35 ± 0.04	1.46 ± 0.15	0.41 ± 0.23	0.98 ± 0.11	8.55 ± 3.65	1.19 ± 0.86		
CBoligo/TC (%)	0.80	0.15	0.03	1.23	0.01	0.19	0.12	0.57	0.10	0.48	0.23	1.69	0.02		
CBoligo/TC ($\times 10^5$)	0.0007	2.58 ± 1.13	0.71 ± 0.34	0.03 ± 0.01	0.03 ± 0.00	0.86 ± 0.74	0.17 ± 0.09	0.18 ± 0.06	0.43 ± 0.06	0.22 ± 0.05	0.21 ± 0.15	4.18 ± 0.89	0.97 ± 0.78		
CBoligo/TC (%)	0.07	0.09	0.02	0.08	0.01	0.98	0.03	0.29	0.26	0.05	0.83	0.02	0.02		
TCB ($\mu\text{gC} \times \text{g}^{-1}$ soil)	3.15 ± 0.90	11.29 ± 0.79	4.06 ± 0.69	0.88 ± 0.11	1.97 ± 0.34	3.06 ± 0.61	2.85 ± 0.32	1.33 ± 0.17	9.17 ± 0.64	1.18 ± 0.18	1.98 ± 0.70	12.93 ± 0.73	30.99 ± 0.63		
Shannon index	0.92 ± 0.06	0.93 ± 0.03	1.02 ± 0.07	0.61 ± 0.01	0.71 ± 0.03	0.67 ± 0.04	1.12 ± 0.03	0.75 ± 0.08	0.97 ± 0.04	0.79 ± 0.07	0.95 ± 0.05	1.05 ± 0.02	1.10 ± 0.00		
OM _{LOI} (%)	2.71 ± 0.11	1.75 ± 0.01	1.95 ± 0.26	2.68 ± 0.12	2.25 ± 0.15	2.72 ± 0.15	2.72 ± 0.06	2.56 ± 0.43	2.62 ± 0.14	2.17 ± 0.21	4.07 ± 0.18	6.15 ± 0.45	10.93 ± 1.04		
pH (in H ₂ O)	5.7	9.2	8.9	8.8	8.3	7.7	8.5	7.4	8.2	8.2	8.2	5.3	6.3		
H ₂ O (%)	14.89	14.02	12.69	12.58	11.95	15.27	8.99	10.5	8.54	15.95	17.39	28.05	28.05		
C (%)	0.69	0.33	0.14	0.70	0.80	0.22	0.90	0.38	0.95	1.18	1.61	1.83	1.83		
N (%)	0.01	0.02	0.01	0.03	0.04	0.01	0.09	0.02	0.08	0.04	0.10	0.10	0.10		
C/N	53.24	17.75	18.11	20.73	17.99	22.08	9.99	24.57	12.05	29.19	16.42	18.04	18.04		
P (mg P ₂ O ₅ /100 g soil)	2.09	1.70	1.52	1.2	1.70	1.47	1.70	1.70	2.12	1.77	1.75	1.17	1.17		
K (mg/100 g soil)	8.44	4.82	3.62	4.82	2.41	2.41	0.60	1.21	1.21	1.21	2.41	2.41	2.41		
Na (mg/100 g soil)	134.80	35.05	40.44	21.57	20.22	22.92	14.83	22.92	18.87	29.66	28.31	29.66	29.66		
Mg (mg/100 g soil)	3.32	1.66	3.32	1.66	0.83	0.83	0.83	0.83	1.66	1.66	1.66	1.66	1.66		
Ca (mg/100 g soil)	1.4	2.8	5.6	0.7	0.7	0.7	0.7	0.7	0.7	0.7	0.7	0.7	2.8		

Transsect 1: A1, B1, C1, D1, E1; transect 2: A2, B2, C2, D2, E2, F2

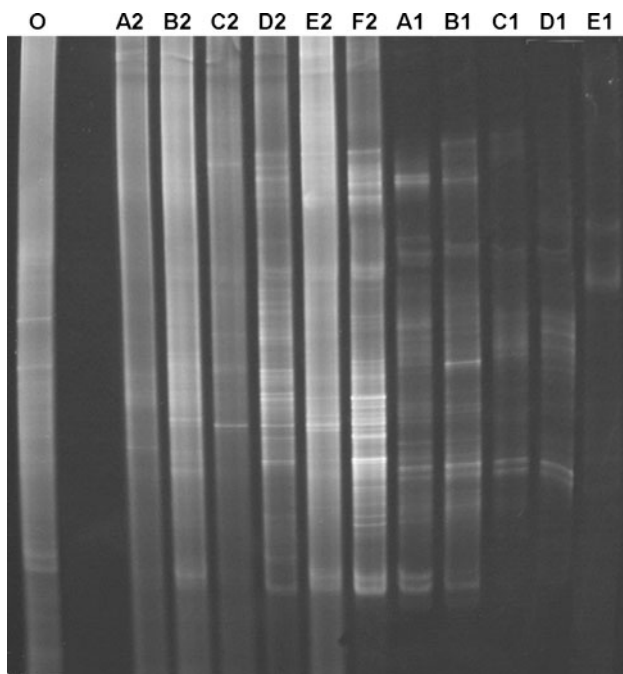


Fig. 1 Denaturing gradient gel electrophoresis band patterns of bacterial communities in samples. Letters at the top are sample labels. See text for details

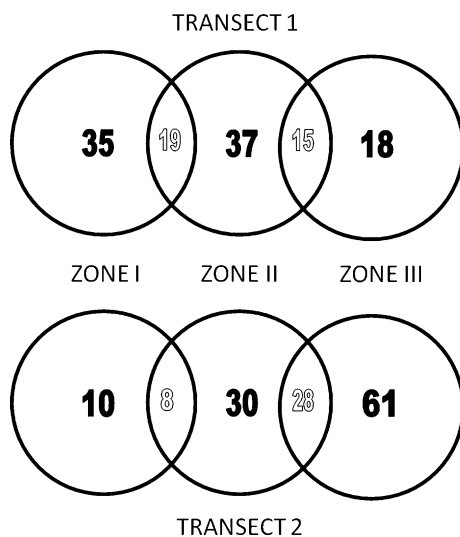


Fig. 2 Number of operational taxonomic units (OTU) by denaturing gradient gel electrophoresis in relation to soil age (zone). Black numbers all OTUs in a zone, white numbers OTUs shared with neighboring zone

constitutive enzyme activity and sole carbon source metabolism, unlike that from stable TR2. Only 11 of the 46 traits occurred with equal frequency in both groups. These were the rarely expressed traits.

The 82 pure cultures whose 16S rRNA genes were partially sequenced fell into 56 OTUs at the 98 % sequence identity level (Table 4). Seventy-five cultures

were classified to the genus level. Actinobacteria were the most numerous and also the most diverse (41 strains, 26 OTUs). Less numerous and less diverse were Bacteroidetes (11 strains, 9 OTUs), Alphaproteobacteria (12 strains, 6 OTUs), Betaproteobacteria (8 strains, 6 OTUs), Firmicutes (6 strains, 3 OTUs), and Gammaproteobacteria (4 strains, 4 OTUs). The nearest neighbors in BLAST searches were from cold environments, such as Antarctica, the Arctic, Himalayas, Alps, and deep-sea sediments.

The dominant phylum among bacteria grown from TR1 and the surface of the glacier was the Actinobacteria (Table 4) represented by *Frigoribacterium* spp.—found on the glacier, *Leifsonia*—present mainly at the interface between the glacier and *Arthrobacter*—mostly isolated from Zones II and III of the first transect, but also Zone III of the second transect.

Proteobacteria were most abundant among bacteria cultivated from TR2. The Alphaproteobacteria (*Sphingomonas*, *Brevundimonas*) occurred in large quantities at sites that became ice-free in the last 10 years, but their numbers decreased with distance from the glacier. The Betaproteobacteria represented by *Polaromonas* were detected on the glacier and along TR2. *Herminiimonas* (Betaproteobacteria) numerically dominated at sites F2 and E2—the earliest exposed as the glacier retreated. *Pseudomonas* (Gammaproteobacteria) was also present at the latter site. Bacteroidetes (*Flavobacterium*, *Pedobacter*) were present in moderate quantities in sites in early and middle stages of glacier foreland succession. Firmicutes (*Sporosarcina*) were found in both transects but only in areas far from the glacier.

Statistics

Simple regression analysis between abundance of the three bacterial fractions (TC, CBcop, and CBoligo) and environmental factors showed that in TR1, these significantly correlated with the soil pH (Table 5). Water and organic carbon content correlated positively with TC only in TR2, whereas the mineral content (P_2O_5 , K, Mg, Ca) in TR1. Time since exposure negatively correlated with TC in TR1 and positively with CB in TR2. Relatively large soil particles (stones, sand) and plant cover had an overall positive effect on the microbiocenosis.

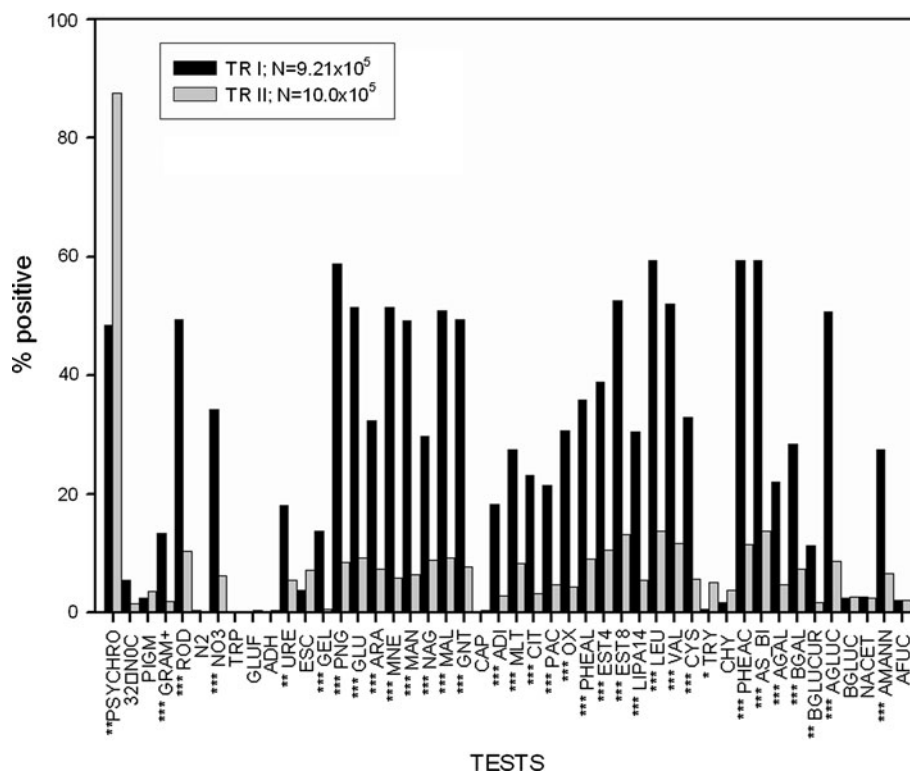
Multiple regression analysis identified those independent variables whose influence was most significant (Table 6). In TR1, Ca and P_2O_5 contents were significant for TC, and Na content for the CB. In TR2, it was the water content and soil age (time since exposure) that had a major impact on the microbiocenosis.

The statistical analysis of principal components (CCA) showed a significant correlation between the taxonomic structure (of OTUs) and selected biological and

Table 3 Proportions of fast-growing (up to 7th day) to slow-growing (days 8–30) colonies in each sampling zone along the two transects

Zone	Sampling points	Transect 1		Sampling points	Transect 2	
		% of fast-growing			% of fast-growing	
		Copiotrophic	Oligotrophic		Copiotrophic	Oligotrophic
I	0	6.52	2.99			
	A1	9.39	1.37	A2	NO	NO
II	B1	NO	NO	B2	6.25	NO
	C1	0.46	NO	C2	0.1	NO
III	D1	NO	NO	D2	10.17	1.99
	E1	0.45	NO	E2	6.41	4.11

NO no colony growth



TESTS

Fig. 3 Differences in the incidence of morphophysiological positive tests among the cultivated bacteria isolated from two transects: *NS* not significant, *TR* transect, *N* number of isolates calculated on the basis of the counts of similar colonies of which representative was tested. Significance level ($*p < 0.05$, $**p < 0.01$, $***p < 0.001$) between frequency of positive responses to tests shown by bacterial communities. Tests conducted: ability to grow at 4, 22, 32 °C; pigmented colony counts (PIGM), Gram stain (GRAM), cell morphology (ROD); reduction of nitrate to nitrite (NO₃), and to nitrogen (N₂); indole production (TRP); fermentation of glucose (GLU_F); arginine dihydrolase (ADH); urease ((URE); β-glucosidase (ESC); gelatin hydrolysis (GEL); cytochrome oxidase (OX); assimilation tests:

glucose (GLU); arabinose (ARA), mannose (MNE); mannitol (MAN); *N*-acetyl-glucosamine (NAG); maltose (MAL); gluconate (GNT); caprate (CAP); adipate (ADI); malate (MLT); citrate (CIT); phenyl-acetate (PAC); enzyme activity tests: alkaline phosphatase (PHAL); esterase—C4 (EST4); esterase lipase—C8 (EST8); lipase—C 14 (LIP14); leucine arylamidase (LEU); valine arylamidase (VAL); cystine arylamidase (CYS); trypsin (TRY); chymotrypsin (CHY); acid phosphatase (PHAC); α-galactosidase (AGAL); β-galactosidase (BGAL); β-glucuronidase (BGLUR), α-glucosidase (AGLUC); β-glucosidase (BGLUC); *N*-acetyl-α-glucosaminidase (NACET); α-mannosidase (AMANN); α-fucosidase (AFUC)

physicochemical parameters of soils (Fig. 4). The main factors determining the microbial structure of the soil in TR1 were physicochemical properties, especially pH. In

soils covered with vegetation (TR2), significant factors affecting the taxonomic structure were mainly biological, particularly the abundance of photoautotrophs (TAC).

Table 4 Taxonomic affiliation of representative strains determined by BLAST search of 16S rRNA gene nucleotide sequences

Category	Sampling site	GenBank accession number (JF728...)	Most related sequence (GenBank)	GenBank accession number	Similarity (%)	Taxonomical position of isolate (RDP classifier)	Origin of GenBank sequence
Actinobacteria	D1	902	<i>Arthrobacter</i> sp. Lc5-1	GU733458.1	98	Family <i>Micrococcaceae</i> / genus <i>Arthrobacter</i>	Antarctic sediments
	A2, B1	923, 976	<i>Arthrobacter</i> sp. R-37013	FR691391.1	100		Lake, Antarctica
	B1, C1, E2	968, 969, 911	<i>Arthrobacter</i> sp. PN21am	HQ396622.1	99		Mont Blanc snow with Saharan dust layer
	C2	918	<i>Arthrobacter</i> sp. PSA A20	DQ628958.1	99		High Arctic glacier
	D1, F2,	934, 974	<i>Arthrobacter sulfonivorans</i> strain T-52	HQ202868.1	99		Western Kunlun Mountains, China
	D2	935	<i>Arthrobacter scleromae</i> strain T-50	HQ202866.1	99		Western Kunlun Mountains, China
	D1	906	<i>Arthrobacter</i> sp. RKS6-4	GQ477171.1	99		Roopkund Glacier of Himalayas, India
	D1, E1	904, 970, 971	<i>Arthrobacter</i> sp. Lc30-1	GU244360.1	99		Antarctic sediments
	D1, E1	910, 956	<i>Arthrobacter</i> sp. Nj-43	AM491459.1	99		Antarctica
	F2	938	Uncultured bacterium clone AKAU3832	DQ125730.1	98		Uranium contaminated soil
	E1	952	<i>Arthrobacter</i> sp. SH-82B	FN377716.1	99		Svalbard:Kongsfjord region
	E2	940	<i>Arthrobacter</i> sp. TP-Snow-C2	HQ327111.1	99		Tibetan Plateau, China
	0, A1, B1	905, 954, 963–966	<i>Rhodoglobus</i> sp.R-36762	FR691401.1	99	Family <i>Microbacteriaceae</i> / genus <i>Leifsonia</i>	Lake, Antarctica
	0	903, 967, 955	<i>Leifsonia rubra</i> clone SE44	AY771758.1	99		Arctic
	D2, E1	975, 909	<i>Leifsonia kafniensis</i> strain KFC-22T	AM889135.1	99		Kafni glacier, Himalaya
	D2	936	<i>Salinibacterium</i> sp. TP-Snow-C10	HQ327119.1	99		Tibetan Plateau, China
	G	929	Arctic sea ice bacterium ARK10173	AF468440.1	100	Family <i>Microbacteriaceae</i> / genus <i>Frigoribacterium</i>	Arctic sea ice
	G	931	Antarctic bacterium strain G064	EU636019.1	100		Collins glacier, Antarctica
	B1	932	<i>Agreia</i> sp. SK3	GQ406816.1	99	Family <i>Microbacteriaceae</i> / genus <i>Agreia</i>	Northeast Greenland
	G	925	Glacier bacterium FJS11	AY315169.1	100	Unclassified <i>Microbacteriaceae</i>	Glacier, New Zealand
	C2	919	Uncultured bacterium clone RT_58	EU644208.1	100		Tundra soil, Siberia
	D2	901	<i>Subtercola boreus</i> strain R-36477	FR682694.1	98		East Antarctica
	F2	937	<i>Subtercola</i> sp. TP-Snow-C8	HQ327117.1	99		Tibetan Plateau, China
	E2	939	<i>Janibacter</i> sp. 0705C10-1	HM222661.1	99	Family <i>Intrasporangiaceae</i> / genus <i>Janibacter</i>	Deep-sea sediments
	B1	933	<i>Intrasporangiaceae</i> bacterium MI-1.2 V18	FN550117.1	98	Unclassified <i>Intrasporangiaceae</i>	Marion Island, sub-Antarctica
	E2	941	<i>Rhodococcus</i> sp. gap-e-79	DQ530478.1	99	Family <i>Nocardiaceae</i> / genus <i>Rhodococcus</i>	Oil contaminated Antarctic sea ice

Table 4 continued

Category	Sampling site	GenBank accession number (JF728...)	Most related sequence (GenBank)	GenBank accession number	Similarity (%)	Taxonomical position of isolate (RDP classifier)	Origin of GenBank sequence
α-Proteobacteria	0	907	<i>Brevundimonas</i> sp. R-37030	FR691409.1	100	Family <i>Caulobacteraceae</i> /genus <i>Brevundimonas</i>	Lake, Antarctica
	B1, A2	973, 961, 977, 962, 899	<i>Brevundimonas</i> sp. R-37014	FR691411.1	100		Lake, Antarctica
	C2	916, 960	<i>Brevundimonas staleyi</i> strain R-36554	FR682698.1	99		East Antarctica
	C2, F2	959, 915	<i>Sphingomonas</i> sp. R-36533	FR682705.1	100	Family <i>Sphingomonadaceae</i> /genus <i>Sphingomonas</i>	East Antarctica
	D2	942	Glacier bacterium FXS25	AY315166.1	98		Glacier, New Zealand
	B2	951	Uncultured bacterium clone SINN640	HM128677.1	99	Unclassified <i>Sphingomonadaceae</i>	Tibetan Lake, China
β-Proteobacteria	G	927	<i>Polaromonas</i> sp. 1006	EF423322.1	99	Family <i>Comamonadaceae</i> /genus <i>Polaromonas</i>	Glacier, China
	G	928, 953	Betaproteobacterium LH14	DQ535025.1	99		Ice-covered lake, McMurdo Dry Valleys, Antarctica
	A2, B2	924, 978	<i>Polaromonas</i> sp. R-36732	FR691430.1	99		Lake, Antarctica
	D2	943	<i>Polaromonas</i> sp. Cr4-35	HM583567.1	98		Glacier cryoconite, Austria
	F2	944	<i>Hermiimonas</i> sp. 166	GU213388.1	99	Family <i>Oxalobacteraceae</i> /genus <i>Hermiimonas</i>	Freshly deglaciated granite sand, Central Alps
	E2	935	<i>Hermiimonas</i> sp. 4-96	GU213333.1	97	Unclassified <i>Oxalobacteraceae</i>	Freshly deglaciated granite sand, Central Alps
γ-Proteobacteria	E1	900	<i>Pseudomonas</i> sp. RKS6-8	GQ477175.1	100	Family <i>Pseudomonadaceae</i> /genus <i>Pseudomonas</i>	India: Roopkund Glacier, Himalayas
	E2	914	<i>Pseudomonas</i> sp. 254	GU213414.1	99		Damma glacier (Central Alps) freshly deposited granite sand
	A2	922	Uncultured <i>Acinetobacter</i> sp. clone XZZLH29	EU703475.1	99	Family <i>Moraxellaceae</i> /genus <i>Acinetobacter</i>	High-altitude lakes, eastern Tibetan plateau
	B2	921	<i>Luteimonas</i> sp. R-37032	FR691433.1	99	Family <i>Xanthomonadaceae</i> /genus <i>Lysobacter</i>	Lake, Antarctica
Bacteroidetes	A1	946	<i>Flavobacterium micromati</i> strain R-36963	FR691440.1	99	Family <i>Flavobacteriaceae</i> /genus <i>Flavobacterium</i>	Antarctica: Transantarctic Mountains, Forlidas Pond
	B2	947	<i>Flavobacterium</i> sp. KOPRI 25152	FJ544919.1	97		Antarctica
	B2	948	<i>Flavobacterium</i> sp. RKS1-3	GQ477164.1	99		India: Roopkund Glacier, Himalayas
	C1	MOR13	Uncultured bacterium clone TP-Snow-14	HQ327168.1	99		China: Tibetan Plateau
	C1	950	<i>Flavobacterium</i> sp. ice-oil-522	DQ521398.1	98		Antarctic sea ice
	G	930	<i>Sejorgia</i> sp. KOPRI 25179	FJ544917.1	99	Family <i>Flavobacteriaceae</i> /genus <i>Chryseobacterium</i>	Arctic
	G	926	<i>Chryseobacterium</i> sp. FRGDSA	AY468464.1	98		Diseased aquatic animals
	B2	920	<i>Pedobacter</i> sp. KFC-76	EF459523.1	99	Family <i>Sphingobacteriaceae</i> /genus <i>Pedobacter</i>	Soil, Kafni Glacier, Himalaya
	E2, F2	912, 958, 979	<i>Pedobacter cryoconitis</i> strain A37	NR_025534.1	99		Alpine glacier cryoconite

Table 4 continued

Category	Sampling site	GenBank accession number (JF728...)	Most related sequence (GenBank)	GenBank accession number	Similarity (%)	Taxonomical position of isolate (RDP classifier)	Origin of GenBank sequence
Firmicutes	D1, E2, F2	957, 972, 913, 980	<i>Bacillus</i> sp. Nj-19	AM491453.2	100	Family <i>Planococcaceae</i> /genus <i>Sporosarcina</i>	Antarctica
	D2	949	Uncultured bacterium clone HAW-RM37-2-B-1017d-UE	FN563243.1	99		Mesophilic biogas digester
	C2	917	<i>Bacillus simplex</i> strain EQH11	FJ999940.1	100	Family <i>Bacillaceae</i> /genus <i>Bacillus</i>	China: Qinghai, Haibei Tibetan Autonomous Prefecture

Table 5 Bacterial abundance (bacterial total count—TC, cultivated copiotrophic bacteria—CBcop, cultivated oligotrophic bacteria—CBoligo) relationships with physical, chemical, and environmental factors

	Transect 1; N = 18			Transect 2; N = 18		
	TC	CB-cop	CB-oligo	TC	CB-cop	CB-oligo
pH	0.74*	0.69*	0.78*	-0.4	-0.62*	-0.73*
H ₂ O	0.01	0.16	0.18	0.92*	0.23	0.31
C	-0.32	0.05	0.04	0.69*	0.42	0.54*
N	-0.41	-0.46	-0.49*	0.44	0.36	0.41
C/N	0.2	0.81*	0.82*	0.37	0.14	0.23
P ₂ O ₅	0.52*	0.78*	0.72*	-0.75*	0.05	0.01
K	0.52*	0.61*	0.74*	0.15	0.13	0.15
Na	0.45	0.85	0.9	0.23	0.16	0.11
Mg	0.41	0.71*	0.79*	0.8*	-0.06	-0.08
Ca	0.85*	0.24	0.31	0.19	-0.2	-0.3
Age	-0.55*	-0.3	-0.43	0.2	0.75*	0.82*
m a.s.l.	-0.55*	-0.33	-0.47*	0.14	0.75*	0.8*
Stones	0.5*	0.27	0.23	-0.35	0.22	0.17
Sand	-0.16	-0.26	-0.18	0.57*	-0.03	0.02
Clay	-0.72*	-0.15	-0.19	-0.14	-0.46	-0.46
Plants				0.7*	0.38	0.45

Significant regression (*) at the $p < 0.05$ level

Discussion

Bacterial communities in postglacial soils of Ecology Glacier (King George Island, Antarctica) were investigated in terms of abundance, physiology, and taxonomic affiliation. Our findings provide a description of changes in the glacier's edge, as well as a comparison of soil microbiocenoses in two transects with different ground stability and plant coverage along lateral moraines.

Glacier ice–foreland interactions

Biogeochemical data were in line with the literature reports on postglacial areas (Bardgett et al. 2007; Nemergut et al.

Table 6 Factors determined on the basis of multiple regression that best explain variability in bacterial numbers for the three main microbiological parameters (dependent variables), bacterial total count—TC, cultivated copiotrophic bacteria—CBcop, cultivated oligotrophic bacteria—CBoligo, in two systems: transect 1 and transect 2

Transect 1		
For TC	Ca—72 %; P ₂ O ₅ —7.4 %,	$R^2 = 0.85, p < 0.01$
For CBcop	Na—72 %,	$R^2 = 0.72, p < 0.05$
For CBoligo	Na—72 %,	$R^2 = 0.82, p < 0.05$
Transect 2		
For TC	H ₂ O—83 %, K—5 %, Age—5 %, Clay—1 %	$R^2 = 0.95, p < 0.05$
For CBcop	Age 56 %	$R^2 = 0.72, p < 0.05$
For CBoligo	Age 67 %, Clay 9 %	$R^2 = 0.82, p < 0.05$

2007). However, total organic carbon was lower than in the high Arctic (Schmidt et al. 2008). The high total carbon contents at point '0' (close to the glacier) were striking, considering the lower levels 100–200 m away from the glacier, where microbial photosynthesis would be expected to provide organic carbon. This might be attributable to the enriching influence of supraglacial meltwater (such as from cryoconite holes) or ancient subglacial sediment carbon (Stibal et al. 2006; Bardgett et al. 2007). The highest TC and CFU counts were determined in a zone closest to the glacier (point '0'). The CB/TC ratio was also high here and might be supported by enrichment of the soil by supraglacial meltwater bacteria (Christner et al. 2003; Mueller and Pollard 2004; Grzesiak et al. 2009). Supraglacial meltwater input may also explain the relatively high count of cyanobacteria at point '0', while their numbers in the newly exposed glacial terrain of Zone I and II, farther from the glacier, showed low values. This is in line with Tscherko et al. (2003a) who reported that heterotrophic microbial communities occur in newly exposed glacial terrain before autotrophic communities. Initial taxonomic structure comparison analysis of soil from the Ecology Glacier foreland and from the glacier's surface ice by DGGE showed they share 30 % of OTUs (by taxonomic affiliation) with Zone I. However, this number decreases

markedly with distance from the glacier, with 6 and 4 % of shared OTUs for Zones II and III, respectively. This indicates that glacial runoff derived bacteria play a role only in early stages of postglacial succession.

Ribosomal DNA analysis of bacteria cultivated from soil close to the glacier, and from the glacial ice, did not detect any common genera between the two. This might reflect the very different pH values of the glacial ice and freshly exposed forefield soil, or high Na⁺ content of the soil. The high TC and composition of the culturable bacteria community in this soil were similar even 2 years after deglaciation (sampling point A1). Therefore, high culturable bacteria counts might be derived from below the glacier. For example, culturable bacteria counts in basal sediment beneath a high Arctic glacier may reach 10⁷ ml⁻¹ (Skidmore et al. 2000; Cheng and Foght 2007). Supraglacial melt water enrichment thus appears more nutritional than microbial in nature. This explanation is supported by the high numbers of copiotrophic and fast-growing bacteria (*r* strategy) at the glacier edge. A similar pattern of development per day of CB in the Ecology Glacier forefield has been reported before (Zdanowski and Węgleński 2001). These observations are consistent with the hypothesis of Sigler and Zeyer (2004), that bacteria able to quickly respond to disturbance and tolerate stress are more abundant in younger soils, where conditions favor *r*-strategists, typical of opportunistic bacteria (Sigler et al. 2002). It should be noted that we incubated samples for cultivation

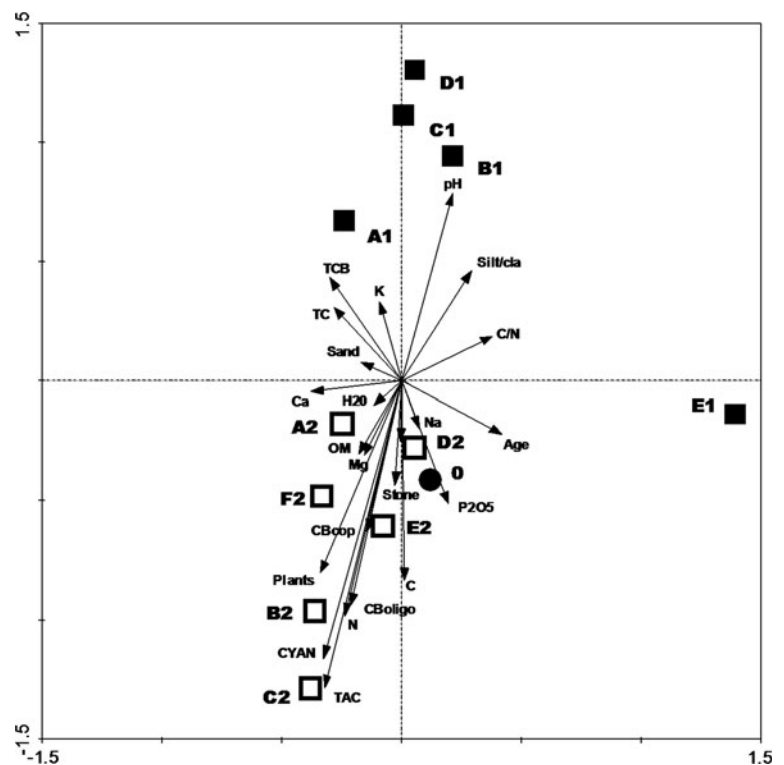
of bacteria under conditions that differed from those used by Sigler et al. (2002), since we avoided incubation temperatures that would be inappropriate for most psychrophilic and psychrotrophic microbes, that is, we used 4 °C instead of 25 °C.

Microbial community along transect 2

A classic succession pattern in terms of soil elements, microbial abundance, and bacterial taxonomic richness was evident along transect 2 (Schipper et al. 2001; Sigler et al. 2002; Schmidt et al. 2008). In this nutrient-limited postglacial environment, the contents of biogenic elements gradually increased. Total and photoautotrophic cell counts followed this trend, reaching a maximum in well-formed soils in Zone III. Sattin et al. (2009) describe a similar observation based on clone libraries during their work on Mendenhall Glacier foreland soils (USA). Counts at sampling points E2 and F2 could have been influenced by significant numbers of edaphic ciliates which control bacterial populations (Mieczan, unpublished).

Stable ground conditions along TR2, with minor freeze-thaw events, have enabled development of plant coverage, which according to the CCA analysis positively influenced bacterial communities along this transect (Fig. 4). Community richness determined by DGGE analysis increased with distance from the glacier (Fig. 2); a phenomenon recognized in ecological studies where the number of

Fig. 4 Canonical correspondence analysis of denaturing gradient gel electrophoresis bands and environmental data. *Empty square* transect 1, *solid square* transect 2, *solid circle* control, bacterial total count (TC), total photoautotrophic count (TAC), cyanobacteria count (CYAN), cultivated copiotrophic bacteria count (CBcop), cultivated oligotrophic bacteria count (CBoligo), total bacterial carbon (TCB), loss on ignition (OM)



species in an undisturbed environment tends to increase with age (Walker and del Moral 2003). The presence of plants and microbial photoautotrophs is especially beneficial for the culturable fraction of the community as indicated by the relatively high CB to TC ratio. This is also reflected in the phenotypic properties of strains isolated from TR2. Plant coverage on TR2 promoting more stable and lower temperatures would explain significantly higher numbers of psychrophilic bacteria along this transect. Moreover, bacteria cultivated from TR2 expressed a narrow range of constitutive enzymes and utilized only a few carbon sources (Fig. 3), implying a stable and perhaps single source of nutrients. Cultivated bacteria from transect 2, that was less affected by cryoturbation than transect 1, were taxonomically more diverse in terms of 16S rRNA gene nucleotide sequences. That no genus dominated the chronosequence indicates favorable conditions for bacterial colonization, and greater likelihood of successful allochthonous introductions.

Members of the generally oligotrophic genus *Brevundimonas* were cultivated from Zones I and II in both transects, but reached very high numbers only in transect 2. *Brevundimonas*, although detected in low numbers at the glacier's edge, developed large populations later in the succession process. Members of this genus have been isolated from various polar habitats, for example, glacial snow in summer (Amato et al. 2006), Antarctic soil and lakes, Arctic deep ice core and snow cover, subglacial outflow waters (Cheng and Foght 2007), and also from the Antarctic aerosol over King George Island (Miteva et al. 2004; Amato et al. 2006; González-Toril et al. 2009). *Polaromonas* sp. seems to be the only surface ice derived genus among the CB bacteria in the moraine soil older than 30 years. However, it was only cultivated here from transect 2, so cryoturbation and large temperature shifts in TR1 may not be favorable for members of this genus.

A non-opportunistic growth strategy and dependence on plant-specific growth factors may be the reason for the high percentage of strains lost during recultivation from late succession period sites (C2, D2, E2, F2). Those samples did, however, yield bacteria from genera considered plant-associated and/or plant-dependent, such as *Sporosarcina*, *Bacillus*, *Pedobacter*, *Herminiimonas*, *Sphingomonas*, and *Rhodococcus*, psychrophilic members of which have been isolated from the lichen and moss rhizospheres in polar regions (Männistö and Häggblom 2006; Lang et al. 2007; Selbmann et al. 2010). Furthermore, *Herminiimonas* sp. has been reported to assimilate oxalic acid produced by lithophytic lichens (Lang et al. 2007). Thus, *Herminiimonas* spp. at the humus-rich F2 sampling point may be associated with lichen communities and their production of secondary metabolites. *Sphingomonas* spp. (Alphaproteobacteria) also are often found in association with plants

(in the rhizosphere) (White et al. 1996); some are nitrogen fixers, suggesting a potential role as plant symbionts (Doty et al. 2009). *Pedobacter* spp. have been isolated from soils, permafrost, glacial, and near glacial environments and from decaying lichens (Bai et al. 2006; Männistö and Häggblom 2006; Hansen et al. 2007; Yoon et al. 2007; Simon et al. 2009). Members of the *Sphingobacteriaceae* such as *Pedobacter* sp. can utilize carbohydrates that are plant derived (Liebner et al. 2008).

Microbial community along transect 1

Transect 1 was selected because of its lack of vegetation, on the premise that this could influence microbial communities. However, it was quickly recognized as undergoing severe cryoturbation, and a process that in many ways impairs the postglacial soil development and bacterial succession determined along TR2. Cryoturbation consequences for soil and microbial community development are clearly visible. The main factors affecting soil microbiocenoses are most likely the freeze–thaw cycle and lack of plant cover. The latter results *inter alia* in a greater daily temperature range. The CCA analysis, however, showed that pH in these circumstances is of crucial importance to the bacterial community. In this respect, a lack of rock-weathering organic acids produced by some mosses and lichens would explain why the pH of the soils lacking vegetation along TR1 never dropped below 8.

Despite cryoturbation and the lack of plant cover, TC and CB values were relatively high in all TR1 soil samples. Counts of edaphic ciliates as low as four individuals per gram of soil would not be expected to significantly affect the size of the bacterial communities (Mieczan, unpublished). Taxonomic diversity among the bacterial community diminished with distance from the glacier (Fig. 2). Pure cultures of bacteria isolated from TR1 expressed a wide range of enzymes and assimilated various carbon compounds, useful characteristics for life in an environment where biogenic elements show a 'patchy' distribution.

Members of the *Leifsonia* genus numerically dominated the CB community in the very early stages of succession in transect 1. This is rather surprising considering their copiotrophic and photoautotroph-associated nature (Evtushenko and Takeuchi 2006). It is more likely that the isolates cultivated here were of subglacial origin, as no representatives of the genus were found in surface ice or anywhere else along the transect. For example, recently described *Leifsonia* species were isolated from cold habitats such as soil near a Himalayan Glacier, Southern Ocean sediment (Pindi et al. 2009), Antarctic pond cyanobacterial mat (Reddy et al. 2003), and Spitsbergen permafrost (Hansen et al. 2007), suggesting this genus is well adapted

to life in polar and alpine ecosystems. One of its ecological functions could be supplying autotrophs with phosphorus and metal ions of biological importance such as Fe, Mn, Ca, K, and Mg by weathering of poorly available sources (insoluble mineral deposits). In this respect, the bioleaching properties of *Leifsonia* spp. have been widely recognized (Hansen et al. 2007; Delvasto et al. 2008; Frey et al. 2010).

A sharp drop in total bacterial abundance and change in the taxonomic composition of the bacteria cultivated occurred between sampling points A1 and B1, with a shift from *Leifsonia* to *Arthrobacter*. We posit that this may be a response to an environmental factor such as soil mineral composition change (causing a pH drop), indicating a limit of glacial influence. Due to their extreme resistance to drying, starvation, temperature shifts, pH changes, and nutritional versatility (Jones and Keddie 2006; Mongodin et al. 2006), *Arthrobacter* strains replace *Leifsonia* spp. in such conditions and are clearly recognized as pioneer bacteria in this study. Along TR1, for example, these bacteria comprised up to 98 % of the CB counts, testimony to their durability, and survival under cryoturbation conditions that would be lethal to other secondary succession bacteria. However, a succession pattern exists even in this genus (Table 4). A penguin rookery close to Ecology Glacier (Trivelpiece et al. 1990; Ciaputa and Sierakowski 1999) is a likely source of *Arthrobacter* spp. Studies have already shown *Arthrobacter* species engaged in guano decomposition are abundant, nutritionally versatile, and adaptable (Zdanowski et al. 2005; Żmuda-Baranowska 2010).

Summary

Our findings indicate that recently deglaciated soils host abundant microorganisms and biogenic elements which appear to be the result of sub- and supraglacial influences. This unique environment maintains its character even several years after deglaciation. However, the microbiological contribution to later succession stages seems limited. A more intense allochthonous colonization begins when glacier-associated factors no longer determine microbial community composition, and ‘pioneer-microorganisms’ start to flourish. Diversity and properties of postglacial microbiocenoses result not only from soil age and composition, but also from differences in soil stability. An undisturbed ground enables vegetation growth and promotes a more taxonomically diverse microbial community, with types of bacteria being plant-associated and less resistant to temperature fluctuations. Cryoturbation, which hinders colonization by plants, causes a gradual

biodiversity impoverishment and emergence of dominance by just a few types of bacteria which are nutritionally versatile and very resistant to abiotic conditions. Therefore, we recognize cryoturbation as an extremely important environmental factor shaping microbial community structure and development in postglacial soils.

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