

## Efficient Synthesis of Functionalized Unsymmetrical Dialkyl Trisulfanes

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**Abstract:** We have developed a convenient method for the synthesis of functionalized unsymmetrical dialkyl trisulfanes under mild conditions in very good yields. The designed method is based on the reaction of (5,5-dimethyl-2-thioxo-1,3,2-dioxaphosphorinan-2-yl)-disulfanyl derivatives **1** with alkyl disulfanyl anions generated *in situ* from *S*-acetyl disulfanyl derivatives **2** and sodium methoxide. The developed method allows for the preparation of unsymmetrical trisulfanes bearing additional hydroxyl, carboxyl or amino functionalities on both sides of the trisulfane functionality.

**Key words:** unsymmetrical trisulfanes, thiols, hydrodisulfanes, *S*-acetyl alkyl disulfanes, phosphorodithioic acid.

Our interest in the chemistry of diorganyl trisulfanes (trisulfides) arises from their diverse roles in living organisms and their occurrence in natural sources.<sup>1,2</sup> In the literature, these compounds are often termed as organic trisulfides, but the IUPAC- recommended nomenclature is trisulfanes.<sup>3</sup> The name trisulfide should only be applied to ionic compounds, such as Na<sub>2</sub>S<sub>3</sub>. Organic trisulfanes have been isolated from shiitake mushrooms,<sup>4</sup> oil made from *Ferula asafetida*,<sup>5</sup> durian fruit,<sup>6</sup> garlic oil<sup>7,8</sup> and Hawaiian algae.<sup>9</sup> It also should be mentioned that dialanyltrisulfane has been detected in acidic wool hydrolysates, but it is not clear whether the related amino acid (HOOC-CH(NH<sub>2</sub>)-CH<sub>2</sub>)S<sub>3</sub> is a part of the wool structure or is formed from cysteine during hydrolysis.<sup>10</sup> In the biochemical literature, dialanyltrisulfane is often incorrectly termed as “cysteine trisulfide”. A natural peptide containing a trisulfane group in place of a disulfane bridge has been isolated from genetically engineered *Escherichia coli* bacteria.<sup>11</sup> It is a derivative of the human growth hormone consisting of 191 amino acids in a single chain with a trisulfane bridge between cysteine (alanyl) residues 182 and 189.<sup>12</sup> A trisulfanyl functionality has also been observed in recombinant DNA-derived methionyl human growth hormone in the bridge between cysteine residues 53 and 165.<sup>13</sup> Calicheamicin<sup>14,15</sup> and esperamicins A<sub>1</sub> and A<sub>2</sub>,<sup>16,17</sup> members of the enediyne class of antibiotics, also contain the trisulfanyl functionality. These natural products are very potent antitumor antibiotics.

There are numerous reactions that can be used to prepare symmetrical organic trisulfanes. The most common methods include the reaction of thiols with sulfur dichloride,<sup>18</sup> the coupling of alkyl halides with sodium trisulfide,<sup>19</sup> and the reaction of thiols or disulfanes with sulfur.<sup>20</sup> Thioalkylation reactions of various thiosulfenate species can also produce trisulfanes. The most suitable substrates include Bunte

salts,<sup>21</sup> metal sulfides,<sup>22</sup> and thiosulfenyl chloride.<sup>23</sup> The latter can also be used for the preparation of unsymmetrical trisulfanes. Other practical procedures involve the reduction of thiosulfonates and disulfanyl sulfides with phosphines,<sup>24</sup> sulfur insertion reactions into thiosulfonates, thiosulfonates,<sup>25</sup> and disulfanes,<sup>26</sup> alkoxide decomposition of sulfenylthiocarbonates,<sup>27</sup> and reactions of thiols with 1,1-thiobis(benzimidazole)<sup>28</sup> or diimidazolylsulfide.<sup>29</sup>

Preparative methods that are efficient for the preparation of symmetrical trisulfanes are very often ineffective for the preparation of unsymmetrical trisulfanes. Indeed, the synthesis of unsymmetrical trisulfanes is more complex. There are known procedures based on the coupling of chlorodisulfanes with *N*-arylamidithiosulfites<sup>30</sup> or thiols<sup>31,32</sup> or the sequential coupling of two thiols using sulfur dichloride.<sup>33</sup> Other procedures involve the desulfurization of unsymmetrical dialkanesulfonic thioanhydrides,<sup>24</sup> or the use of (often) unstable hydrodisulfanes.<sup>34</sup>

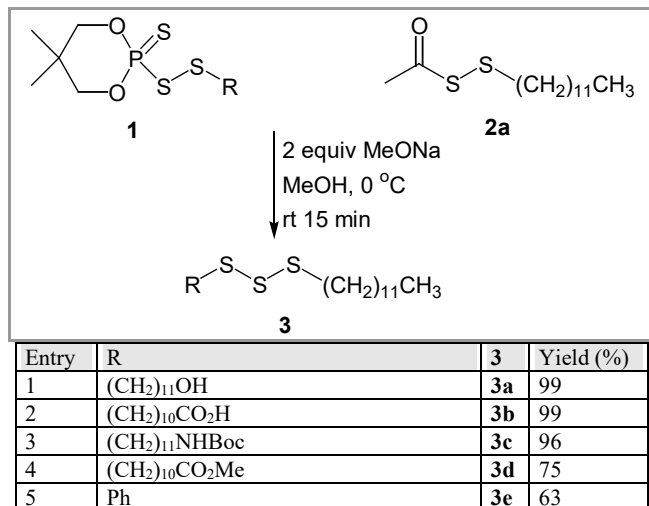
Moderate yields and/or the formation of undesirable polysulfane side products are the major drawbacks of the methods presented above. The likelihood that pure trisulfanes can be separated from the reaction mixture is very poor. The best purification method is crystallization, but clearly this can only be applied to solid products. Moreover, the scopes of the presented methods are limited by the availability of reagents and the chemical reactivity of the additional functional groups.

We have previously demonstrated the preparation of functionalized unsymmetrical molecules such as dialkyl disulfanes, alkyl-aryl disulfanes,<sup>35</sup> ‘bioresistant’ disulfanes,<sup>36</sup> unsymmetrical disulfanes of L-cysteine and L-cystine,<sup>37</sup> and diaryl disulfanes<sup>38</sup> based on the readily available 5,5-dimethyl-2-thioxo-1,3,2-dioxaphosphorinane-2-disulfanyl derivatives **1**. These disulfanyl derivatives **1** of phosphorodithioic acid were also convenient for the preparation of  $\alpha$ -sulfenylated carbonyl compounds<sup>39</sup> and symmetrical trisulfanes.<sup>40</sup>

The limitations of our previous method for the preparation of unsymmetrical trisulfanes<sup>41</sup> have encouraged us to develop a new synthetic strategy for the preparation of unsymmetrical trisulfanes bearing additional functionalities on either side of the trisulfane. The idea is based on the reaction of electrophilic disulfanyl derivatives of phosphorodithioic acid **1** with a nucleophilic

dodecyldisulfanyl anion generated *in situ* from *S*-acetyl dodecyldisulfane **2a** and sodium methoxide (Table 1).

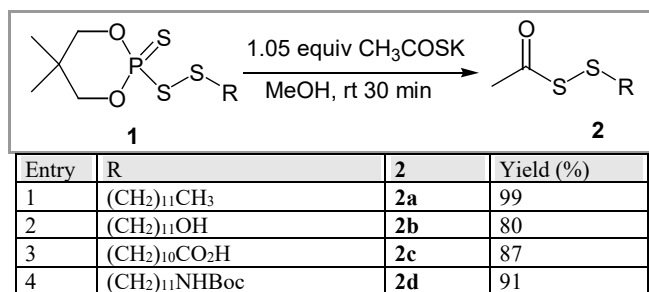
Table 1 Formation of Unsymmetrical Trisulfanes **3** from *S*-Acetyl Dodecyldisulfane **2a**



It is known that the preparation of polysulfanes is frequently hampered by the formation of their homologues with a variable number of sulfur atoms.<sup>32</sup> The purification of such a mixture is, at best, difficult and, in most cases, impossible due to the similar properties of the products. We were able to avoid the formation of side products by using a small excess of the disulfanyl derivatives **1** (1.05 equivalent). In this case, the potentially unstable dodecyldisulfanyl anion was consumed very quickly and sulfur extrusion or exchange with product **3** was not observed.

The *S*-acetyl alkyldisulfanes **2** are readily available from disulfanyl derivatives **1** and potassium thioacetate (Table 2).

Table 2 Preparation of *S*-Acetyl Alkyldisulfanes **2**



We selected disulfanyl derivatives **1** and *S*-acetyl alkyldisulfanes **2** bearing hydroxy, carboxy, methyl ester, tert-butoxycarbonylamino, and phenyl functionalities to examine the scope and limitations of this method for the preparation of functionalized unsymmetrical trisulfanes **3**. All of the above alkyl

derivatives afforded, after purification, unsymmetrical trisulfanes **3** in very good yield (Table 3).

As shown in Table 3, the same unsymmetrical trisulfane **3** can be obtained in two different ways. For example, **3b** can be prepared from **1c** and **2a** or from **1a** and **2b**. Both approaches gave product **3b** in very good yield (Table 3). However, the reaction of disulfanyl derivative **1f** (R<sup>1</sup> = Ph) with *S*-acetyl alkyldisulfanes **2** was successful only for compound **2a**. Unfortunately, the other *S*-acetyl alkyldisulfanes **2b-d** provided products **3i**, **3l** and **3n**, that were contaminated with symmetrical and unsymmetrical di- and tetrasulfanes, respectively. Although di-, tri- and tetrasulfanes could not be separated by column chromatography, the presence of these impurities can be easily confirmed by <sup>1</sup>H NMR. The chemical shift of the methylene group (triplet in CDCl<sub>3</sub>) adjacent to the sulfur for di-, tri- and tetrasulfanes are 2.66, 2.87 and 3.05, respectively. The versatility of the method is limited by very fast sulfur extrusions and exchange reactions in the case of aryl alkyl trisulfanes **3i**, **3l**, and **3n**. It seems that aromatic groups can promote these side reactions because the arylthiolate anion is a very good leaving group.

In conclusion, a convenient method for the preparation of unsymmetrical dialkyl trisulfanes **3**, bearing hydroxy, carboxy, methyl ester, or tert-butoxycarbonylamino groups on either or both sides of the trisulfane has been developed. Reactions of **1** with a variety of **2** in the presence of sodium methoxide in methanol at room temperature were generally complete within 30 minutes and gave exclusively unsymmetrical dialkyl trisulfanes **3** in very good yields after purification. Because the reactions of the *S*-acetyl alkyldisulfanes **2** proceeded with a small excess of **1** under mild reaction conditions and in a short time, thiol-trisulfane exchange and sulfur extrusion did not occur during the reaction. The simplicity and very good yields of this method make it one of the most attractive approaches for the preparation of functionalized unsymmetrical dialkyl trisulfanes.

### Typical Procedure for Trisulfane Preparation

A solution of MeONa (2.0 mmol) in dry MeOH (2 mL) was added to a solution of **1** (1.05 mmol) and **2** (1.0 mmol) in dry MeOH (20 mL) at 0 °C under a N<sub>2</sub> atmosphere. Then, the ice bath was removed, and the mixture was stirred for 30 min at r.t. After evaporation of the solvent under reduced pressure, the residue was purified by column chromatography. The addition of small amount of AcOH prior to purification on a silica gel column inhibited sulfur extrusion during separation. The yields are reported in Table 3.

Table 3 Synthesis of Unsymmetrical Trisulfanes 3<sup>a</sup>

R <sup>1</sup>	(CH <sub>2</sub> ) <sub>11</sub> CH <sub>3</sub> <b>1a</b>	(CH <sub>2</sub> ) <sub>11</sub> OH <b>1b</b>	(CH <sub>2</sub> ) <sub>10</sub> CO <sub>2</sub> H <b>1c</b>	(CH <sub>2</sub> ) <sub>11</sub> NHBoc <b>1d</b>	(CH <sub>2</sub> ) <sub>10</sub> CO <sub>2</sub> Me <b>1e</b>	Ph <b>1f</b>
R <sup>2</sup>						
(CH <sub>2</sub> ) <sub>11</sub> CH <sub>3</sub> <b>2a</b>	-	<b>3a</b> (99)	<b>3b</b> (99)	<b>3c</b> (96)	<b>3d</b> (75)	<b>3e</b> (63)
(CH <sub>2</sub> ) <sub>11</sub> OH <b>2b</b>	<b>3a</b> (71)	-	<b>3f</b> (72)	<b>3g</b> (78)	<b>3h</b> (60)	<b>3i</b> (0)
(CH <sub>2</sub> ) <sub>10</sub> CO <sub>2</sub> H <b>2c</b>	<b>3b</b> (85)	<b>3f</b> (70)	-	<b>3j</b> (73)	<b>3k</b> (62)	<b>3l</b> (0)
(CH <sub>2</sub> ) <sub>11</sub> NHBoc <b>2d</b>	<b>3c</b> (87)	<b>3g</b> (76)	<b>3j</b> (74)	-	<b>3m</b> (77)	<b>3n</b> (0)

<sup>a</sup> The yields (%) of the isolated product are reported in parentheses

### Representative Analytical Data

#### 11-(11-Hydroxyundec-1-yl-trisulfanyl)undecanoic acid (**3f**)

Chromatography (CH<sub>2</sub>Cl<sub>2</sub> : EtOAc, 10:1); yield: 0.326 g (72%); white solid; mp: 49–52 °C. IR (KBr):  $\nu$  = 3345 (w), (OH), 2918 (s), 2848 (m), (CH), 1710, (m), (C=O), 1472 (m), (S-CH<sub>2</sub>), 1036 (m), (O-CH<sub>2</sub>), 716 (m), (CH<sub>2</sub>) cm<sup>-1</sup>. <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.25–1.48 (m, 28H, CH<sub>2</sub>), 1.53–1.77 (m, 6H, CH<sub>2</sub>), 2.34 (t,  $J$  = 7.3 Hz, 2H, C(O)CH<sub>2</sub>), 2.87 (t,  $J$  = 7.3 Hz, 4H, S-CH<sub>2</sub>), 3.18 (bs, 2H, OH, COOH), 3.64 (t,  $J$  = 6.5 Hz, 2H, O-CH<sub>2</sub>). <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>):  $\delta$  = 178.5, 62.9, 38.9, 33.9, 32.7, 29.7, 29.6, 29.5, 29.4, 29.3, 29.1, 29.0, 28.9, 28.8, 28.6, 28.5, 28.2, 25.7, 24.7. Signals: expected, 22; observed, 19. ESI-HRMS:  $m/z$  [M + Na]<sup>+</sup> calcd for C<sub>22</sub>H<sub>44</sub>NaO<sub>3</sub>S<sub>3</sub>: 475.2350; found: 475.2344.

#### *N*-(*tert*-Butoxycarbonyl)-11-(11-hydroxyundec-1-yl-trisulfanyl)-1-undecylamine (**3g**)

Chromatography (CH<sub>2</sub>Cl<sub>2</sub> : EtOAc, 10:1); yield: 0.420 g (78%); colorless oil. IR (KBr):  $\nu$  = 3381 (m), (NH), 3345 (w), (OH), 2920 (s), 2850 (s), (CH), 1685, (s), (C=O), 1637 (m), (NH), 1170 (m), (C-N), 720 (w), (CH<sub>2</sub>) cm<sup>-1</sup>. <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.20–1.45 (m, 33H, CH<sub>2</sub>, OH), 1.46 (s, 9H, *t*Bu), 1.65–1.77 (m, 4H, S-C-CH<sub>2</sub>), 2.88 (t,  $J$  = 7.3 Hz, 4H, S-CH<sub>2</sub>), 3.00–3.20 (m, 2H, N-CH<sub>2</sub>), 3.64 (t,  $J$  = 6.5 Hz, 2H, O-CH<sub>2</sub>), 4.48 (bs, 1H, NH). <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>):  $\delta$  = 155.8, 78.8, 62.9, 40.6, 39.1, 38.8, 31.8, 30.2, 30.1, 29.7, 29.5, 29.4, 29.3, 29.2, 29.1, 29.0, 28.8, 28.5, 28.4, 26.8, 22.7. Signals: expected, 25; observed, 21. ESI-HRMS:  $m/z$  [M + Na]<sup>+</sup> calcd for C<sub>27</sub>H<sub>55</sub>NNaO<sub>3</sub>S<sub>3</sub>: 560.3242; found: 560.3245.

#### 11-(11-Hydroxyundec-1-yl-trisulfanyl)undecanoic acid methyl ester (**3h**)

Chromatography (petroleum ether : CH<sub>2</sub>Cl<sub>2</sub>, 2:1); yield: 0.280 g (60%); yellow solid; mp: 26–28 °C. IR (KBr):  $\nu$  = 3378 (w), (OH), 2922 (s), 2852 (m), (CH), 1743, (m), (C=O), 1472 (m), (S-CH<sub>2</sub>), 1036 (m), (O-CH<sub>2</sub>), 720 (m), (CH<sub>2</sub>) cm<sup>-1</sup>. <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.02–1.77 (m, 35H, CH<sub>2</sub>, OH), 2.22–2.34 (m, 2H, CH<sub>2</sub>), 2.87 (t,  $J$  = 7.3 Hz, 4H, S-CH<sub>2</sub>), 3.64 (t,  $J$  = 6.5 Hz, 2H, O-CH<sub>2</sub>), 3.67 (s, 3H, COOCH<sub>3</sub>). <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>):  $\delta$  = 174.7, 63.0, 51.4, 39.4,

39.2, 38.8, 34.4, 34.0, 32.7, 29.7, 29.5, 29.4, 29.3, 29.2, 29.1, 29.0, 28.8, 28.6, 28.5, 28.4, 25.7, 24.9. Signals: expected 23, observed 22. ESI-HRMS:  $m/z$  [M + H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>47</sub>O<sub>3</sub>S<sub>3</sub>: 467.2687; found: 467.2691.

#### 11-[11-(*N*-*tert*-Butoxycarbonylamino)undec-1-yl-trisulfanyl]undecanoic acid (**3j**)

Chromatography (CH<sub>2</sub>Cl<sub>2</sub> : EtOAc, 10:1); yield: 0.403 g (73%); white solid; mp: 54–56 °C. IR (KBr):  $\nu$  = 3381 (m), (NH), 2920 (s), 2850 (s), (CH), 1895, (m), (C=O), 1684, (s), (C=O), 1639 (m), (NH), 1172 (m), (C-N), 730 (w), (CH<sub>2</sub>) cm<sup>-1</sup>. <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.20–1.44 (m, 30H, CH<sub>2</sub>), 1.46 (s, 9H, *t*Bu), 1.66–1.76 (m, 4H, S-C-CH<sub>2</sub>), 2.36 (t,  $J$  = 7.3 Hz, 2H, CH<sub>2</sub>COO), 2.88 (t,  $J$  = 7.3 Hz, 4H, S-CH<sub>2</sub>), 3.00–3.20 (m, 2H, N-CH<sub>2</sub>), 4.48 (bs, 1H, NH). <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>):  $\delta$  = 179.9, 156.0, 78.9, 40.7, 39.2, 38.9, 34.0, 31.9, 29.6, 29.6, 29.5, 29.3, 29.2, 29.1, 29.0, 28.8, 28.7, 28.5, 24.6, 22.7. Signals: expected, 25; observed, 20. ESI-HRMS:  $m/z$  [M + H]<sup>+</sup> calcd for C<sub>27</sub>H<sub>54</sub>NO<sub>4</sub>S<sub>3</sub>: 552.3215; found: 552.3220.

**Supporting Information** for this article consisting of experimental details and spectroscopic data for compounds **1**, **2** and **3** is available online at <http://www.thieme-connect.com/ejournals/toc/synlett>.

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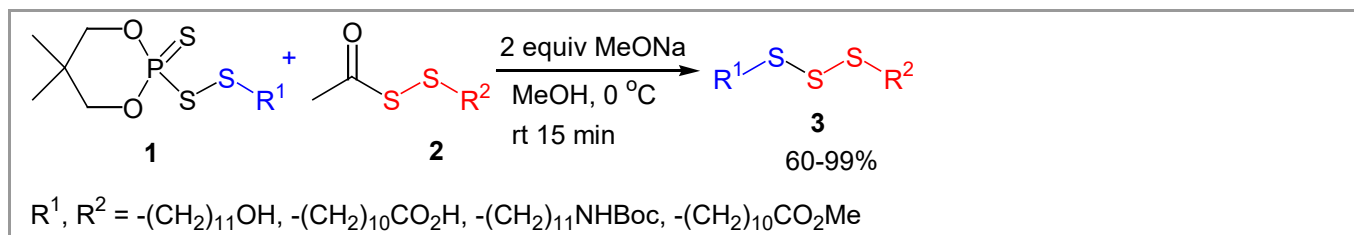
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