- Diethyl carbonate as green extraction solvent for chlorophenols determination with dispersive
   liquid-liquid microextraction.
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# 9 ABSTRACT

10 The principles of green analytical chemistry indicate that the search for greener organic 11 solvents for extraction applications is crucial. In this study diethyl carbonate (DEC) is proved 12 to be green solvent, as it is relatively nontoxic, obtainable from renewable resources and it is 13 biodegradable. Here it is applied as extraction solvent for chlorophenols determination in 14 water samples with dispersive liquid-liquid microextraction. The multiobjective optimization for 7 chlorophenols is done with design of experiment combined with Derringers desirability 15 16 function. The optimized parameters on extraction step are 300 µL of DEC volume, 0.5 mL of 17 methanol, 8 mL of water sample and 7.5% of inorganic salt addition. The values of LOQ are 0.01 and 0.3  $\mu$ g L<sup>-1</sup>, CVs are between 7.1-17.3 % for 7 CPs. In this study DEC is proved to be 18 19 promising, green solvent, applicable in analytical extractions.

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21 Keywords: DLLME; chlorophenols; green analytical chemistry; green solvent;
22 microextraction

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### 25 1. Introduction

One of the aims of green analytical chemistry is introduction of greener solvents to analytical practice.<sup>1</sup> Supercritical fluids application requires costly and relatively nonstandard apparatus. The application of ionic liquids also requires nonstandard solutions as their lack of volatility makes them incompatible with gas chromatography. Therefore, there is strong need to develop solventless analytical methodologies<sup>2</sup> or methodologies based on extraction with greener organic solvents.<sup>3</sup>

32 The definition of green solvent is not straightforward. The three main aspects that need to be 33 considered are: health issues in case of exposure, environmental concerns in case of their release and safety during their storage, handling and application.<sup>4</sup>Within each aspect of 34 35 greenness there are few assessment criteria to be considered and because of unavailability of data the assessment is even harder to perform. There are systems that help to select the 36 37 solvent within green chemistry framework, that are known as solvent selection guides. They combine many greenness criteria into easy to interpret output.<sup>5</sup> The closest homologue to 38 39 diethyl carbonate - dimethyl carbonate is assessed as green solvent, causing only some problems in the safety area.<sup>6</sup>Diethyl carbonate (DEC) is a linear carbonate ester that is 40 classified as organic carbonate.<sup>7</sup> The application of this colorless, transparent liquid mainly 41 involves being an alternative fuel or fuel additive due to its high oxygen content.<sup>8,9,10</sup> It 42 43 influences reduction CO<sub>2</sub> and particulates emissions from engines. Many studies describe DEC application as an electrolyte for lithium ion batteries.<sup>11,12</sup> It could also be an excellent 44 solvent, that is why it is widely used in pharmaceutical products, fertilizer, pesticide and dyes 45 manufacture.<sup>13,14</sup> Organic carbonates are characterized by many outstanding properties, for 46 47 instance polarity, low toxicity, low bio-accumulation or relatively high biodegrability.

48 Dispersive liquid-liquid microextraction (DLLME) is a liquid sample preparation technique 49 that applies the mixture of two organic solvents to perform the extraction rapidly.<sup>15,16</sup>The first 50 solvent plays the role of extractant and has to be immiscible with water. The first extraction 51 solvents applied in DLLME were chlorinated solvents and other non green organic solvents. 52 The development of DLLME mode applying solvents lighter than water allowed to apply more organic solvents and these solvents are of greener nature.<sup>17</sup> The examples of such 53 solvents can be hexane, methyl tert-butyl ether or toluene. As these solvents are indicated by 54 55 green solvent selection guides as problematic the search for green extraction solvents is still 56 urgent problem. The second solvent applied in DLLME is dispersive solvent, which has to be 57 miscible with both aqueous sample and extraction solvent. In analytical practice the most commonly applied dispersive solvents are methanol, acetone and acetonitrile,<sup>18</sup> two first are 58 59 considered to be green and the third one is claimed to be problematic.

60 Chlorophenols (CPs) are group of environmental pollutants that are toxic and hardly undergo 61 degradation processes in both aerobic and anaerobic conditions.<sup>19</sup> They are used as wood 62 preservation agents and are substrates in pesticides production processes. They can be formed 63 during water disinfection process or during pulp bleaching. Therefore, their occurrence is 64 detected in surface and groundwaters.<sup>20</sup>

The aim of the study is to show the applicability of diethyl carbonate as analytical extraction solvent. There is a strong need to search for new, green solvents to be applied in analytical chemistry. In this study the applicability is shown at the example of chlorophenols determination with dispersive liquid-liquid microextraction.

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#### 2. Materials and Methods

2.1. Reagents

The analytical standards of: 2,4-dichlorophenol (2,4-DCP), 2,6-dichlorophenol (2,6-DCP),
2,4,6-trichlorophenol (2,4,6- TCP), 2,3,4-trichlorophenol (2,3,4-TCP), 2,3,4,5tetrachlorophenol (2,3,4,5-TeCP), 2,3,4,6-tetrachlorophenol (2,3,4,6-TeCP),

pentachlorophenol (PCP).Diethyl carbonate (anhydrous, 99%) and acetic anhydride used for
derivatization were purchased from Sigma-Aldrich (Germany). 2,4,6-tribromophenol (2,4,6TBP) was used as an internal standard and K<sub>2</sub>SO<sub>4</sub>were also purchased from Sigma-Aldrich
(Germany).

Stock standard solution of CPs was prepared in methanol (1, 2 or 5 mL), with appropriate concentration levels and stored in a refrigerator in the dark. All aqueous solutions were prepared with appropriate amount of ultrapure water obtained from a Mili-Q<sup>®</sup> apparatus (Merck KGaA, Germany).

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#### 85 2.2. Instrumentation

Centrifugation of samples was carried out by centrifuge supplied from Eppendorf SG 86 87 (Germany). Chromatographic analysis of CPs was performed with GC8000 Top (USA), 88 electron capture detector ECD850 (TermoQuest, Italy) and on-column injector. Separation 89 and identification of chlorophenols were obtained by applying the DB-5 column (30m x 90 0.32mm x 0.25µm) (Agilent Technolgies, USA) and Guard Column (6 m x 0.32mm) (Zebron 91 Phenomenex, USA). The initial temperature was 130 °C (held for 8 min) and followed by heating to 250 °C at rate 15 °C min<sup>-1</sup> held for 1 min. Hydrogen, as carrier gas, was supplied 92 93 by hydrogen generator (HG2600, Claind Italy) and inlet pressure of 130 kPa was applied. 94 Nitrogen (purity 99,999%) was used as make-up gas for ECD and detector operated at 330 °C.

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### 2.3. Dispersive liquid-liquid microextraction

In order to perform the simultaneous DLLME and derivatization, in the optimized conditions
8 mL of an aqueous solution was transferred into 10-mL glass test tube (OMNILAB,
Germany) with screw cap and PTFE/silicone membrane (Agilient Technologies, USA) and

sample was treated with mixture of 0.5 mL of methanol and 0.3 mL of diethyl carbonate that contained 50  $\mu$ L of acetic anhydride. Such mixture was rapidly injected into aqueous sample with using 2-mL syringe (Polfa, Poland). A cloudy solution was formed and centrifugation for 2 min at 3200 rpm was applied to separate extraction phase from sample. A 2  $\mu$ L of extract was carefully taken from the surface of the sample in the vial with a 10- $\mu$ L microsyringe (Hamilton, USA) and injected into GC for analysis. For easy recovery of this light extraction solvent from the surface of sample, specially designed sampler was applied.<sup>21</sup>

107 To clean the glassware after centrifugation it was rinsed with tap water and washed with soap. 108 Then it was soaked overnight in 5%  $HNO_3$  solution, rinsed very thoroughly with deionized 109 water and placed in an ultrasonic bath (Bandelin Sonorex, Germany) for ~15 minutes. Then 110 the glassware was oven-dried at 120 °C in furnace (Poland).

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### 112 2.4.Experimental design

113 Central composite design is used to optimize the volumes of sample, extraction and dispersive 114 solvents of DLLME applied for isolation and enrichment of CPs. It is designed for these 115 three factors as it is shown in the table 1. The type dispersive solvent optimised in previous 116 step in this study, while the extraction solvent is assumed to be diethyl carbonate.

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Table 1. Factors and their values applied in experimental central composite plan

			1			
	-1.68	-1	0	1	1.68	units
Vextr	216	250	300	350	384	μL
Vdisp	0.33	0.5	0.75	1	1.17	mL
Vsamp	2.64	4	6	8	9.36	mL

120 The structure of central design plan is presented in the table 2. All the calculations to obtain 121 central composite design plan and figure of response surface are drawn with Statistica 122 software.

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# Table 2. Design matrix for central composite plan

No experiment	V extr	V disp	V sampl
4	-1	1	1
14	0	0	1.68
6	1	-1	1
1	-1	-1	-1
9	-1.68	0	0
2	-1	-1	1
12	0	1.68	0
15 C	0	0	0
18 C	0	0	0
8	1	1	1
3	-1	1	-1
13	0	0	-1.68
17	0	0	0
7	1	1	-1
16 C	0	0	0
5	1	-1	-1
10	1.68	0	0
19C	0	0	0
11	0	-1.68	0

#### 125 2.5.Desirability Function

126 To perform multi-objective optimization usually a desirability function was applied. In this 127 case multi-objectiveness is expressed as selection of optimal parameters for all 7 analytes at 128 the same time. Together with completely desirable response (d = 1) and completely 129 undesirable response (d = 0), for every optimization criterion (peak area to be maximized) 130 desirability function equation are defined. In this case linear functions between desirable and 131 undesirable response are selected. The desirable response (d = 1) is above 90 % of the range 132 between minimum and maximum values of response for given analyte. The undesirable 133 response (d = 0) is below 10 % of the range between minimum and maximum values of 134 response for given analyte. Then functions for every optimization goal are combined into a 135 single score called global desirability (D) as it is presented with equation 1. If any of 136 optimized analytes has undesirable response, then D = 0. Application of desirability function approach allows for giving different relative importance of criteria  $(r_1, r_2 \hdots r_n)$  but in this 137 138 study equal importance for all analytes is assigned.

139	$D = \left(d_1^{r_1} \times \ d_2^{r_2} \times \dots \times d_n^{r_n}\right)^{\frac{1}{\Sigma r i}} \tag{1}$
140	Global desirability function is input data to draw response surface for optimization of sample
141	volume and dispersive and extraction solvents and obtain their optimized values.
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143	3. Results and Discussion
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145	3.1.Chromatographic separation
146	The chromatographic separation was aimed at minimization of separation time and separation
147	of analytes peaks from solvent peak and impurities that elute mainly at the beginning of the
148	chromatographic run. As some impurities elute in the region of retention times of analytes, the
149	optimization of chromatographic separation was also aimed at assurance that none of analytes
150	co-elutes with impurities. Less ghost peaks are obtained indeed during blank sample analysis
151	in the after 5th minute and slightly less peaks before this time. These impurities may originate
152	from derivatization reagent and/or side reactions and from dispersive solvent.

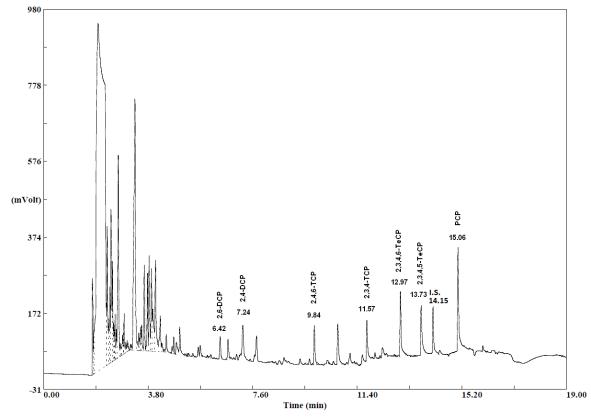
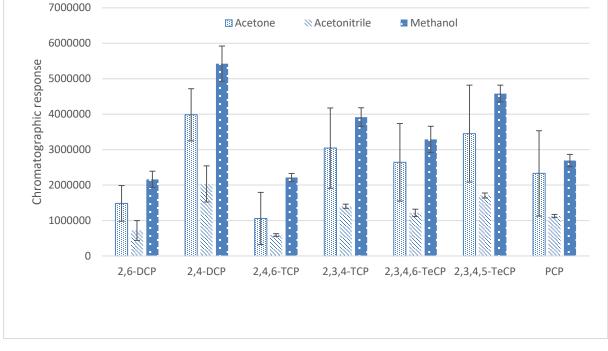


Figure 1. Chromatogram showing separation of 7 CPs and I.S. -2,4,6-TBP.The concentrations are 1µg L<sup>-1</sup> (except for I.S. and PCP 0.5 µg L<sup>-1</sup>) spiked in ultrapure water.

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158 3.2.Selection of dispersive solvent

159 The selection of dispersive solvent was based on comparison of means of peak areas for the analysis of standard solutions with the concentration of  $1\mu g L^{-1}(0.5 \mu g L^{-1}$  for PCP). The 160 volume of disperser solvent was 1 mL, while the volume of diethyl carbonate was 0.3 mL 161 162 while the volume of sample was 8 mL. Acetone gave moderate chromatographic response but 163 the precision (n = 3) expressed as coefficient of variance (CV) was between 18.49 and 69.56 164 %. The application of acetonitrile gave poor chromatographic response but CVs were between 3.92 and 8.52 for all CPs except for dichlorophenols, for 2,6-DCP it was 38.90 % and for 2,4-165 166 DCP it was 25.07 %. Methanol gave the best response and the values of CV were between 167 5.11 and 11.24 % for all analytes. Methanol was selected as the most appropriate dispersive 168 solvent and the procedure was optimized for diethyl carbonate and methanol as extraction and



169 dispersive solvents. The results are presented in the figure 2.

- 171 Figure 2. The chromatographic responses (n = 3) and standard deviations for different 172 dispersive solvents.
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174 3.3. Optimization of sample, extraction solvent and dispersive solvent volumes

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176 Above-described central composite design was applied to obtain the optimal volumes. The concentration of analytes in the samples used for optimization was 1  $\mu$ g L<sup>-1</sup>. The values of 177 178 global desirability for given extraction conditions are presented in the table 3. The worst 179 conditions are for experimental point 13 with very low volume of sample. For this point 4 out 180 of 7 analytes gave undesirable response. 181

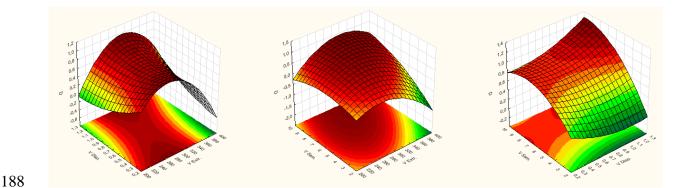
Table 3. The extraction conditions with their values of calculated global desirability.

No	V Extr. [µL]	V Disp. [mL]	V Sam. [mL]	D
4	250	1	8	0.634
14	300	0.75	9.36	0.894
6	350	0.5	8	0.523
1	250	0.5	4	0.747

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9	216	0.75	6	0
2	250	0.5	8	0.980
12	300	1.17	6	0.915
15 C	300	0.75	6	0.936
18 C	300	0.75	6	0.898
8	350	1	8	0.927
3	250	1	4	0.623
13	300	0.75	2.64	0
17	300	0.75	6	0.788
7	350	1	4	0
16 C	300	0.75	6	0.462
5	350	0.5	4	0.209
10	384	0.75	6	0.231
19C	300	0.75	6	0.778
11	300	0.33	6	0.610

Based on global desirability calculated for all experimental design conditions response surface was drawn and it is presented in figure 3. The response is not desirable at low volume of sample and for extreme values of extraction solvent volume. The volume of dispersive solvent does not influence much the desirability of response.



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Figure 3. The response surface for central composition design. Global desirability for 7 CPs.

191 The polynomial equation for the response function is following:

D =-2.94424+0.0337895\* Vextr -0.000079424\* Vextr<sup>2</sup> -3.25828\* Vdisp +0.489155\* Vdisp<sup>2</sup> 0.0950929\* Vsam -0.0202707\* Vsam<sup>2</sup>0.0066808\* Vextr \* Vdisp +0.00124701\* Vextr \*
Vsam +0.0983837\* Vdisp \*Vsam

196 Critical values that are obtained with solving of response surface equation are V extr. = 297 197  $\mu$ L; V disp. = 0.51 mL; V sam. = 7.99 mL. Global desirability value obtained by the solution 198 is D = 0.869. As a result, 300  $\mu$ L of DEC, 0.5 mL of methanol and 8 mL of water sample 199 were selected as optimal parameters.

- 200
- 201 3.4. Salting out effect

To investigate the effect of the salt on the extraction efficiency K<sub>2</sub>SO<sub>4</sub> was used. No salt 202 203 added, 5 %, 7.5 % and 10 % of salt were considered as procedural options. There was no 204 influence on the extraction efficiency however the addition of salt resulted in much easier 205 separation of phases during centrifugation phase. Without adding of the salt dispersed system 206 partially remained despite performing centrifugation. As a result the volume of extraction 207 phase collected at the surface of the sample was so small that it was inconvenient or 208 sometimes impossible to collect extract for analysis. It was decided to apply addition of salt 209 resulting in 7.5% of salt concentration.

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3.5. Metrological parameters

Limit of detection (LOD) was estimated on the basis of signal to noise ratio. The value of LOD was for S/N = 3. Limit of quantitation (LOQ) was calculated according to the relation LOQ = 3 \* LOD. Coefficient of variance (CV) was determined at the level of concentration 1  $\mu g L^{-1}$  for n = 5 repetitions. Linearity range was investigated up to 5  $\mu g L^{-1}$  as these are maximal concentrations of CPs in environmental waters. The results of determination of metrological parameters of the procedure are presented in table 4.

Table 4.Basic validation parameters of the procedure

Analyte	LOD [µg L <sup>-1</sup> ]	LOQ [µg L <sup>-1</sup> ]	CV [%]	Linearity range $[\mu g L^{-1}]$
2,6-DCP	0.05	0.15	17.3	LOQ - 5
2,4-DCP	0.1	0.3	9.0	LOQ - 5

2,4,6-TCP	0.01	0.03	13.6	LOQ - 5
2,3,4-TCP	0.01	0.03	10.5	LOQ – 5
2,3,4,6-TeCP	0.005	0.015	7.2	LOQ – 5
2,3,4,5-TeCP	0.005	0.015	9.9	LOQ – 5
PCP	0.0033	0.01	7.1	LOQ – 5

## 221 3.6. Comparison with other procedures

222 Table 5 shows the comparison of LODs, CVs and extraction solvents with previously reported 223 analytical procedures for CPs determination, based on solvent extraction. Almost all given 224 procedures incorporate solvents that are commonly considered as less green ones, some of 225 them causing environmental problems. Other sample preparation techniques based on solvent 226 extraction are hollow fiber liquid phase microextraction (HF-LPME) and single drop 227 microextraction (SDME). The values of LODs, obtained with other than LC based 228 procedures, are in similar range as in this study. On the other hand, other procedures are 229 characterized by better precisions. This is probably due to the fact that we have selected lower 230 concentrations to determine CVs, but such concentration is typical for CPs present in the 231 environmental waters. Our methodology is characterized by poor precision for 2,6-DCP, 232 neglecting this analyte, the poorest precision, expressed as CV, would be 13.6 %. The detailed 233 investigation on extraction solvents greenness indicates that DEC can only be compared with 234 butyl acetate, that is recommended by solvent selection guides. Additionally, procedure with its application presented by Bagheri et al.<sup>24</sup> is characterized by equally good and satisfactory 235 236 value of LOD. Thus, option is another available option for CPs determination in water 237 samples baring in mind green analytical chemistry approach.

Table 5. CPs determination in water samples - comparison with other procedures based on solvent extraction.

Procedure	Analytes	Extraction solvent	LOD [µg L <sup>-1</sup> ]	CV [%]	EF	Reference
HF- LPME- GC-MS	4-CP	toluene	3.2	10	30	22

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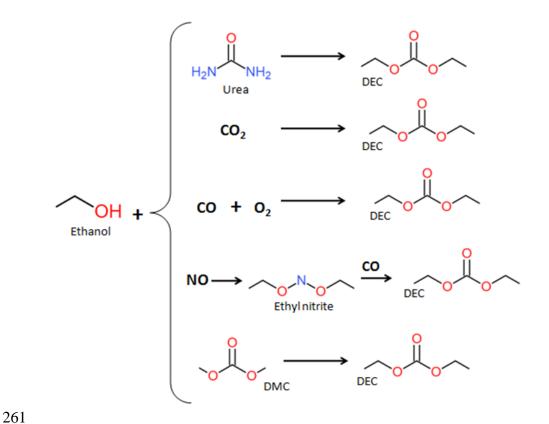
HF- LPME- GC-MS	2,4-DCP, PCP	toluene	0.015	< 13	123, 136	23
SDME- GC-MS	2-CP, 4-CP, 24-DCP, 246-TCP	butyl acetate	0.01 – 0.021	< 10	71-134	24
SDME- LC-UV- VIS	2-CP, 2,4-DCP, 2,6-DCP	methyl cyanide	6 – 23	< 9	-	25
DLLME- GC-ECD	2-CP, 3-CP, 4-CP, 23-DCP, 2,4-DCP, 2,5-DCP, 2,6-DCP, 3,4-DCP,3,5-DCP, 2,3,4-TCP, 2,4,5-TCP, 2,3,5-TCP, 2,3,6-TCP, 3,4,5-TCP, 2,3,4,5- TeCP, 2,3,4,6-TeCP, 2,3,5,6-TeCP, PCP	chlorobenzene	0.001 – 1	<5	287-906	26
DLLME- GC-ECD	2,4-DCP, 2,6-DCP, 2,4,6-TCP, 2,3,4-TCP, 2,3,4,5-TeCP, 2,3,4,6- TeCP, PCP	diethyl carbonate	0.0033 - 0.1	< 17.3	?	This study

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## 243 3.7.Comparison of DEC greenness with other solvents

The most commonly used methods of greenness evaluation applied to solvents are EHS (environmental, health and safety) assessment and life-cycle assessment (LCA).<sup>27,28</sup> The first one is a screening method that aims to identify potential hazards of chemicals. The second one is used for an assessment of emissions to the environment as well as resource use over the full life-cycle of a solvent, including the production, the use, potential recycling, and the disposal.

Beginning with LCA, the oldest known method for DEC production is phosgenation of ethanol,<sup>29</sup> however it is undesired way of synthesis, due to toxic substrate. Since phosgene route of organic carbonates has been abandoned, non-phosgene synthesis have been developed. They include among others oxidative carbonylation of ethanol, transestefication of carbonate, alcoholysis of urea, ethanolysis of  $CO_2$  and decarbonylation of diethyl oxalate. Some of them are summarized in Figure 4.<sup>30</sup>And it can be noted that there is a significant growth of publications concerning DEC production via non-phosgene routes since 2009 year. Recently, the most frequently used method of DEC production is synthesis from urea due to its cheap and non-toxic raw materials. Considering potential recycling, and the disposal of diethyl carbonate, it also seems to be relatively non-hazardous. When it is released into environment, it decomposes to benign  $CO_2$  and ethanol.<sup>31</sup>



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Fig. 4. Possible non-phosgene methods for DEC production<sup>30</sup>

Another method of assessment is EHS data interpretation. This information mainly relays on physical and chemical properties, toxicity, environmental and safety aspects of the substances to be assessed. The simplest way of dealing with data is looking through the Material Safety Data Sheets. Solvents that could be considered a green should be characterized by relatively high flash point to prevent potential fire ignition. The boiling point ought to be between 70-139 °C, because low temperatures may generate vapours and high ones may influence negatively on easy solvent recycling and complicates the work-up and downstream unit operations, for instance drying.<sup>6</sup> Also octanol-water partition coefficient should not exceed value log Kow > 3, which denotes high potential of bioaccumulation.<sup>32,33</sup> The best score for biodegradation in 28-days test would be 100% as for example hexane shows, however level of biodegradability of DEC is satisfactory. According to International Agency for Research on Cancer (IARC) diethyl carbonate is not classified as carcinogenic or potentially carcinogenic substance, in contrary to formerly used benzene and chloroform.

The first researches on DEC toxicity were carried out in 1966<sup>34</sup> and then 1978.<sup>35</sup> In the first 277 278 one, 100-week study with rats consuming water containing up to 0.3% DEC was presented. 279 No effect on survival, growth, clinical chemistry, hematology and pathology of the rats was 280 found. Later long-term toxicity of DEC in mice was studied. During 83 weeks drinking water 281 with 0-1000 ppm DEC was given to group of 48 male and 50 female mice. Also no effect on 282 mortality, rate of body-weight gain or the incidence of histopathological findings, including 283 tumors was found. The latest assessment on DEC was conducted in 2018 by Kumar Das et al.<sup>36</sup> and the potential of DEC as a green solvent and the fact that there is not 284 285 much available information about it was emphasized.

286 An attempt to make a comprehensive solvents evaluation in the point of sustainable 287 development (including DEC) for medicine chemistry was also made by GlaxoSmithKline (GSK).<sup>37,38</sup>Solvent selection guide (SSG) is the system to provide concise and useful 288 289 information for scientists and engineers to help with green solvent selection decision 290 problems. The SSG includes the most commonly used solvents within GSK operations. Its 291 approach is based on relative ranking of associated environmental, health and safety issues for 292 each solvent. Solvents assessment is prepared based on collected data including different 293 areas of interest (categories), for instance incineration, recycling, biotreatment, VOC emissions, impact on aquatic and air, health hazard, exposure potential, flammability, 294 reactivity, LCA. More details of their descriptions may be found in SSG.<sup>38</sup> Based on SSG 295

results, diethyl carbonate is classified as a green solvent. Moreover DEC is one of the recommended solvent alternatives. In reference to mostly used solvents, DEC is characterized by good score values within each category.

DEC is less commonly used solvent than traditional extraction solvents such as toluene, chloroform, etc. Further research on its properties and especially toxicity endpoints is needed. However, according to results from studies carried out so far, DEC could be considered as a potentially green solvent. The comparison of DEC with other extraction solvents, presented in table 6 shows that it is less toxic, biodegradable and relatively safe in handling.

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Table 6. Comparison of parameters of DEC with other extraction solvents

Solvent	MW [g/mol]	F.P. [°C]	B.P. [°C]	Log K <sub>OW</sub>	Biodegradability in % (28 day test)	LD50 towards rats administered orally [mg kg <sup>-1</sup> ]	Toxicity towards Daphnia Magna EC50 [mg/L]
Diethyl carbonate 105-58-8	118.13	29	127	1.21	75	4876	103 mg/L/48h
Hexane 110-54-3	86.18	-25	69	4	100	25000	3.8 mg/L/24 h
Chloroform 67-66-3	119.38	Does not flash	60	1.97	0	695	79 mg/L/24 h
Toluene 108-88-3	92.14	4	110.5	2.65	86	> 5000	6 mg/L/48 h
Diethyl ether 60-29-7	74.12	-40	34.6	1.05	7	1215	165 mg/L/24 h

4. Conclusions

307 Diethyl carbonate is greener option to be applied as extraction analytical solvent than 308 commonly used organic solvents. It can be obtained from renewable resources and is safer in 309 use and much less toxic than the most of organic solvents. It is characterized by low toxicity 310 and high biodegradability. DEC was applied as extraction solvent for determination of CPs in 311 water samples by means of DLLME. Methanol was used as dispersive solvent and optimal 312 extraction parameters were selected. Estimated values of LODs and CVs were comparable 313 with other analytical procedures for CPs determination. The results of the study show that diethyl carbonate is promising green, extraction solvent for analytical chemistry. It has a potential to be applied for other extraction purposes and is worth future investigations.

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322 References

<sup>1</sup> S. Armenta, S. Garrigues and M. de la Guardia, *TrAC Trends Anal. Chem.*, 2008, **27**(6), 497-511.

<sup>2</sup> A. Spietelun, Ł. Marcinkowski, M. de la Guardia and J. Namieśnik, *J. Chromatogr. A*, 2013, 1321, 1-13.

<sup>3</sup> L. Kocúrová, I. S. Balogh and V. Andruch, *Microchem. J.*, 2013, **110**, 599-607.

<sup>4</sup> C. Capello, U. Fischer and K. Hungerbühler, *Green Chem.*, 2007, **9**(9), 927-934.

<sup>5</sup> F. P. Byrne, S. Jin, G. Paggiola, T. H. M. Petchey, J. H. Clark, T. J. Farmer, A. J. Hunt, C.
R. McElroy, J. Sherwood, *Sustainable Chem. Processes*, 2016, 4(1), 1-24.

<sup>6</sup> D. Prat, A. Wells, J. Hayler, H. Sneddon, C. R. McElroy, S. Abou-Shehada and P. J. Dunn, *Green Chem.*, 2015, **18**(1), 288-296.

<sup>7</sup> B. Schaffner, F. Schaffner, S. P. Verevkin, A. Borner, *Chem.Rev.*, 2010, **110(8)**, 4554-4581.

<sup>8</sup> N. S. Roh, B. C. Dunn, E. M. Eyring, R. J. Pugmire and H. L. Meuzelaar, *Fuel process. Technol.*, 2003, **83(1-3)**, 27-38.

<sup>9</sup> J. Gu, Y. Gao, X. Xu, J. Wu, L. Yu, Z. Xin and S. Sun, *Fuel*, 2018, **216**, 781-786.

<sup>10</sup> D. Li, W. Fang, Y. Xing, Y. Guo and R. Lin, *J. hazard. Mater.*, 2009, **161(2-3)**, 1193-1201.

<sup>11</sup> F. Mueller, N. Loeffler, G. T. Kim, T. Diemant, R. J. Behm and S. Passerini, *ChemSusChem*, 2016, **9(11)**, 1290-1298.

<sup>12</sup> H. Zheng, B. Li, Y. Fu, T. Abe and Z. Ogumi, *Electrochim. acta*, 2006, **52(4)**, 1556-1562.

<sup>13</sup> S. Udayakumar, A. Pandurangan and P. K. Sinha, *Appl. Catal. A: General*, 2005, **287(1)**, 116-128.

<sup>14</sup> J. Olejniczak, J. Staniewski and J. Szymanowski, *Anal. Chim. Acta*, 2005, **535(1-2)**, 251-257.

<sup>15</sup> P. Viñas, N. Campillo, I. López-García and M. Hernández-Córdoba, *Anal. Bioanal. Chem.*,
2014, 406(8), 2067-2099.

<sup>16</sup> J. Ma, W. Lu and L. Chen, *Curr. Anal. Chem.*, 2012, **8**(1), 78-90.

<sup>17</sup> L. Kocúrová, I. S. Balogh, J. Šandrejová and V. Andruch, *Microchem. J.*, 2012, **102**, 11-17.

<sup>18</sup> Y. Wu, L. Dai, J. Cheng, F. Guo and J. Li, *Chromatographia*, 2010, **72**(7-8), 695-699.

<sup>19</sup> M. Pera-Titus, V. García-Molina, M. A. Baños, J. Giménez and S. Esplugas, *Appl. Catal. B*:, 2004, **47**(**4**), 219-256.

<sup>20</sup> M. Czaplicka, *Sci. Total Environ.*, 2004, **322(1-3)**, 21-39.

<sup>21</sup> P. Bigus, A. Szalkiewicz, J. Namiesnik and M. Tobiszewski, *Curr. Anal. Chem.*, 2017, 13(3), 262-268.

<sup>22</sup> L. Hou, G. Shen, H. K. Lee, J. Chromatogr. A, 2003, **985(1-2)**, 107-116.

<sup>23</sup> C. Basheer, H. K. Lee, J. Chromatogr. A, 2004, **1057**, 163-169

<sup>24</sup> H. Bagheri, A. Saber, S. R. Mousavi, J. Chromatogr. A, 2004, **1046**, 27-33

<sup>25</sup> H. Xu, Y. Liao, J. Yao, J. Chromatogr. A, 2007, **1167**, 1-8

- <sup>26</sup> N. Fattahi, Y. Assadi, M.R.M. Hosseini, E.Z. Jahromi, *J. Chromatogr. A*, 2007, **1157**, 23-29
- <sup>27</sup> G. Koller, U. Fischer, K. Hungerbühler, *Ind. Eng. Chem. Res*, 2000, **39**(**4**), 960-972.

<sup>28</sup> Environmental management - Life cycle assessment – Principles and framework, EN ISO
14040, European Committee for Standardisation, Brussels, Belgium, 1997.

<sup>29</sup> I. E. Muskat and F. Strain, US Pat., 2 379 250, 1941.

<sup>30</sup> K. Shukla and V. C. Srivastava, *RSC Adv.*, 2016, **6(39)**, 32624-32645.

<sup>31</sup> A. A. G. Shaikh and S. Sivaram, *Chem. Rev.*, 1996, **96(3)**, 951-976.

<sup>32</sup> D. J. Miller and P. McWilliams, *Tenside Surfact. Det.*, 2010, **47**(1), 28-33.

<sup>33</sup> A. Verliefde, N. Van Vliet, G. Amy, B. Van der Bruggen, J. C. Van Dijk, *Water Pract. Technol.*, 2006, **1**(**4**)

<sup>34</sup> G. Bornmann and A. Loeser, Archiv für Toxikologie, 1966, **22(2)**, 98-114.

<sup>35</sup> D. Brown, I. F. Gaunt, J. Hardy, I. S. Kiss and K. R. Butterworth, *Toxicology*, 1978, **10**, 291-295.

<sup>36</sup> A. K. Das, B. N. Rajasekhar, S. Krishnakumar, J. Quant. Spectrosc. Radiat. Transfer, 2018, **217**, 53-62.

<sup>37</sup> R. K. Henderson, C. Jiménez-González, D. J. Constable, S. R. Alston, G. G. Inglis, G. Fisher and A. D. Curzons, *Green Chem.*, 2011, 13(4), 854-862.

<sup>38</sup> C. M. Alder, J. D. Hayler, R. K. Henderson, A. M. Redman, L. Shukla, L. E. Shuster, H. F. Sneddon, *Green Chem.*, 2016, **18**(**13**), 3879-3890.