



Phase I and phase II metabolism simulation of antitumor-active 2-hydroxyacridinone with electrochemistry coupled on-line with mass spectrometry.

Journal:	<i>Xenobiotica</i>
Manuscript ID	TXEN-2018-0223.R1
Manuscript Type:	Original Article
Keywords:	<i>in vitro</i> metabolism, metabolic activation, electrochemical oxidation, electrochemistry-mass spectrometry, metabolite electrosynthesis, reactive metabolite, glutathione S-conjugate

SCHOLARONE™
Manuscripts

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Title:

Phase I and phase II metabolism simulation of antitumor-active 2-hydroxyacridinone with electrochemistry coupled on-line with mass spectrometry.

For Peer Review Only

Abstract

1. Here, we report the metabolic profile and the results of associated metabolic studies of 2-hydroxyacridinone (2-OH-AC), the reference compound for antitumor-active imidazo- and triazoloacridinones.
2. Electrochemistry coupled with mass spectrometry was applied to simulate the general oxidative metabolism of 2-OH-AC for the first time. The reactivity of 2-OH-AC products to biomolecules was also examined. The usefulness of the electrochemistry for studying the reactive drug metabolite trapping (conjugation reactions) was evaluated by the comparison with conventional electrochemical (controlled-potential electrolysis) and enzymatic (microsomal incubation) approaches.
3. 2-OH-AC oxidation products were generated in an electrochemical thin-layer cell. Their tentative structures were assigned based on tandem mass spectrometry in combination with accurate mass measurements. Moreover, the electrochemical conversion of 2-OH-AC in the presence of reduced glutathione and/or *N*-acetylcysteine unveiled the formation of reactive metabolite-nucleophilic trapping agent conjugates (m/z 517 and m/z 373, respectively) via the thiol group. This glutathione S-conjugate was also identified after electrolysis experiment as well as was detected in liver microsomes.
4. Summing up, the present work illustrates that the electrochemical simulation of metabolic reactions successfully supports the results of classical electrochemical and enzymatic studies. Therefore, it can be a useful tool for synthesis of drug metabolites, including reactive metabolites.

Keywords:

in vitro metabolism; metabolic activation; electrochemical oxidation; electrochemistry-mass spectrometry; metabolite electrosynthesis; reactive metabolite; glutathione S-conjugate;

For Peer Review Only

Introduction

Solid understanding of the metabolic pathways and the biotransformation mechanisms of new drug candidates is a crucial point in the drug discovery and development processes. Overall, it allows to elucidate the **metabolic** activation as well as the deactivation routes of new biologically active compounds, especially in respect to their possible toxicity (Park *et al.* 2011). The identification of metabolites helps to eliminate the inappropriate candidates at an early stage, before the more expensive development phases will be performed (Bussy & Boujtita 2014). Particularly, it is very important that the formation of chemically reactive intermediates is checked (Baillie *et al.* 2002), because an increasing number of reports indicate that they are responsible for the majority of rapid and unexpected drug toxic effects (Kalgutkar & Soglia 2005; Srivastava *et al.* 2010; Orhan & Vermeulen 2011). The relationship between drug metabolism and adverse drug reactions was first demonstrated with the analgesic agent acetaminophen (Jollow *et al.* 1973; Larson 2007). Reactive intermediates, generated usually via cytochrome P450 (P450)-catalyzed oxidative reactions, have the potential for covalent binding to cellular nucleophiles such as purine and pyrimidine bases of DNA or thiols of proteins, and form stable adducts. Adduct formation may alter biological functions of these biomolecules what ultimately leads to a toxic response (Brandon *et al.* 2003).

Metabolic activation of new drugs to reactive intermediates is currently assessed in the presence or the absence of reactive endogenous nucleophiles, such as reduced glutathione (GSH), *N*-acetylcysteine (NAC) or β -lactoglobulin A (β LGA), a model protein. In practise, the reaction usually involves the *in vitro* incubation of a studied compound with an excess of the selected chemical-trapping agent in hepatocytes or liver microsomes (as a source of cytochrome P450 isoenzymes). *In vivo* experiments involve laboratory animals (Kalgutkar & Soglia 2005). However, performing these biological schemes is usually laborious and time-consuming. Despite continuous improvements in metabolite-identification tools, the identification of some reactive metabolites remains difficult due to the matrix complexity, low concentration or their binding to matrix bio-components (Bussy *et al.* 2015). Isolated

1
2
3 hepatocytes have only a very limited proliferative potential *in vitro* and a correspondingly
4 short life-span in primary culture. Their fenotype is unstable and the level and activities of
5 enzymes, such as cytochromes P450, fall quickly in a few days (Jennings & Strauss 1999).
6
7 Also liver microsomes offer a limited reproducibility. Additionally, because of genetic
8 polymorphisms, variations in the gene expression for individual drug-metabolising
9 isoenzymes in each organism have to be taken into account (Pinto & Dolan 2011; Zanger &
10 Schwab 2013).
11
12

13
14
15
16 As an alternative to the existing methods for studies on drug metabolism and toxicity, the
17 electrochemical simulation of P450-catalyzed phase I reactions, mostly initiated by a single-
18 electron transfer, has been developed (Volk *et al.* 1992; Lohmann & Karst 2008; Faber *et al.*
19 2011). Overall, it allows to simulate a wide variety of oxidation-reduction (redox) processes
20 occurring in living organisms (Jurva *et al.* 2003; Nouri-Nigjeh *et al.* 2011). The combination of
21 electrochemistry (EC) coupled on-line with mass spectrometry (MS) creates a powerful
22 platform for rapid generation (in the electrochemical cell) and detection (by mass
23 spectrometry) of a series of metabolic products, including observation of reactive metabolites
24 (Permentier *et al.* 2008; Faber *et al.* 2011; Bussy & Boujtita 2014). Use of EC/MS improves
25 the conventional methods of drug metabolism studies. It is a purely instrumental technique
26 with a simple set-up that enables the generation of drug metabolites in the absence of
27 biological matrices in the reaction medium. So, the application of EC/MS helps to overcome
28 many of the laborious tasks related to isolation and identification of metabolic products
29 formed *in vitro* (cultured hepatocytes, liver microsomes, purified enzymes) or *in vivo* (urine,
30 plasma, *etc.*) (Orhan & Vermeulen 2011; Faber *et al.* 2011). Moreover, on-line combining EC
31 with MS can provide valuable information about metabolically labile sites in a drug molecule
32 and predict its reliable metabolic profile in a much shorter time (Jurva *et al.* 2003; Permentier
33 *et al.* 2008).
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51

52 Recently extensive research have been conducted in our group to determine the possible
53 metabolic pathways of potential antitumor drugs and their model compounds. The objective
54 of the investigations presented here was to develop and evaluate an on-line EC/MS method
55
56
57
58
59
60

1
2
3 for the simulation of oxidative metabolism of 2-hydroxyacridinone (2-OH-AC) (a structure in
4 the frame in Figure 1), a simple reference compound for high antitumor-active imidazo- and
5 triazoloacridinones.
6
7

8
9 The preliminary studies in this field were performed earlier by the application of cyclic
10 voltammetry, spectroelectrochemical measurements, and controlled-potential electrolysis
11 (Mazerska *et al.* 1997, 2002). These experiments indicated the ability of 2-OH-AC to undergo
12 oxidative metabolic activation in the living organism. However, no data for acridinone
13 derivatives are yet available using the direct coupling EC with MS. The studies presented
14 here were undertaken with respect to the usefulness of EC/MS technique for further
15 investigation on the oxidative metabolic activation and the identification of potential reactive
16 metabolites formed during the metabolism of antitumor-active imidazoacridinone C-1311 and
17 triazoloacridinone C-1305. These compounds, developed in our laboratory, have
18 demonstrated significant cytotoxic and antitumor activities (Cholody *et al.* 1990, 1996;
19 Kusnierczyk *et al.* 1994). The data obtained so far indicate that they have different spectrum
20 of antitumor activity and exhibit various mechanisms of action at the molecular level
21 (Dziegielewski & Konopa 1996; Skladanowski *et al.* 1996; Dziegielewski *et al.* 2002; Lemke
22 *et al.* 2005; Augustin *et al.* 2006). Metabolic activation is the leading concept for the reason
23 how imidazo- and triazoloacridinone derivatives cause high antitumor effect (Dziegielewski &
24 Konopa 1998; Mazerska *et al.* 2001; Koba & Konopa 2007). It is assumed that the hydroxyl
25 group in the acridinone ring provides the susceptibility of these compounds to oxidative
26 metabolism. The formation of the intermediate, characterized as a quinone imine structure,
27 that expresses antitumor activity, has been shown, for example, in the case of 9-
28 hydroxyellipticine (Harding & Grummitt 2003). A known quinone imine with activity against
29 human tumor cells is actinomycin D (Marks & Venditti 1976).
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49

50 In this work, the ability of EC/MS to expedite the generation and identification of the main
51 phase I metabolites of 2-OH-AC will be discussed. The formation of the reactive 2-OH-AC
52 intermediate metabolite and the possibility to simulate its covalent binding to biomolecule
53 (*i.e.*, glutathione (GSH) and/or *N*-acetylcysteine (NAC) as biomarkers of metabolic activity;
54
55
56
57
58
59
60

1
2
3 phase II metabolism) are also taken into account. Furthermore, to show the capability of
4 electrochemistry to simulate certain P450-mediated reactions, the results obtained by EC
5 were also compared with those gained after controlled-potential electrolysis (CPE) and
6 conventional *in vitro* studies by conducting incubations of 2-OH-AC with human and rat liver
7 microsomes (HLMs and RLMs, respectively). The products of electrolysis and enzymatic
8 transformations of 2-OH-AC were analysed by reversed-phase liquid chromatography (LC)
9 with UV-Vis detection and/or diode array detection, and monitored by MS. The relation
10 between the products generated electrochemically and enzymatically for the model
11 compound 2-OH-AC can provide a clue to the nature of their metabolic pathway initiation
12 (Volk *et al.* 1992). It opens up further perspectives directed to the search for more effective
13 and less toxic antitumor drugs among acridinone derivatives. It is significant because
14 improves the risk evaluation for potential drugs in the optimal chemotherapy schedules
15 designed for individual patients.
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

List of abbreviations:

CPE, controlled-potential electrolysis;

CV, cyclic voltammetry, cyclic voltammogram;

EC, electrochemistry, electrochemical cell;

ESI, electrospray ionization;

GC, glassy carbon;

GSH, glutathione (reduced form);

HLMs, human liver microsomes;

2-OH-AC, 2-hydroxyacridinone;

m/z, mass-to-charge ratio;

MS, mass spectrometry, mass spectrometer;

MS/MS, tandem mass spectrometry;

NADPH, β -nicotinamide adenine dinucleotide 2'-phosphate tetrasodium salt (reduced form);

NAC, *N*-acetylcysteine;

P450, cytochrome P450;

PBS, phosphate-buffered saline;

Q-TOF, quadrupole-time of flight;

RLMs, rat liver microsomes;

LC, liquid chromatography;

Parts of this work were presented at the 13th European ISSX Meeting (Glasgow, Scotland, 2015) and at the 2nd Congress BIO 2016 (Wrocław, Poland, 2016).

Materials and methods

Chemicals and enzymes

An acridinone derivative, a 2-hydroxyacridinone (2-OH-AC) was synthesized in our laboratory according to the method described earlier (Acheson 1973). The following chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA): *N*-acetylcysteine (NAC), dipotassium phosphate (K_2HPO_4), disodium phosphate (Na_2HPO_4), formic acid (HCOOH), L-glutathione reduced (GSH), monopotassium phosphate (KH_2PO_4), monosodium phosphate (NaH_2PO_4), and semicarbazide hydrochloride. Methanol (gradient grade for liquid chromatography) and β -nicotinamide adenine dinucleotide 2'-phosphate tetrasodium salt (β -NADPH) were obtained from Merck KGaA (Darmstadt, Germany). Ammonium formate ($HCOONH_4$) was ordered from Fisher Scientific (Loughborough, UK). Aluminum oxide (Al_2O_3) powder in form of 1- μ m alumina suspension, for polishing of electrodes, was delivered by TESTING Sp z o.o. (Katowice, Poland). All other commercially available chemicals and reagents were of the highest possible grade available. Ultrapure water ($0.056 \mu S \cdot cm^{-1}$), used in all the experiments, was passed through a Milli-Q water purification system from Merck KGaA (Darmstadt, Germany) or water distillation system from Hydrolab Sp. z o.o. sp.k. (Straszyn, Poland).

Pooled human liver microsomes (HLMs), mixed gender, from 50 donors (protein concentration, $20 \text{ mg} \cdot \text{mL}^{-1}$; P450 content, $411 \text{ pmol} \cdot \text{mg}^{-1}$ protein) and pooled rat liver microsomes (RLMs) from untreated, male Sprague-Dawley rats (protein concentration, $20 \text{ mg} \cdot \text{mL}^{-1}$; P450 content, $680 \text{ pmol} \cdot \text{mg}^{-1}$ protein) were purchased from Tebu-bio (Le Perray-En-Yvelines, France).

General instrumentation

Electrochemical measurements

Electrochemical measurements of the oxidation-reduction properties of 2-OH-AC were performed with controlled-potential electrolysis (CPE) and by electrochemical metabolism simulation in a three-electrode thin-layer cell (EC).

1
2
3 CPE was performed using an Autolab (Eco Chemie B.V., Utrecht, The Netherlands),
4 model PGSTAT 12 potentiostat, controlled via producer's software. The three-electrode
5 system, consisting of a cylinder glassy carbon (GC) working electrode ($\phi = 3$ mm and 13 mm
6 long; $A = 1.296$ cm²), an Ag/AgCl/3 M KCl reference electrode and a platinum-wire counter
7 electrode, were employed. During chronoamperometric measurements the working electrode
8 as well as the analyzed solution were placed in a glass tube closed with a "Vicor" plug to
9 separate the electrolyzed solution from the main solution.
10
11

12
13
14
15
16
17 Simulation of the oxidative metabolism reactions of 2-OH-AC was accomplished in an
18 electrochemical thin-layer cell equipped with a disc glassy carbon (GC) working electrode (ϕ
19 = 8 mm; $A = 0.502$ cm²) and a Pd/H₂ reference electrode (reactor cell; Antec Leyden,
20 Zoeterwoude, The Netherlands). Carbon-loaded PTFE (polytetrafluoroethylene) served as
21 auxiliary electrode. The cell potentials were applied using a ROXY EC System (Antec
22 Leyden) controlled by Antec Dialogue software (Antec Leyden). The outlet of the
23 electrochemical cell was connected directly to an electrospray ionization (ESI) source of
24 a quadrupole-time of flight (Q-TOF) mass spectrometer (Agilent Technologies, Santa Clara,
25 CA, USA) (Figure 1a). S-Conjugate formation of 2-OH-AC and GSH (NAC) was established
26 using a T-piece and a 100 μ L mixing coil placed between reactor cell and the ESI source
27 (Figure 1b). The relevant EC conditions can be viewed in Table 1.
28
29
30
31
32
33
34
35
36
37

38 **Liquid chromatographic analyses**

39
40 For all analyses of 2-OH-AC, liquid chromatographic (LC) separations were performed on
41 a reversed-phase 5- μ m Suplex pKb-100 analytical column (4.6 mm x 250 mm, C18)
42 (Supelco Inc., Bellefonte, PA, USA) with Waters Associates HPLC system (Waters Co.,
43 Milford, MA, USA). It was equipped with a model 600E system controller, a model 7725i
44 Rheodyne injector, and a model 2996 photodiode array detector (DAD) controlled with
45 Millennium software (Waters Co.). The LC analyses were carried out at a flow rate of 1
46 mL·min⁻¹ with the following mobile phase system: a linear gradient from 15% to 80%
47 methanol in 0.05 M aqueous ammonium formate buffer (pH 3.40 adjusted with formic acid)
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 for 25 min, followed by a linear gradient from 80% to 100% methanol in ammonium formate
4 buffer, pH 3.40, for 3 min. The eluates were monitored at 380 nm.

6 **Mass spectrometry**

7
8 Mass spectrometric detection, identification, and fragmentation of 2-OH-AC products
9 were carried out in the positive-ion mode by recording full scan spectra (m/z 100 – 600). To
10 ensure accurate mass during the experiment, the mass spectrometer was calibrated on a
11 daily basis, prior to sample analysis. The corresponding conditions for the ESI-Q-TOF-
12 MS(/MS) measurements are listed in Table 1.

18 **Controlled-potential electrolysis of 2-OH-AC**

19
20 The controlled-potential electrolysis was performed in 0.02 M phosphate-buffered saline
21 (PBS), pH 7.40, obtained by mixing Na_2HPO_4 and KH_2PO_4 , and adding the appropriate
22 amounts of NaCl and KCl to reach their concentrations of 0.15 and 0.002 M, respectively.
23 Each time before the electrolysis experiments the GC working electrode was briefly polished
24 with 1- μm Al_2O_3 powder on a wet pad. After polishing step, to remove alumina completely
25 from the surface, the electrode was rinsed with a direct stream of ultrapure water. To
26 eliminate the electric noise the electrochemical cell was placed in a Faraday cage. The
27 chronoamperometric experiments of 0.5 mM 2-OH-AC in the absence and the presence of
28 10-fold excess of GSH were performed at 0.50 V. The electrolysis progress was monitored
29 by voltammetric, spectroscopic, chromatographic, and spectrometric methods. The
30 experiments have been performed at room temperature (21 °C).

42 **Electrochemical simulation of the oxidative metabolism of 2-OH-AC**

43
44 For the electrochemical conversion, a 10- μM 2-OH-AC solution in electrolyte (0.1%
45 formic acid in water/methanol, 50/50, v/v) was pumped (SP2 – ROXY dual piston syringe
46 pump; Antec Leyden) through the electrochemical cell at a constant flow rate of 30 $\mu\text{L}\cdot\text{min}^{-1}$.
47 The potential of working electrode in the reactor cell was ramped between 0 and 2500 mV
48 (scan rate 10 $\text{mV}\cdot\text{s}^{-1}$). The outlet of the reactor cell was connected on-line to the ESI-MS
49 source (Figure 1a). To identify potential aldehyde products the oxidized sample was
50 collected in a vial containing semicarbazide (dissolved in ultrapure water) to give a final
51
52
53
54
55
56
57
58
59
60

1
2
3 concentration of 5 mM. After collection, the samples from the trapping experiments were kept
4
5 at 25 °C for 1 h before analysis by LC/MS.

6
7 The properties of the sample solvent may influence the conversion efficiency in the
8
9 electrochemical cell. Therefore, the electrochemical simulation of the oxidative metabolism of
10
11 2-OH-AC has been also performed in two different electrolyte solutions containing
12
13 acetonitrile (0.1% formic acid or 20 mM ammonium formate (pH 3.40), respectively, in
14
15 water/acetonitrile, 50/50, v/v).

16 ***On-line trapping of oxidation product with glutathione (N-acetylcysteine)***

17
18 To identify possible S-conjugate(s) with GSH (NAC), the EC/MS set-up described above
19
20 was slightly modified (Figure 1b). A 10- μM 2-OH-AC solution in electrolyte was pumped
21
22 through the electrochemical cell at a constant flow rate of 30 $\mu\text{L}\cdot\text{min}^{-1}$. 100 μM GSH (NAC) in
23
24 ultrapure water was added at the same flow rate to the effluent of the electrochemical cell via
25
26 a T-piece into a 100 μL mixing coil. The effluent from the mixing coil was injected directly into
27
28 the ESI-MS interface.

29 ***In vitro microsomal incubations***

30
31
32 Human or rat liver microsomal incubations (1 $\text{mg}\cdot\text{mL}^{-1}$ protein) were performed with 50
33
34 μM 2-OH-AC, 1 mM NADPH, and 1 mM GSH in a potassium phosphate-buffered solution
35
36 (0.1 M, pH 7.40) at 37 °C for 1 h, in a total volume of 100 μL . The incubations were
37
38 terminated by adding ice-cold methanol to the incubation mixtures (50:50, v/v). The samples
39
40 were then vortexed, placed in ice for 10 min, and centrifuged at 10000 x g for 10 min.
41
42 Aliquots of the supernatants (150 μL) were then analyzed directly by reversed-phase LC with
43
44 UV-Vis detection and/or diode array and multiple wavelength detection, and monitored by
45
46 MS. The three types of control incubations were applied, the first without test compound, the
47
48 second without NADPH, and the third without GSH.

Results

In order to provide an important insight into the mechanism of antitumor action of imidazo- and triazoloacridinones, the electrochemical oxidation of their simpler reference compound,

2-OH-AC, was investigated. The ability of 2-OH-AC to undergo oxidative metabolic activation was studied by applying the direct combination of electrochemistry and mass spectrometry (EC/MS). For 2-OH-AC no EC/MS data were found before. Electrochemical conversion of 2-OH-AC was conducted in parallel to controlled-potential electrolysis (CPE) and *in vitro* experiments using liver microsomes.

Controlled-potential electrolysis of 2-OH-AC

At first, cyclic voltammetry (CV) was used to initially investigate the electrochemistry of 2-OH-AC at a glassy carbon electrode in a range of positive potentials and to determine the optimum operating range of voltages for further EC/MS studies. In order to confirm the postulated oxidation mechanism of 2-OH-AC action and to identify the oxidation products, controlled-potential electrolysis of 2-OH-AC (0.5 mM in 0.02 M PBS buffer, pH 7.40) was performed at 0.50 V. To support the possibility of the formation of the stable adduct between 2-OH-AC intermediate and GSH, the tests were performed in the absence and the presence of GSH (5 mM in ultrapure water). The electrolysis measurements were carried out for 24 hours and the progress of the process was monitored periodically by application of voltammetric, spectroscopic, chromatographic, and spectrometric methods.

Data for the CV are provided in the supplemental material. In the solution of pure 2-OH-AC one anodic peak (a1) and three cathodic peaks (c1, c2, c3) were obtained. The appearance of three cathodic peaks indicated that the oxidation product of 2-OH-AC underwent subsequent chemical reactions. The addition of 10-fold excess of GSH to 2-OH-AC solution resulted in a shift of the oxidation peak of 2-OH-AC of about 30 mV to a more positive potential. In turn, the signal appearing in the cathodic part of the CV at circa 0.32 V virtually disappeared and two other cathodic peaks were substantially depressed. These

1
2
3 changes confirmed the interaction between 2-OH-AC and GSH, and the reducing effect of
4
5 GSH against the oxidation reaction products.

6
7 Assuming that during oxidation process of 2-OH-AC two electrons are exchanged, the
8
9 total electrolysis of 0.5 mM compound should be achieved at the time of obtaining the charge
10
11 of 200 mC. Unfortunately, despite long electrolysis (over 1 day) the maximum charge which
12
13 was achieved was only 29 mC. A possible explanation of this situation is that the oxidation
14
15 products of 2-OH-AC strongly adsorbed on the electrode surface and effectively blocked it.
16
17 With electrolysis progress the intensity of the current signals drastically decreased (Figure
18
19 **2a**). In addition, a new oxidation signal at circa 0.06 V (**a2**) was also well visible. It was
20
21 related to the third cathodic peak (**c1**). The introduction of GSH to the solution of 2-OH-AC
22
23 significantly minimized the effects of the electrolysis process. This observation may point out
24
25 the presence of interactions between oxidation products of 2-OH-AC and GSH. The CPE of a
26
27 mixture of 2-OH-AC and GSH (Figure **2b**) revealed the anodic signal at circa 0.42 V (**a1**) with
28
29 increasing intensity. Also a new signal emerged at circa 0.31 V (**a2**) and its height increased
30
31 with time of electrolysis. This product peak was due to the reduction of the transformed 2-
32
33 OH-AC molecule.

34 ***Electrochemical simulation of the oxidative metabolism of 2-OH-AC (phase I*** 35 ***metabolism)***

36
37
38 The electrochemical oxidation products of 2-OH-AC were generated and identified using
39
40 on-line **EC/MS** (Figure **1a**). The applied set up enabled us to simulate metabolic P450-
41
42 catalyzed reactions occurring in the liver. To reduce the complexity of the system these
43
44 experiments were carried out only with flow injection EC/MS without using an LC column.

45
46 The best electrochemical conversion of 2-OH-AC into its expected products was
47
48 achieved using the scan mode in the potential range of 0 – 2.5 V. To provide a concise
49
50 overview of the oxidation products, 2-D mass voltammograms were generated by plotting the
51
52 extracted ion intensities versus the progress of the electrochemical oxidation. Mass
53
54 spectrometry allowed the identification of the formed products by an increasing signal
55
56 intensity of the corresponding mass-to-charge (m/z) ratio. Furthermore, for structure
57
58
59
60

1
2
3 elucidation of the detected products, fragment ions, generated by in-source fragmentation in
4 the ESI interface, have been studied.

5
6 A representative 2-D mass voltammogram resulting from the oxidation of 2-OH-AC within
7 the scanned range is shown in **Figure 3**. The 2-OH-AC molecule was easily protonated and
8 detected with high intensities as $[M+H]^+$ ion (m/z 212) in the positive ionization mode of the
9 ESI-Q-TOF mass spectrometer. No products were observed in the solution of pure 2-OH-AC
10 without any electrochemical potential (cell off) (the inset of top left corner in **Figure 3**),
11 whereas significant drop in an extracted ion intensity of 2-OH-AC was noticed when the
12 potential was applied (cell on) (the inset of top right corner in **Figure 3**). This change is
13 attributed to the oxidation of 2-OH-AC into eight products in an electrochemical thin-layer
14 cell. All products are summarized in Table 2 and will be discussed in detail, with specific
15 reference to their accurate mass data and fragmentation patterns.

16
17 Furthermore, we noticed that the ratios of oxidation products of 2-OH-AC were strongly
18 dependent on the solvent composition. Most products identified using methanol-containing
19 electrolyte, with the exception of P4 (m/z 240) and P8 (m/z 421), have been also observed in
20 solutions of acetonitrile. No **additional** oxidation products of 2-OH-AC were detected under
21 these conditions. Higher signal intensity of the selected mass ions was caused by that
22 methanol was better than acetonitrile in diminishing the adsorption of the electrochemical
23 products on the surface of the working electrode. Moreover, methanol/water electrolyte
24 solution produced the lowest mass background noise in positive total ion chromatogram
25 (data not shown). **A summary of the electrochemical products observed under different
26 electrolyte conditions is available in the supplemental material.**

27 ***Characterization of GSH conjugate formed from electrochemical oxidation of 2-OH-AC*** 28 ***(phase II metabolism)***

29
30 In this work, the **EC/MS** set-up described above was extended to a system allowing the
31 study of the conjugation of 2-OH-AC oxidation products with a reactive endogenous
32 nucleophile, such as reduced GSH or NAC. GSH and NAC were selected because of a
33 simple structure with their soft nucleophilic thiol group and their relevance in living organisms
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49

(Evans *et al.* 2004). The instrumental set-up for electrochemical simulation of conjugation reactions (phase II reactions) is shown in Figure 1b.

Observation of the potential conjugation reaction(s) between the oxidation product(s) of 2-OH-AC and GSH was possible in the form of a 2-D mass voltammogram (Figure 4a).

The mass spectra were acquired to confirm the formation of GSH S-conjugate (Figure 5). As expected, no additional signals besides those associated with protonated 2-OH-AC (m/z 212) and GSH (m/z 308) were observed at cell off (Figure 5a and a zoom in the mass spectrum from the circle range). However, in the potential range of 0 – 2.5 V (cell on) mass spectrum revealed one m/z signal representing potential GSH S-conjugate (Figure 5b and a zoom in the mass spectrum from the circle range). Ion with a m/z ratio of 517 was observed with a weak intensity which was increasing only in the case when an extracted ion intensity of protonated 2-OH-AC was decreasing. It was confirmed by the accurate mass measurements that m/z 517 represents the conjugate of molecular formula $C_{23}H_{24}N_4O_8S$, consistent with a product of the 2-OH-AC oxidation and one molecule of GSH. This result is in accordance with the assumption that GSH traps soft electrophiles with its thiol group (Inoue *et al.* 2015).

To confirm that the ion at m/z 517 is originating from GSH conjugate of 2-OH-AC, the fragmentation spectrum was recorded (Figure 5c). The MS/MS of m/z 517 ion produced two fragment ions at m/z 388 and 244, respectively. The accurate values of m/z correlated well with calculated m/z (Table 3). The first was consistent with the neutral loss of anhydroglutamic acid (-129), whereas the second (-275) was assigned as a cleavage adjacent to the cysteinyl thioether moiety with charge retention on the 2-OH-AC molecule. Thus, the fragmentation pattern of m/z 517 ion suggested the presence of at least one GSH moiety. As for GSH, formation of NAC S-conjugate with a m/z ratio of 373 was also observed in similar experiments (Figure 4b). The MS/MS of 373 ion revealed the presence of the fragment ion at m/z 244 (data not shown), exactly the same that occurred in MS/MS of m/z 517 ion.

Identification of GSH conjugate of 2-OH-AC from *in vitro* microsomal incubations and controlled-potential electrolysis

Trapping experiments with GSH in *in vitro* microsomal incubations and during controlled-potential electrolysis of 2-OH-AC were carried out to generally assess the feasibility of GSH S-conjugate formation. The results obtained from these approaches were compared with those from a purely instrumental electrochemical simulation. Figure 6a presents the representative high performance LC chromatograms recorded for enzymatic and electrochemical oxidation of 2-OH-AC in the absence and the presence of 10-fold excess of GSH.

Due to various expressions of the different isoforms of P450 in each organism, the metabolism in human and rat liver microsomes may result in different metabolites or in a different quantitative distribution of the metabolites (Martignoni *et al.* 2006). When experiments were performed using both types of microsomal fractions, in the case of 2-OH-AC we observed only slight differences in the intensity of the individual peaks, hence the further discussion will be based on the results from RLM studies. The microsomal incubations included three controls, one without 2-OH-AC to rule out any potential interference/contamination from endogenous compounds, the second without NADPH, a cofactor for cytochrome P450 activities, to assess the metabolic dependence of GSH conjugate formation, and the third without GSH. No conjugate was detected in the “no compound” and “no NADPH” control reactions (data shown in the supplemental material). LC analysis of microsomal samples without GSH revealed that three main metabolites were formed under the conditions studied (Figure 6a). Chromatogram taken after 60 min incubation of 2-OH-AC with 10-fold excess of GSH represents one additional chromatographic peak. It is noteworthy that it was not seen in the control incubations without NADPH, which suggests an NADPH-dependent oxidation of 2-OH-AC to this metabolite. LC analysis of the reaction mixture throughout the course of controlled-potential electrolysis revealed one main product (Figure 6a). Compared to enzymatically generated metabolites,

1
2
3 two of them were missing. As before, one additional peak was observed only where GSH
4 was present in excess, and its intensity depended on the progress of electrolysis.
5

6 In both approaches the UV-Vis spectra of the peaks at 18.6 min differed significantly from
7 that of the substrate (the inset in Figure 6a). The shift towards longer wavelengths may
8 indicate that a more extensive delocalized electron system exists in this product in
9 comparison to that in 2-OH-AC. It was subsequently identified by ESI-MS as GSH S-
10 conjugate with the mass ion at m/z 517 (data not shown). Therefore, the above results
11 agreed with the data obtained from EC/MS or EC/MS/MS measurements and confirmed the
12 existence of a reactive intermediate in the oxidative pathway of 2-OH-AC. However, due to
13 the low concentration of the S-conjugate in the EC/MS system it was not possible to obtain
14 its chromatographic peak and UV-Vis spectrum. Also other products showing quite good
15 intensities in the EC/MS system, were found in small or trace amounts in LC analysis (Figure
16 6b). Some of them were not detected, which is likely to be a consequence of their low
17 stability. Nevertheless, it is worth to note that generally the EC system allowed to obtain a
18 wider set of phase I metabolites than the use of classical approaches, probably because of
19 clean matrix.
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Discussion

The toxic effects of drugs and other xenobiotics arise not only from the compound itself but also from its metabolites (Baumann *et al.* 2010). Hence, during drug development process particular attention is paid to drug metabolite formation and identification of metabolic pathways. Progress in this research area depends critically on the improvement of methods involved in the generation and analysis of various types of drug metabolites, with special respect to the characterization of reactive metabolites (Prakash *et al.* 2008). In this work different approaches for the investigation of the 2-OH-AC oxidative metabolism, that may occurs in the living organism, were performed. The oxidative electrochemical behavior of 2-OH-AC was first investigated by the direct combination of electrochemistry and mass spectrometry. The applicability of the electrochemistry in drug metabolite synthesis was evaluated by conventional electrochemical (controlled-potential electrolysis) and enzymatic (microsomal incubation) approaches.

The electrochemical conversion of 2-OH-AC in an electrochemical thin-layer cell was successfully achieved (Figure 3). Table 2 consists of list of major products related to 2-OH-AC oxidative metabolism and m/z ratios of the protonated species $[M+H]^+$ used for mass spectra interpretation. The resulting molecular formulas were deduced by their accurate masses. The isotopic patterns correlated well with theoretical calculations. For all compounds, the deviations between the calculated and measured m/z values were less than 4 ppm.

The proposed chemical reactions that may occur in the electrochemical cell are summarized in Figure 7. Most of 2-OH-AC oxidation products seem to be species containing additional oxygen atom and can exist in at least two tautomeric forms. According to previous studies on the oxidative transformations of 2-OH-AC (Mazerska *et al.* 2002), the most likely oxidation site in the 2-OH-AC molecule is the position *ortho* to the hydroxyl group. This relates to the positions 1 or 3 of 2-OH-AC, wherein position 1 remains preferential. Thus, the mass ion at m/z 226 is proposed to correspond to 1,2-orthoquinone (P1a), while the signal at m/z 228 may include monohydroxylation product (P3). The molecular ion observed with little

1
2
3 signal intensity at m/z 421 was identified to be the dimer formed from two substrate
4 molecules of radical structure linked together at position 1 of 2-OH-AC (P8). The chemical
5 structures of these three products have been previously determined by means of MS and
6 NMR spectroscopy (Mazerska *et al.* 2002).
7
8
9

10 Other products of oxidative metabolism of 2-OH-AC have been identified for the first time
11 upon electrochemical oxidation. The P2 compound with the mass ion at m/z 227 may have
12 been formed via *C*- or *N*-hydroxylation because the mass corresponds to an addition of
13 15.99 Da to the parent compound with a lack of one hydrogen atom. As a result, the P2
14 molecule should contain a positive charge, probably located on the nitrogen atom of the
15 acridinone ring. Further, considering the mesomeric effects in electron density distribution,
16 the loss of one hydrogen atom in P2 compound would indicate the possibility of the formation
17 other than P1a structures for the mass ion at m/z 226, including P1b.
18
19
20
21
22
23
24
25

26 The products showing the mass ions at m/z 240 (P4), 242 (P5), 256 (P6), and 258 (P7)
27 have not been definitively identified. However, we propose here only their tentative
28 structures. The accurate mass of ion at m/z 242 (P5) differed from the accurate mass of ion
29 at m/z 226 (P1) by exactly 16.03 Da. This value may indicate the presence of the additional
30 one carbon and four hydrogen atoms, but not a single oxygen atom (15.99 Da), somewhere
31 within the P5 molecule. During the electrochemical and/or the ionization processes some
32 side reactions can take place, like the reaction of electrolyte components with
33 electrochemically generated intermediates and sometimes they are unexpected. Based on
34 the literature data (Sichilongo *et al.* 2011; Wang *et al.* 2011), electro-oxidation of methanol
35 from electrolyte may give a strongly electrophilic methylum (CH_3^+) or methyl radical ($\cdot\text{CH}_3$).
36 Thus, we can assume that it is quite likely that compound containing a methyl group in its
37 structure was formed. The probable methylation of 2-OH-AC derivative have not been
38 avoided even when acetonitrile was present in the electrolyte solution (supplemental
39 material). The possible location of attachment is speculated to be an electronegative nitrogen
40 atom due to valence considerations. Presumably further two-electron two-proton
41 dehydrogenation of P5 provided P4 (m/z 240), with molecular mass ion decreased by 2.01
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 Da. In turn, a product with the mass ion at m/z 258 (P7) could have been created by the
4 hydroxylation of the methyl group of P5, and its further two-electron two-proton
5 dehydrogenation gave a signal at m/z 256 (P6).
6
7

8
9 It should be pointed out that the signals at m/z 240 and m/z 256 may correspond to
10 aldehyde products as well. To test whether the hydroxymethyl group at the nitrogen atom in
11 P5 or in P7 products underwent further oxidation, trapping experiments were performed with
12 semicarbazide. It is a small molecule trapping agent that can form a Schiff base with
13 aldehydes in the process mimicking reactions between aldehyde metabolites with lysine
14 residues on proteins (Evans *et al.* 2004). Semicarbazide was added to the electrochemical
15 oxidation mixture of 2-OH-AC followed by LC/MS analysis. Trapping with semicarbazide did
16 not give any conjugates what excluded the presence of aldehyde products in oxidative
17 transformations of 2-OH-AC.
18
19
20
21
22
23
24
25

26 To simulate the phase II metabolism of 2-OH-AC, electrochemical oxidation of the
27 compound was carried out in the presence of an excess amount of nucleophilic trapping
28 agent. EC/MS successfully predicted the formation of chemically reactive metabolite that
29 spontaneously reacted with reduced glutathione (Figures 4a and 5) and/or *N*-acetylcysteine
30 (Figure 4b) to form conjugates (m/z 517 and m/z 373, respectively) via the thiol group.
31 General structures for probable GSH and NAC conjugates of 2-OH-AC are shown in Figure
32 8. We suspect that 2-OH-AC is likely to undergo a P450-catalyzed oxidative dehydrogenation
33 which results in the formation of a reactive intermediate in the form of a quinone imine
34 (Guengerich 2007), as, *e.g.*, it takes place in a metabolic pathway of the acetaminophen
35 (Jollow *et al.* 1973; Larson 2007). For this mechanism, a quinone imine of 2-OH-AC has to
36 be the precursor of the adduct as the corresponding quinone imine would have the m/z ratio
37 of 210 in the positive-ion mode detection. However, this intermediate product, presumably
38 due to its short life span and high reactivity, was not directly detected in the mass
39 voltammogram of pure 2-OH-AC. It is important to note that the same type of GSH S-
40 conjugate was detected after *in vitro* liver microsomal incubations in an NADPH-dependent
41 manner and in the reaction mixture from the electrolysis experiment (Figure 6). Quinone
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 imines are well known reactive intermediates which very often undergo adduct formation with
4 crucial cellular compounds such as GSH, structural proteins, enzymes and/or DNA (Bolton *et*
5 *al.* 2000; Zhou *et al.* 2005). As a consequence, these compounds can cause a variety of
6 hazardous effects *in vivo*, including acute cytotoxicity, immunotoxicity, and carcinogenesis,
7 especially in the case of depleted levels of cellular GSH (Lohmann *et al.* 2010). However, the
8 fact that reactive metabolite of 2-OH-AC may exist and, as it has been shown, may
9 undergo
10 adduct formation does not consequently imply that it causes toxicity (Evans *et al.* 2004) so
11 further studies are needed to elucidate whether this metabolite contributes to the 2-OH-AC
12 toxicity *in vivo*.
13
14
15
16
17
18
19

20 The results of our studies clearly prove that EC-based approach has the potential to
21 simulate the majority of oxidative metabolism reaction, including the simulation of reactive
22 metabolite formation and its binding to biomolecules (e.g., GSH). The observed
23 dehydrogenation, hydroxylation and oxidation of acridinone molecule are in general
24 agreement with reports on the types of P450-catalyzed reactions being simulated by
25 electrochemistry (Lohmann & Karst 2008; Lohmann *et al.* 2010). Our observations on
26 product structures may be advantageous from the viewpoint of the structural factors that
27 influence the reactivity and functional group interactions of the compound, providing a good
28 approximation of what may occur *in vivo*.
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Conclusions

The present study demonstrated the significance of electrochemistry coupled on-line with mass spectrometry in drug metabolism studies. This combination appeared to be a suitable tool for a simulation of some types of oxidative drug metabolic reactions related to cytochromes P450, and for studying the formation of reactive metabolites. In this investigation, 2-hydroxyacridinone (2-OH-AC), the reference compound for antitumor-active imidazo- and triazoloacridinone derivatives, was easily oxidized in an electrochemical thin-layer cell to several different products. To trap potentially reactive metabolite(s) two types of nucleophilic trapping agents, reduced GSH and NAC, both with nucleophilic thiol group, were used. This allowed us to identify and characterize a novel GSH (NAC) S-conjugate of 2-OH-AC. The postulated reactive quinone imine metabolite of 2-OH-AC may potentially be involved in a number of biochemical transformations, and can be responsible for the antitumor activity of acridinone derivatives. The role of this reactive metabolite requires further investigation.

The electrochemical method we proposed here represented a good alternative for classical metabolic studies with the principal advantage which is the absence of proteins in the reaction medium. It will be very useful in further studies on metabolic transformations of antitumor imidazo- and triazoloacridinone drugs. Considering pharmacological aspect, our findings may provide important guidelines for the further modification of the acridinone compounds so they contribute to the design and the development of safer and more effective therapeutic agents.

Acknowledgments

This work was supported by the National Science Center (Poland) under Grant number 2012/07/D/NZ7/03395..

For Peer Review Only

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Declaration of interest statement

The authors confirm that this article content has no conflicts of interest.

For Peer Review Only

References

Acheson RM. (1973). Acridines. In: Acheson RM, ed. The chemistry of heterocyclic compounds. New York, Interscience Publishers, Inc., p. 141.

Augustin E, Moś-Rompa A, Skwarska A, Witkowski JM, Konopa J. (2006). Induction of G2/M phase arrest and apoptosis of human leukemia cells by potent antitumor triazoloacridinone C-1305. *Biochem Pharmacol* 72: 1668-1679.

Baillie TA, Cayen MN, Fouda H, Gerson RJ, Green JD, Grossman SJ, *et al.* (2002). Drug metabolites in safety testing. *Toxicol Appl Pharmacol* 182: 188-196.

Baumann A, Lohmann W, Rose T, Ahn KC, Hammock BD, Karst U, *et al.* (2010). Electrochemistry-mass spectrometry unveils the formation of reactive triclocarban metabolites. *Drug Metab Dispos* 38: 2130-2138.

Bolton JL, Trush MA, Penning TM, Dryhurst G, Monks TJ. (2000). Role of quinones in toxicology. *Chem Res Toxicol* 13: 135-160.

Brandon EF, Raap CD, Meijerman I, Beijnen JH, Schellens JH. (2003). An update on in vitro test methods in human hepatic drug biotransformation research: pros and cons. *Toxicol Appl Pharmacol* 189: 233-246.

Bussy U, Boisseau R, Thobie-Gautier C, Boujtita M. (2015). Electrochemistry-mass spectrometry to study reactive drug metabolites and CYP450 simulations. *Trends Anal Chem* 70: 67-73.

Bussy U, Boujtita M. (2014). Advances in the electrochemical simulation of oxidation reactions mediated by cytochrome P450. *Chem Res Toxicol* 27: 1652-1668.

1
2
3 Cholody WM, Horowska B, Paradziej-Lukowicz J, Martelli S, Konopa J. 1996. Structure-
4 activity relationship for antineoplastic imidazoacridinones: synthesis and antileukemic activity
5 in vivo. *J Med Chem* 39: 1028-1032.
6
7

8
9
10 Cholody WM, Martelli S, Konopa J. (1990). 8-Substitued 5-[(aminoalkyl)amino]-6H-v-
11 triazolo[4,5,1-de]acridin-6-ones as potential antineoplastic agents. Synthesis and biological
12 activity. *J Med Chem* 33: 2852-2856.
13
14

15
16
17 Cholody WM, Martelli S, Lukowicz J, Konopa J. (1990). 5-[(Aminoalkyl)amino]imidazo[4,5,1-
18 de]acridin-6-ones as a novel class of antineoplastic agents. Synthesis and biological activity.
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

J Med Chem 33: 49-52.

Dziegielewski J, Konopa J. (1996). Interstrand crosslinking of DNA induced in tumor cells by
a new group of antitumor imidazoacridinones. *Proc Am Assoc Cancer Res* 37: 410.

Dziegielewski J, Konopa J. (1998). Characterisation of covalent binding to DNA of antitumor
imidazoacridinone C-1311, after metabolic activation. *Ann Oncol Suppl* 1: 137.

Dziegielewski J, Slusarski B, Konitz A, Skladanowski A, Konopa J. (2002). Intercalation of
imidazoacridinones to DNA and its relevance to cytotoxic and antitumor activity. *Biochem
Pharmacol* 63: 1653-1662.

Evans DC, Watt AP, Nicoll-Griffith DA, Baillie TA. (2004). Drug-protein adducts: An industry
perspective on minimizing the potential for drug bioactivation in drug discovery and
development. *Chem Res Toxicol* 17: 3-16.

1
2
3 Faber H, Jahn S, Kunнемeyer J, Simon H, Melles D, Vogel M, *et al.* (2011).
4 Electrochemistry/liquid chromatography/mass spectrometry as a tool in metabolism studies.
5 *Angew Chem* 50: A52-58.
6
7

8
9
10 Guengerich FP. (2007). Oxidative, reductive and hydrolytic metabolism of drugs. In: Zhang
11 D, Zhu M, Humphreys WG, eds. *Drug metabolism in drug design and development*. New
12 York, John Willey & Sons, Inc., p. 265.
13
14

15
16
17
18 Harding MM, Grummitt AR. (2003). 9-hydroxyellipticine and derivatives as chemotherapy
19 agents. *Mini Rev Med Chem* 3: 67-76.
20
21

22
23
24 Inoue K, Fukuda K, Yoshimura T, Kusano K. (2015). Comparison of the reactivity of trapping
25 reagents toward electrophiles: cysteine derivatives can be bifunctional trapping reagents.
26 *Chem Res Toxicol* 28: 1546-1555.
27
28

29
30
31
32 Jennings GS, Strauss M. (1999). Immortalization of hepatocytes through targeted
33 deregulation of the cell cycle. In: Al-Rubeai M, ed. *Cell engineering*. the Netherlands, Kluwer
34 Academic Publishers, p. 259.
35
36

37
38
39
40 Jollow DJ, Mitchell JR, Potter WZ, Davis DC, Gillette JR, Brodie BB. (1973). Acetaminophen-
41 induced hepatic necrosis. II. Role of covalent binding in vivo. *J Pharmacol Exp Ther* 187:
42 195-202.
43
44

45
46
47
48 Jurva U, Wikstrom HV, Weidolf L, Bruins AP. (2003). Comparison between
49 electrochemistry/mass spectrometry and cytochrome P450 catalyzed oxidation reactions.
50 *Rapid Commun Mass Spectrom* 17: 800-810.
51
52

1
2
3 Kalgutkar AS, Soglia JR. (2005). Minimising the potential for metabolic activation in drug
4 discovery. *Expert Opin Drug Metab Toxicol* 1: 91-142.
5
6

7
8 Koba M, Konopa J. (2007). Interactions of antitumor triazoloacridinones with DNA,. *Acta*
9 *Biochim Pol* 54: 297-306.
10
11

12
13
14 Kusnierczyk H, Cholody WM, Paradziej-Lukowicz J, Radzikowski C, Konopa J. (1994).
15 Experimental antitumor activity and toxicity of the selected triazolo- and imidazoacridinones.
16 *Arch Immunol Ther Exper (Warsz)* 42: 415-423.
17
18
19

20
21
22 Larson AM. (2007). Acetaminophen hepatotoxicity. *Clin Liver Dis* 11: 525-548.
23
24

25
26 Lemke K, Wojciechowski M, Laine W, Bailly C, Colson P, Baginski M, *et al.* (2005). Induction
27 of unique structural changes in guanine-rich DNA regions by the triazoloacridone C-1305, a
28 topoisomerase II inhibitor with antitumor activities. *Nucleic Acids Res* 33: 6034-6047.
29
30
31

32
33
34 Lohmann W, Karst U. (2008). Biomimetic modeling of oxidative drug metabolism: strategies,
35 advantages and limitations. *Anal Bioanal Chem* 391: 79-96.
36
37

38
39
40 Lohmann W, Baumann A, Karst U. (2010). Electrochemistry and LC-MS for metabolite
41 generation and identification: tools, technologies and trends. *LC•GC Europe* January: 1-7.
42
43
44

45
46 Marks TA, Venditti JM. (1976). Potentiation of actinomycin D or adriamycin antitumor activity
47 with DNA. *Cancer Research* 36: 496-504.
48
49

50
51
52 Martignoni M, Groothuis GM, de Kanter R. (2006). Species differences between mouse, rat,
53 dog, monkey and human CYP-mediated drug metabolism, inhibition and induction. *Expert*
54 *Opin Drug Metab Toxicol* 2: 875-894.
55
56
57

1
2
3 Mazerska Z, Dziegielewski J, Konopa J. (2001). Enzymatic activation of a new antitumour
4 drug 5-diethylaminoethylamino-8-hydroxyimidazo-acridinine, C-1311, observed after its
5 intercalation into DNA. *Biochem Pharmacol* 61: 685-694.
6
7

8
9
10 Mazerska Z, Zamponi S, Marassi R, Martelli S, Konopa J. (1997). Electrochemical oxidation
11 of antitumor imidazoacridinone derivatives and the reference 2-hydroxyacridinone. *J.*
12 *Electroanal Chem* 427: 71-78.
13
14

15
16
17 Mazerska Z, Zamponi S, Marassi R, Sowinski P, Konopa J. (2002). The products of electro-
18 and photochemical oxidation of 2-hydroxyacridinone, the reference compound of antitumor
19 imidazoacridinone derivatives. *J Electroanal Chem* 521: 144-154.
20
21
22

23
24
25 Nouri-Nigjeh E, Bischoff R, Bruins AP, Permentier HP. (2011). Electrochemistry in the
26 mimicry of oxidative drug metabolism by cytochrome P450s. *Curr Drug Metab* 12: 359-371.
27
28
29

30
31
32 Orhan H, Vermeulen NPE. (2011). Conventional and novel approaches in generating and
33 characterization of reactive intermediates from drugs/drug candidates. *Curr Drug Metab* 12:
34 383-394.
35
36
37

38
39
40 Park BK, Boobis A, Clarke S, Goldring CE, Jones D, Kenna JG, *et al.* (2011). Managing the
41 challenge of chemically reactive metabolites in drug development. *Nat Rev Drug Discov* 10:
42 292-306.
43
44
45

46
47
48 Permentier HP, Bruins AP, Bischoff R. (2008). Electrochemistry-mass spectrometry in drug
49 metabolism and protein research. *Mini Rev Med Chem* 8: 45-56.
50
51

52
53
54 Pinto N, Dolan ME. (2011). Clinically relevant genetic variations in drug metabolizing
55 enzymes. *Curr Drug Metab* 12: 487-497.
56
57

1
2
3 Prakash C, Sharma R, Gleave M, Nedderman A. (2008). In vitro screening techniques for
4 reactive metabolites for minimizing bioactivation potential in drug discovery. *Curr Drug Metab*
5 9: 952-964.
6
7

8
9
10 Sichilongo KF, Famuyiwa SO, R. Kibechu R. (2011). Pre-electrospray ionisation manifold
11 methylation and post-electrospray ionisation manifold cleavage/ion cluster formation
12 observed during electrospray ionisation of chloramphenicol in solutions of methanol and
13 acetonitrile for liquid chromatography-mass spectrometry employing a commercial
14 quadrupole ion trap mass analyser. *Eur J Mass Spectrom (Chichester)* 17: 255-264.
15
16
17

18
19 Skladanowski A, Plisov SY, Konopa J, Larsen AK. (1996). Inhibition of DNA topoisomerase II
20 by imidazoacridinones, new antineoplastic agents with strong activity against solid tumors.
21 *Mol Pharmacol* 49: 772-780.
22
23
24

25
26 Srivastava A, Maggs JL, Antoine DJ, Williams DP, Smith DA, Park BK. (2010). Role of
27 reactive metabolites in drug-induced hepatotoxicity. *Handb Exp Pharmacol* 196 (2010) 165-
28 194.
29
30
31

32
33 Volk KJ, Yost RA, Brajter-Toth A. (1992). Electrochemistry on line with mass spectrometry
34 insight into biological redox reactions. *Anal Chem* 64: 21A-26A.
35
36
37

38
39 Wang L, Chai Y, Tu P, Sun C, Pan Y. (2011). Formation of [M + 15](+) ions from aromatic
40 aldehydes by use of methanol: in-source aldolization reaction in electrospray ionization mass
41 spectrometry. *J Mass Spectrom* 46: 1203-1210.
42
43
44

45
46 Zanger UM, Schwab M. (2013). Cytochrome P450 enzymes in drug metabolism: Regulation
47 of gene expression, enzyme activities, and impact of genetic variation. *Pharmacol Therapeut*
48 138: 103-141.
49
50
51

1
2
3
4 Zhou S, Chan E, Duan W, Huang M, Chen YZ. (2005). Drug bioactivation, covalent binding
5 to target proteins and toxicity relevance. Drug Metab Rev 37: 41-213.
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

For Peer Review Only

1
2
3 **Tables with captions**

4 **Table 1**

5 EC and ESI-Q-TOF-MS(/MS) parameters as applied in direct EC/MS(/MS) and in MS(/MS)
6
7 experiments for determination of accurate masses of product ions and getting ion
8
9 fragmentation.
10
11

	Parameter	Value or setting
EC settings	Flow rate	30 $\mu\text{L}\cdot\text{min}^{-1}$
	Potential	0 – 2.5 V (10 mV steps)
	EC operating mode	scan
	Cycle	continuous
MS(/MS) settings	The range of m/z	100 – 600
	Ion source	dual ESI
	Ion polarity	positive
	MS operating mode	scan
	Capillary voltage	3500 V
	Nebulizer gas (N_2) pressure	35 psig
	Drying gas (N_2) flow	10 $\text{L}\cdot\text{min}^{-1}$
	Drying gas temperature	325 $^{\circ}\text{C}$
	Fragmentor	175 V
	Skimmer	45 V
	OCT 1 RF V_{pp}	750 V
	Rate	1,5 $\text{spectra}\cdot\text{s}^{-1}$
MS/MS method	targeted	
- Slope	- 4 m/z	
- Offset	- 5 V	

Table 2

A summary of molecular formulas of 2-OH-AC and its products found after electrochemical oxidation on a GC working electrode in a potential range of 0 – 2.5 V versus Pd/H₂. Base

Product	Measured <i>m/z</i> / Da	Calculated <i>m/z</i> / Da ^a	Deviation / ppm	Molecular formula of [M+H] ⁺ ion	MS/MS fragment ions	Predicted modification of 2-OH-AC molecule
2-OH-AC	212.0712	212.0706	2.9	C ₁₃ H ₁₀ NO ₂	-	-
P1a, b	226.0501	226.0499	1.1	C ₁₃ H ₈ NO ₃	212	- 2H + O
P2	227.0578	227.0577	0.5	C ₁₃ H ₉ NO ₃	212	- H + O
P3	228.0648	228.0655	-3.1	C ₁₃ H ₁₀ NO ₃	212, 226, 227	+ O
P4	240.0658	240.0655	1.2	C ₁₄ H ₁₀ NO ₃	212, 227 , 228	+ CO
P5	242.0818	242.0812	2.6	C ₁₄ H ₁₂ NO ₃	212, 228 , 240	+ 2H + CO
P6	256.0614	256.0604	3.8	C ₁₄ H ₁₀ NO ₄	212, 226, 228 , 240, 242	+ CO ₂
P7	258.0758	258.0761	-1.2	C ₁₄ H ₁₂ NO ₄	212, 226, 228 , 240, 242, 256	+ 2H + CO ₂
P8	421.1176	421.1183	-1.6	C ₂₆ H ₁₇ N ₂ O ₄	212	2-OH-AC dimer

fragment ions are shown in bold.

^a Calculated using Molecular Mass Calculator freeware version v2.02.

Table 3

A summary of neutral losses and fragmentations for ions attributed to the respective glutathione S-conjugate of the 2-OH-AC.

MS/MS collision-induced dissociation of glutathione S-conjugate			2-OH-AC		
			Measured <i>m/z</i> / Da	Calculated <i>m/z</i> / Da ^a	Deviation / ppm
Neutral losses	Parent	<i>m/z</i> / Da	517.1382	517.1387	1.04
	Glycine	75.0320	None	442.1067	None
	Anhydroglutamic acid	129.0426	388.0963	388.0962	-0.39
	Glutamine	146.0691	None	371.0696	None
	γ-Glu-Ala-Gly	275.1117	244.0421 ^b	244.0427	2.34
	Glutathione (S-oxide)	322.0709	None	195.0679	None

^a Calculated using Molecular Mass Calculator freeware version v2.02.

^b Neutral loss of γ-Glu-Ala-Gly + 2H.

Figure captions

Figure 1

Instrumental set-ups used for (a) the electrochemical simulation of the oxidative metabolism of 2-OH-AC and (b) the investigation of the reactivity of 2-OH-AC oxidation products towards GSH (NAC).

Legend: 1 – infusion syringe pump, 2 – potentiostat, 3 – electrochemical thin-layer cell (EC) – reactor cell, 4 – mass spectrometer (MS), 5 – T-piece and 100 μ L mixing coil

Chemical structures of the investigated 2-hydroxyacridinone (2-OH-AC) compound and glutathione (GSH)/N-acetylcysteine (NAC), used as nucleophilic trapping agents, are presented in the frame.

Figure 2

Cyclic voltammograms of 0.5 mM 2-OH-AC (a) alone and (b) in the mixture with 5 mM GSH in 0.02 M PBS buffer, pH 7.40, versus electrolysis progress. Experimental conditions: potential range -0.2 – 1.2 V; scan rate 100 $\text{mV}\cdot\text{s}^{-1}$; T = 21 $^{\circ}\text{C}$; ϕ GC 3 mm.

Figure 3

Two-dimensional plot (2-D mass voltammogram) of 10 μM 2-OH-AC oxidation at a GC working electrode (extracted ion intensity versus the progress of the electrochemical oxidation; positive-ion mode). The m/z ratios shown correspond to the protonated 2-OH-AC and its products (see legend). Experimental conditions: potential range 0 – 2.5 V; scan rate 10 $\text{mV}\cdot\text{s}^{-1}$, continuous; T = 21 $^{\circ}\text{C}$; ϕ GC 8 mm. The insets show the representative mass spectrum of 2-OH-AC without voltage applied to the electrochemical cell (cell off; top left corner) and after electrochemical oxidation (cell on; top right corner).

Figure 4

Two-dimensional plots (2-D mass voltammograms) of 10 μM 2-OH-AC oxidation at a GC working electrode in the presence of 100 μM (a) GSH and (b) NAC (extracted ion intensity versus the progress of the electrochemical oxidation; positive-ion mode). The m/z ratios shown correspond to the protonated 2-OH-AC and GSH or NAC S-conjugate (see legend).

1
2
3 Experimental conditions: potential range 0 – 2.5 V; scan rate 10 mV·s⁻¹, continuous; T = 21
4 °C; ϕ GC 8 mm.

5
6
7 **Figure 5**

8
9 The representative mass spectrum of a mixture of 2-OH-AC and GSH (a) without voltage
10 applied to the electrochemical cell (cell off) and (b) after electrochemical oxidation (cell on).
11
12 The insets show a zoom in the mass range from m/z 510 to 520 (positive-ion mode). (c)
13
14 Fragmentation spectrum of the conjugation product m/z 517.

15
16
17 **Figure 6**

18
19 (a) The representative high performance LC chromatograms of the reaction mixtures
20 obtained after metabolism simulation of 0.5 mM 2-OH-AC in the absence (black line) and the
21 presence (grey dashed line) of 5 mM GSH in RLM incubations (enzymatic oxidation) and
22 from controlled-potential electrolysis (oxidation by CPE). The inset shows UV-Vis spectra of
23
24 GSH S-conjugate and 2-OH-AC. (b) The representative extracted ion chromatograms for 2-
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
OH-AC oxidation in electrochemical cell.

30
31
32 **Figure 7**

32
33 Proposed oxidation reaction pathways of 2-OH-AC observed in the electrochemical cell.
34
35 Tentative structures were derived on the basis of accurate mass measurements and MS/MS
36
37 fragmentation patterns. Probable isomeric compounds are gathered in encircled blocks.

38
39
40 **Figure 8**

40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
Schematic representation of the proposed mechanism of the 2-OH-AC electrochemical
oxidation and GSH (NAC) S-conjugate formation.

Title:

Phase I and phase II metabolism simulation of antitumor-active 2-hydroxyacridinone with electrochemistry coupled on-line with mass spectrometry.

For Peer Review Only

Abstract

1. Here, we report the metabolic profile and the results of associated metabolic studies of 2-hydroxyacridinone (2-OH-AC), the reference compound for antitumor-active imidazo- and triazoloacridinones.
2. Electrochemistry coupled with mass spectrometry was applied to simulate the general oxidative metabolism of 2-OH-AC for the first time. The reactivity of 2-OH-AC products to biomolecules was also examined. The usefulness of the electrochemistry for studying the reactive drug metabolite trapping (conjugation reactions) was evaluated by the comparison with conventional electrochemical (controlled-potential electrolysis) and enzymatic (microsomal incubation) approaches.
3. 2-OH-AC oxidation products were generated in an electrochemical thin-layer cell. Their tentative structures were assigned based on tandem mass spectrometry in combination with accurate mass measurements. Moreover, the electrochemical conversion of 2-OH-AC in the presence of reduced glutathione and/or *N*-acetylcysteine unveiled the formation of reactive metabolite-nucleophilic trapping agent conjugates (m/z 517 and m/z 373, respectively) via the thiol group. This glutathione S-conjugate was also identified after electrolysis experiment as well as was detected in liver microsomes.
4. Summing up, the present work illustrates that the electrochemical simulation of metabolic reactions successfully supports the results of classical electrochemical and enzymatic studies. Therefore, it can be a useful tool for synthesis of drug metabolites, including reactive metabolites.

Keywords:

in vitro metabolism; metabolic activation; electrochemical oxidation; electrochemistry-mass spectrometry; metabolite electrosynthesis; reactive metabolite; glutathione S-conjugate;

For Peer Review Only

Introduction

Solid understanding of the metabolic pathways and the biotransformation mechanisms of new drug candidates is a crucial point in the drug discovery and development processes. Overall, it allows to elucidate the metabolic activation as well as the deactivation routes of new biologically active compounds, especially in respect to their possible toxicity (Park *et al.* 2011). The identification of metabolites helps to eliminate the inappropriate candidates at an early stage, before the more expensive development phases will be performed (Bussy & Boujtita 2014). Particularly, it is very important that the formation of chemically reactive intermediates is checked (Baillie *et al.* 2002), because an increasing number of reports indicate that they are responsible for the majority of rapid and unexpected drug toxic effects (Kalgutkar & Soglia 2005; Srivastava *et al.* 2010; Orhan & Vermeulen 2011). The relationship between drug metabolism and adverse drug reactions was first demonstrated with the analgesic agent acetaminophen (Jollow *et al.* 1973; Larson 2007). Reactive intermediates, generated usually via cytochrome P450 (P450)-catalyzed oxidative reactions, have the potential for covalent binding to cellular nucleophiles such as purine and pyrimidine bases of DNA or thiols of proteins, and form stable adducts. Adduct formation may alter biological functions of these biomolecules what ultimately leads to a toxic response (Brandon *et al.* 2003).

Metabolic activation of new drugs to reactive intermediates is currently assessed in the presence or the absence of reactive endogenous nucleophiles, such as reduced glutathione (GSH), *N*-acetylcysteine (NAC) or β -lactoglobulin A (β LGA), a model protein. In practise, the reaction usually involves the *in vitro* incubation of a studied compound with an excess of the selected chemical-trapping agent in hepatocytes or liver microsomes (as a source of cytochrome P450 isoenzymes). *In vivo* experiments involve laboratory animals (Kalgutkar & Soglia 2005). However, performing these biological schemes is usually laborious and time-consuming. Despite continuous improvements in metabolite-identification tools, the identification of some reactive metabolites remains difficult due to the matrix complexity, low concentration or their binding to matrix bio-components (Bussy *et al.* 2015). Isolated

1
2
3 hepatocytes have only a very limited proliferative potential *in vitro* and a correspondingly
4 short life-span in primary culture. Their fenotype is unstable and the level and activities of
5 enzymes, such as cytochromes P450, fall quickly in a few days (Jennings & Strauss 1999).
6
7 Also liver microsomes offer a limited reproducibility. Additionally, because of genetic
8 polymorphisms, variations in the gene expression for individual drug-metabolising
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

hepatocytes have only a very limited proliferative potential *in vitro* and a correspondingly short life-span in primary culture. Their fenotype is unstable and the level and activities of enzymes, such as cytochromes P450, fall quickly in a few days (Jennings & Strauss 1999). Also liver microsomes offer a limited reproducibility. Additionally, because of genetic polymorphisms, variations in the gene expression for individual drug-metabolising izoenzymes in each organism have to be taken into account (Pinto & Dolan 2011; Zanger & Schwab 2013).

As an alternative to the existing methods for studies on drug metabolism and toxicity, the electrochemical simulation of P450-catalyzed phase I reactions, mostly initiated by a single-electron transfer, has been developed (Volk *et al.* 1992; Lohmann & Karst 2008; Faber *et al.* 2011). Overall, it allows to simulate a wide variety of oxidation-reduction (redox) processes occurring in living organisms (Jurva *et al.* 2003; Nouri-Nigjeh *et al.* 2011). The combination of electrochemistry (EC) coupled on-line with mass spectrometry (MS) creates a powerful platform for rapid generation (in the electrochemical cell) and detection (by mass spectrometry) of a series of metabolic products, including observation of reactive metabolites (Permentier *et al.* 2008; Faber *et al.* 2011; Bussy & Boujtita 2014). Use of EC/MS improves the conventional methods of drug metabolism studies. It is a purely instrumental technique with a simple set-up that enables the generation of drug metabolites in the absence of biological matrices in the reaction medium. So, the application of EC/MS helps to overcome many of the laborious tasks related to isolation and identification of metabolic products formed *in vitro* (cultured hepatocytes, liver microsomes, purified enzymes) or *in vivo* (urine, plasma, *etc.*) (Orhan & Vermeulen 2011; Faber *et al.* 2011). Moreover, on-line combining EC with MS can provide valuable information about metabolically labile sites in a drug molecule and predict its reliable metabolic profile in a much shorter time (Jurva *et al.* 2003; Permentier *et al.* 2008).

Recently extensive research have been conducted in our group to determine the possible metabolic pathways of potential antitumor drugs and their model compounds. The objective of the investigations presented here was to develop and evaluate an on-line EC/MS method

1
2
3 for the simulation of oxidative metabolism of 2-hydroxyacridinone (2-OH-AC) (a structure in
4 the frame in Figure 1), a simple reference compound for high antitumor-active imidazo- and
5 triazoloacridinones.
6
7

8
9 The preliminary studies in this field were performed earlier by the application of cyclic
10 voltammetry, spectroelectrochemical measurements, and controlled-potential electrolysis
11 (Mazerska *et al.* 1997, 2002). These experiments indicated the ability of 2-OH-AC to undergo
12 oxidative metabolic activation in the living organism. However, no data for acridinone
13 derivatives are yet available using the direct coupling EC with MS. The studies presented
14 here were undertaken with respect to the usefulness of EC/MS technique for further
15 investigation on the oxidative metabolic activation and the identification of potential reactive
16 metabolites formed during the metabolism of antitumor-active imidazoacridinone C-1311 and
17 triazoloacridinone C-1305. These compounds, developed in our laboratory, have
18 demonstrated significant cytotoxic and antitumor activities (Cholody *et al.* 1990, 1996;
19 Kusnierczyk *et al.* 1994). The data obtained so far indicate that they have different spectrum
20 of antitumor activity and exhibit various mechanisms of action at the molecular level
21 (Dziegielewski & Konopa 1996; Skladanowski *et al.* 1996; Dziegielewski *et al.* 2002; Lemke
22 *et al.* 2005; Augustin *et al.* 2006). Metabolic activation is the leading concept for the reason
23 how imidazo- and triazoloacridinone derivatives cause high antitumor effect (Dziegielewski &
24 Konopa 1998; Mazerska *et al.* 2001; Koba & Konopa 2007). It is assumed that the hydroxyl
25 group in the acridinone ring provides the susceptibility of these compounds to oxidative
26 metabolism. The formation of the intermediate, characterized as a quinone imine structure,
27 that expresses antitumor activity, has been shown, for example, in the case of 9-
28 hydroxyellipticine (Harding & Grummitt 2003). A known quinone imine with activity against
29 human tumor cells is actinomycin D (Marks & Venditti 1976).
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49

50 In this work, the ability of EC/MS to expedite the generation and identification of the main
51 phase I metabolites of 2-OH-AC will be discussed. The formation of the reactive 2-OH-AC
52 intermediate metabolite and the possibility to simulate its covalent binding to biomolecule
53 (*i.e.*, glutathione (GSH) and/or *N*-acetylcysteine (NAC) as biomarkers of metabolic activity;
54
55
56
57
58
59
60

1
2
3 phase II metabolism) are also taken into account. Furthermore, to show the capability of
4 electrochemistry to simulate certain P450-mediated reactions, the results obtained by EC
5 were also compared with those gained after controlled-potential electrolysis (CPE) and
6 conventional *in vitro* studies by conducting incubations of 2-OH-AC with human and rat liver
7 microsomes (HLMs and RLMs, respectively). The products of electrolysis and enzymatic
8 transformations of 2-OH-AC were analysed by reversed-phase liquid chromatography (LC)
9 with UV-Vis detection and/or diode array detection, and monitored by MS. The relation
10 between the products generated electrochemically and enzymatically for the model
11 compound 2-OH-AC can provide a clue to the nature of their metabolic pathway initiation
12 (Volk *et al.* 1992). It opens up further perspectives directed to the search for more effective
13 and less toxic antitumor drugs among acridinone derivatives. It is significant because
14 improves the risk evaluation for potential drugs in the optimal chemotherapy schedules
15 designed for individual patients.
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

List of abbreviations:

CPE, controlled-potential electrolysis;

CV, cyclic voltammetry, cyclic voltammogram;

EC, electrochemistry, electrochemical cell;

ESI, electrospray ionization;

GC, glassy carbon;

GSH, glutathione (reduced form);

HLMs, human liver microsomes;

2-OH-AC, 2-hydroxyacridinone;

m/z, mass-to-charge ratio;

MS, mass spectrometry, mass spectrometer;

MS/MS, tandem mass spectrometry;

NADPH, β -nicotinamide adenine dinucleotide 2'-phosphate tetrasodium salt (reduced form);

NAC, *N*-acetylcysteine;

P450, cytochrome P450;

PBS, phosphate-buffered saline;

Q-TOF, quadrupole-time of flight;

RLMs, rat liver microsomes;

LC, liquid chromatography;

Parts of this work were presented at the 13th European ISSX Meeting (Glasgow, Scotland, 2015) and at the 2nd Congress BIO 2016 (Wrocław, Poland, 2016).

Materials and methods

Chemicals and enzymes

An acridinone derivative, a 2-hydroxyacridinone (2-OH-AC) was synthesized in our laboratory according to the method described earlier (Acheson 1973). The following chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA): *N*-acetylcysteine (NAC), dipotassium phosphate (K_2HPO_4), disodium phosphate (Na_2HPO_4), formic acid (HCOOH), L-glutathione reduced (GSH), monopotassium phosphate (KH_2PO_4), monosodium phosphate (NaH_2PO_4), and semicarbazide hydrochloride. Methanol (gradient grade for liquid chromatography) and β -nicotinamide adenine dinucleotide 2'-phosphate tetrasodium salt (β -NADPH) were obtained from Merck KGaA (Darmstadt, Germany). Ammonium formate ($HCOONH_4$) was ordered from Fisher Scientific (Loughborough, UK). Aluminum oxide (Al_2O_3) powder in form of 1- μ m alumina suspension, for polishing of electrodes, was delivered by TESTING Sp z o.o. (Katowice, Poland). All other commercially available chemicals and reagents were of the highest possible grade available. Ultrapure water ($0.056 \mu S \cdot cm^{-1}$), used in all the experiments, was passed through a Milli-Q water purification system from Merck KGaA (Darmstadt, Germany) or water distillation system from Hydrolab Sp. z o.o. sp.k. (Straszyn, Poland).

Pooled human liver microsomes (HLMs), mixed gender, from 50 donors (protein concentration, $20 \text{ mg} \cdot \text{mL}^{-1}$; P450 content, $411 \text{ pmol} \cdot \text{mg}^{-1}$ protein) and pooled rat liver microsomes (RLMs) from untreated, male Sprague-Dawley rats (protein concentration, $20 \text{ mg} \cdot \text{mL}^{-1}$; P450 content, $680 \text{ pmol} \cdot \text{mg}^{-1}$ protein) were purchased from Tebu-bio (Le Perray-En-Yvelines, France).

General instrumentation

Electrochemical measurements

Electrochemical measurements of the oxidation-reduction properties of 2-OH-AC were performed with controlled-potential electrolysis (CPE) and by electrochemical metabolism simulation in a three-electrode thin-layer cell (EC).

1
2
3 CPE was performed using an Autolab (Eco Chemie B.V., Utrecht, The Netherlands),
4 model PGSTAT 12 potentiostat, controlled via producer's software. The three-electrode
5 system, consisting of a cylinder glassy carbon (GC) working electrode ($\phi = 3$ mm and 13 mm
6 long; $A = 1.296$ cm²), an Ag/AgCl/3 M KCl reference electrode and a platinum-wire counter
7 electrode, were employed. During chronoamperometric measurements the working electrode
8 as well as the analyzed solution were placed in a glass tube closed with a "Vicor" plug to
9 separate the electrolyzed solution from the main solution.
10
11

12
13
14
15
16
17 Simulation of the oxidative metabolism reactions of 2-OH-AC was accomplished in an
18 electrochemical thin-layer cell equipped with a disc glassy carbon (GC) working electrode (ϕ
19 = 8 mm; $A = 0.502$ cm²) and a Pd/H₂ reference electrode (reactor cell; Antec Leyden,
20 Zoeterwoude, The Netherlands). Carbon-loaded PTFE (polytetrafluoroethylene) served as
21 auxiliary electrode. The cell potentials were applied using a ROXY EC System (Antec
22 Leyden) controlled by Antec Dialogue software (Antec Leyden). The outlet of the
23 electrochemical cell was connected directly to an electrospray ionization (ESI) source of
24 a quadrupole-time of flight (Q-TOF) mass spectrometer (Agilent Technologies, Santa Clara,
25 CA, USA) (Figure 1a). S-Conjugate formation of 2-OH-AC and GSH (NAC) was established
26 using a T-piece and a 100 μ L mixing coil placed between reactor cell and the ESI source
27 (Figure 1b). The relevant EC conditions can be viewed in Table 1.
28
29
30
31
32
33
34
35
36
37

38 **Liquid chromatographic analyses**

39
40 For all analyses of 2-OH-AC, liquid chromatographic (LC) separations were performed on
41 a reversed-phase 5- μ m Suplex pKb-100 analytical column (4.6 mm x 250 mm, C18)
42 (Supelco Inc., Bellefonte, PA, USA) with Waters Associates HPLC system (Waters Co.,
43 Milford, MA, USA). It was equipped with a model 600E system controller, a model 7725i
44 Rheodyne injector, and a model 2996 photodiode array detector (DAD) controlled with
45 Millennium software (Waters Co.). The LC analyses were carried out at a flow rate of 1
46 mL·min⁻¹ with the following mobile phase system: a linear gradient from 15% to 80%
47 methanol in 0.05 M aqueous ammonium formate buffer (pH 3.40 adjusted with formic acid)
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 for 25 min, followed by a linear gradient from 80% to 100% methanol in ammonium formate
4 buffer, pH 3.40, for 3 min. The eluates were monitored at 380 nm.
5

6 **Mass spectrometry**

7
8 Mass spectrometric detection, identification, and fragmentation of 2-OH-AC products
9 were carried out in the positive-ion mode by recording full scan spectra (m/z 100 – 600). To
10 ensure accurate mass during the experiment, the mass spectrometer was calibrated on a
11 daily basis, prior to sample analysis. The corresponding conditions for the ESI-Q-TOF-
12 MS(/MS) measurements are listed in Table 1.
13
14
15
16
17

18 **Controlled-potential electrolysis of 2-OH-AC**

19
20 The controlled-potential electrolysis was performed in 0.02 M phosphate-buffered saline
21 (PBS), pH 7.40, obtained by mixing Na_2HPO_4 and KH_2PO_4 , and adding the appropriate
22 amounts of NaCl and KCl to reach their concentrations of 0.15 and 0.002 M, respectively.
23 Each time before the electrolysis experiments the GC working electrode was briefly polished
24 with 1- μm Al_2O_3 powder on a wet pad. After polishing step, to remove alumina completely
25 from the surface, the electrode was rinsed with a direct stream of ultrapure water. To
26 eliminate the electric noise the electrochemical cell was placed in a Faraday cage. The
27 chronoamperometric experiments of 0.5 mM 2-OH-AC in the absence and the presence of
28 10-fold excess of GSH were performed at 0.50 V. The electrolysis progress was monitored
29 by voltammetric, spectroscopic, chromatographic, and spectrometric methods. The
30 experiments have been performed at room temperature (21 °C).
31
32
33
34
35
36
37
38
39
40
41

42 **Electrochemical simulation of the oxidative metabolism of 2-OH-AC**

43
44 For the electrochemical conversion, a 10- μM 2-OH-AC solution in electrolyte (0.1%
45 formic acid in water/methanol, 50/50, v/v) was pumped (SP2 – ROXY dual piston syringe
46 pump; Antec Leyden) through the electrochemical cell at a constant flow rate of 30 $\mu\text{L}\cdot\text{min}^{-1}$.
47 The potential of working electrode in the reactor cell was ramped between 0 and 2500 mV
48 (scan rate 10 $\text{mV}\cdot\text{s}^{-1}$). The outlet of the reactor cell was connected on-line to the ESI-MS
49 source (Figure 1a). To identify potential aldehyde products the oxidized sample was
50 collected in a vial containing semicarbazide (dissolved in ultrapure water) to give a final
51
52
53
54
55
56
57
58
59
60

1
2
3 concentration of 5 mM. After collection, the samples from the trapping experiments were kept
4
5 at 25 °C for 1 h before analysis by LC/MS.

6
7 The properties of the sample solvent may influence the conversion efficiency in the
8
9 electrochemical cell. Therefore, the electrochemical simulation of the oxidative metabolism of
10
11 2-OH-AC has been also performed in two different electrolyte solutions containing
12
13 acetonitrile (0.1% formic acid or 20 mM ammonium formate (pH 3.40), respectively, in
14
15 water/acetonitrile, 50/50, v/v).

16 ***On-line trapping of oxidation product with glutathione (N-acetylcysteine)***

17
18 To identify possible S-conjugate(s) with GSH (NAC), the EC/MS set-up described above
19
20 was slightly modified (Figure 1b). A 10- μ M 2-OH-AC solution in electrolyte was pumped
21
22 through the electrochemical cell at a constant flow rate of 30 μ L \cdot min⁻¹. 100 μ M GSH (NAC) in
23
24 ultrapure water was added at the same flow rate to the effluent of the electrochemical cell via
25
26 a T-piece into a 100 μ L mixing coil. The effluent from the mixing coil was injected directly into
27
28 the ESI-MS interface.

29 ***In vitro microsomal incubations***

30
31
32 Human or rat liver microsomal incubations (1 mg \cdot mL⁻¹ protein) were performed with 50
33
34 μ M 2-OH-AC, 1 mM NADPH, and 1 mM GSH in a potassium phosphate-buffered solution
35
36 (0.1 M, pH 7.40) at 37 °C for 1 h, in a total volume of 100 μ L. The incubations were
37
38 terminated by adding ice-cold methanol to the incubation mixtures (50:50, v/v). The samples
39
40 were then vortexed, placed in ice for 10 min, and centrifuged at 10000 x g for 10 min.
41
42 Aliquots of the supernatants (150 μ L) were then analyzed directly by reversed-phase LC with
43
44 UV-Vis detection and/or diode array and multiple wavelength detection, and monitored by
45
46 MS. The three types of control incubations were applied, the first without test compound, the
47
48 second without NADPH, and the third without GSH.

Results

In order to provide an important insight into the mechanism of antitumor action of imidazo- and triazoloacridinones, the electrochemical oxidation of their simpler reference compound,

2-OH-AC, was investigated. The ability of 2-OH-AC to undergo oxidative metabolic activation was studied by applying the direct combination of electrochemistry and mass spectrometry (EC/MS). For 2-OH-AC no EC/MS data were found before. Electrochemical conversion of 2-OH-AC was conducted in parallel to controlled-potential electrolysis (CPE) and *in vitro* experiments using liver microsomes.

Controlled-potential electrolysis of 2-OH-AC

At first, cyclic voltammetry (CV) was used to initially investigate the electrochemistry of 2-OH-AC at a glassy carbon electrode in a range of positive potentials and to determine the optimum operating range of voltages for further EC/MS studies. In order to confirm the postulated oxidation mechanism of 2-OH-AC action and to identify the oxidation products, controlled-potential electrolysis of 2-OH-AC (0.5 mM in 0.02 M PBS buffer, pH 7.40) was performed at 0.50 V. To support the possibility of the formation of the stable adduct between 2-OH-AC intermediate and GSH, the tests were performed in the absence and the presence of GSH (5 mM in ultrapure water). The electrolysis measurements were carried out for 24 hours and the progress of the process was monitored periodically by application of voltammetric, spectroscopic, chromatographic, and spectrometric methods.

Data for the CV are provided in the supplemental material. In the solution of pure 2-OH-AC one anodic peak (a1) and three cathodic peaks (c1, c2, c3) were obtained. The appearance of three cathodic peaks indicated that the oxidation product of 2-OH-AC underwent subsequent chemical reactions. The addition of 10-fold excess of GSH to 2-OH-AC solution resulted in a shift of the oxidation peak of 2-OH-AC of about 30 mV to a more positive potential. In turn, the signal appearing in the cathodic part of the CV at circa 0.32 V virtually disappeared and two other cathodic peaks were substantially depressed. These

1
2
3 changes confirmed the interaction between 2-OH-AC and GSH, and the reducing effect of
4
5 GSH against the oxidation reaction products.

6
7 Assuming that during oxidation process of 2-OH-AC two electrons are exchanged, the
8
9 total electrolysis of 0.5 mM compound should be achieved at the time of obtaining the charge
10
11 of 200 mC. Unfortunately, despite long electrolysis (over 1 day) the maximum charge which
12
13 was achieved was only 29 mC. A possible explanation of this situation is that the oxidation
14
15 products of 2-OH-AC strongly adsorbed on the electrode surface and effectively blocked it.
16
17 With electrolysis progress the intensity of the current signals drastically decreased (Figure
18
19 2a). In addition, a new oxidation signal at circa 0.06 V (a2) was also well visible. It was
20
21 related to the third cathodic peak (c1). The introduction of GSH to the solution of 2-OH-AC
22
23 significantly minimized the effects of the electrolysis process. This observation may point out
24
25 the presence of interactions between oxidation products of 2-OH-AC and GSH. The CPE of a
26
27 mixture of 2-OH-AC and GSH (Figure 2b) revealed the anodic signal at circa 0.42 V (a1) with
28
29 increasing intensity. Also a new signal emerged at circa 0.31 V (a2) and its height increased
30
31 with time of electrolysis. This product peak was due to the reduction of the transformed 2-
32
33 OH-AC molecule.

34 ***Electrochemical simulation of the oxidative metabolism of 2-OH-AC (phase I*** 35 ***metabolism)*** 36 37

38
39 The electrochemical oxidation products of 2-OH-AC were generated and identified using
40
41 on-line EC/MS (Figure 1a). The applied set up enabled us to simulate metabolic P450-
42
43 catalyzed reactions occurring in the liver. To reduce the complexity of the system these
44
45 experiments were carried out only with flow injection EC/MS without using an LC column.

46
47 The best electrochemical conversion of 2-OH-AC into its expected products was
48
49 achieved using the scan mode in the potential range of 0 – 2.5 V. To provide a concise
50
51 overview of the oxidation products, 2-D mass voltammograms were generated by plotting the
52
53 extracted ion intensities versus the progress of the electrochemical oxidation. Mass
54
55 spectrometry allowed the identification of the formed products by an increasing signal
56
57 intensity of the corresponding mass-to-charge (m/z) ratio. Furthermore, for structure
58
59
60

1
2
3 elucidation of the detected products, fragment ions, generated by in-source fragmentation in
4 the ESI interface, have been studied.
5

6
7 A representative 2-D mass voltammogram resulting from the oxidation of 2-OH-AC within
8 the scanned range is shown in Figure 3. The 2-OH-AC molecule was easily protonated and
9 detected with high intensities as $[M+H]^+$ ion (m/z 212) in the positive ionization mode of the
10 ESI-Q-TOF mass spectrometer. No products were observed in the solution of pure 2-OH-AC
11 without any electrochemical potential (cell off) (the inset of top left corner in Figure 3),
12 whereas significant drop in an extracted ion intensity of 2-OH-AC was noticed when the
13 potential was applied (cell on) (the inset of top right corner in Figure 3). This change is
14 attributed to the oxidation of 2-OH-AC into eight products in an electrochemical thin-layer
15 cell. All products are summarized in Table 2 and will be discussed in detail, with specific
16 reference to their accurate mass data and fragmentation patterns.
17
18
19
20
21
22
23
24
25

26
27 Furthermore, we noticed that the ratios of oxidation products of 2-OH-AC were strongly
28 dependent on the solvent composition. Most products identified using methanol-containing
29 electrolyte, with the exception of P4 (m/z 240) and P8 (m/z 421), have been also observed in
30 solutions of acetonitrile. No additional oxidation products of 2-OH-AC were detected under
31 these conditions. Higher signal intensity of the selected mass ions was caused by that
32 methanol was better than acetonitrile in diminishing the adsorption of the electrochemical
33 products on the surface of the working electrode. Moreover, methanol/water electrolyte
34 solution produced the lowest mass background noise in positive total ion chromatogram
35 (data not shown). A summary of the electrochemical products observed under different
36 electrolyte conditions is available in the supplemental material.
37
38
39
40
41
42
43
44
45

46 ***Characterization of GSH conjugate formed from electrochemical oxidation of 2-OH-AC*** 47 48 ***(phase II metabolism)*** 49

50
51 In this work, the EC/MS set-up described above was extended to a system allowing the
52 study of the conjugation of 2-OH-AC oxidation products with a reactive endogenous
53 nucleophile, such as reduced GSH or NAC. GSH and NAC were selected because of a
54 simple structure with their soft nucleophilic thiol group and their relevance in living organisms
55
56
57
58
59
60

1
2
3 (Evans *et al.* 2004). The instrumental set-up for electrochemical simulation of conjugation
4
5 reactions (phase II reactions) is shown in Figure 1b.

6
7 Observation of the potential conjugation reaction(s) between the oxidation product(s) of
8
9 2-OH-AC and GSH was possible in the form of a 2-D mass voltammogram (Figure 4a).

10
11 The mass spectra were acquired to confirm the formation of GSH S-conjugate (Figure 5).
12
13 As expected, no additional signals besides those associated with protonated 2-OH-AC (m/z
14
15 212) and GSH (m/z 308) were observed at cell off (Figure 5a and a zoom in the mass
16
17 spectrum from the circle range). However, in the potential range of 0 – 2.5 V (cell on) mass
18
19 spectrum revealed one m/z signal representing potential GSH S-conjugate (Figure 5b and a
20
21 zoom in the mass spectrum from the circle range). Ion with a m/z ratio of 517 was observed
22
23 with a weak intensity which was increasing only in the case when an extracted ion intensity
24
25 of protonated 2-OH-AC was decreasing. It was confirmed by the accurate mass
26
27 measurements that m/z 517 represents the conjugate of molecular formula $C_{23}H_{24}N_4O_8S$,
28
29 consistent with a product of the 2-OH-AC oxidation and one molecule of GSH. This result is
30
31 in accordance with the assumption that GSH traps soft electrophiles with its thiol group
32
33 (Inoue *et al.* 2015).

34
35 To confirm that the ion at m/z 517 is originating from GSH conjugate of 2-OH-AC, the
36
37 fragmentation spectrum was recorded (Figure 5c). The MS/MS of m/z 517 ion produced two
38
39 fragment ions at m/z 388 and 244, respectively. The accurate values of m/z correlated well
40
41 with calculated m/z (Table 3). The first was consistent with the neutral loss of
42
43 anhydroglutamic acid (-129), whereas the second (-275) was assigned as a cleavage
44
45 adjacent to the cysteinyl thioether moiety with charge retention on the 2-OH-AC molecule.
46
47 Thus, the fragmentation pattern of m/z 517 ion suggested the presence of at least one GSH
48
49 moiety. As for GSH, formation of NAC S-conjugate with a m/z ratio of 373 was also observed
50
51 in similar experiments (Figure 4b). The MS/MS of 373 ion revealed the presence of the
52
53 fragment ion at m/z 244 (data not shown), exactly the same that occurred in MS/MS of m/z
54
55 517 ion.
56
57
58
59
60

1
2
3 **Identification of GSH conjugate of 2-OH-AC from *in vitro* microsomal incubations and**
4 **controlled-potential electrolysis**
5

6 Trapping experiments with GSH in *in vitro* microsomal incubations and during controlled-
7 potential electrolysis of 2-OH-AC were carried out to generally assess the feasibility of GSH
8 S-conjugate formation. The results obtained from these approaches were compared with
9 those from a purely instrumental electrochemical simulation. Figure 6a presents the
10 representative high performance LC chromatograms recorded for enzymatic and
11 electrochemical oxidation of 2-OH-AC in the absence and the presence of 10-fold excess of
12 GSH.
13
14
15
16
17
18
19

20 Due to various expressions of the different isoforms of P450 in each organism, the
21 metabolism in human and rat liver microsomes may result in different metabolites or in a
22 different quantitative distribution of the metabolites (Martignoni *et al.* 2006). When
23 experiments were performed using both types of microsomal fractions, in the case of 2-OH-
24 AC we observed only slight differences in the intensity of the individual peaks, hence the
25 further discussion will be based on the results from RLM studies. The microsomal
26 incubations included three controls, one without 2-OH-AC to rule out any potential
27 interference/contamination from endogenous compounds, the second without NADPH, a
28 cofactor for cytochrome P450 activities, to assess the metabolic dependence of GSH
29 conjugate formation, and the third without GSH. No conjugate was detected in the “no
30 compound’ and “no NADPH” control reactions (data shown in the supplemental material). LC
31 analysis of microsomal samples without GSH revealed that three main metabolites were
32 formed under the conditions studied (Figure 6a). Chromatogram taken after 60 min
33 incubation of 2-OH-AC with 10-fold excess of GSH represents one additional
34 chromatographic peak. It is noteworthy that it was not seen in the control incubations without
35 NADPH, which suggests an NADPH-dependent oxidation of 2-OH-AC to this metabolite. LC
36 analysis of the reaction mixture throughout the course of controlled-potential electrolysis
37 revealed one main product (Figure 6a). Compared to enzymatically generated metabolites,
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 two of them were missing. As before, one additional peak was observed only where GSH
4 was present in excess, and its intensity depended on the progress of electrolysis.
5

6
7 In both approaches the UV-Vis spectra of the peaks at 18.6 min differed significantly from
8 that of the substrate (the inset in Figure 6a). The shift towards longer wavelengths may
9 indicate that a more extensive delocalized electron system exists in this product in
10 comparison to that in 2-OH-AC. It was subsequently identified by ESI-MS as GSH S-
11 conjugate with the mass ion at m/z 517 (data not shown). Therefore, the above results
12 agreed with the data obtained from EC/MS or EC/MS/MS measurements and confirmed the
13 existence of a reactive intermediate in the oxidative pathway of 2-OH-AC. However, due to
14 the low concentration of the S-conjugate in the EC/MS system it was not possible to obtain
15 its chromatographic peak and UV-Vis spectrum. Also other products showing quite good
16 intensities in the EC/MS system, were found in small or trace amounts in LC analysis (Figure
17 6b). Some of them were not detected, which is likely to be a consequence of their low
18 stability. Nevertheless, it is worth to note that generally the EC system allowed to obtain a
19 wider set of phase I metabolites than the use of classical approaches, probably because of
20 clean
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
clean matrix.

Discussion

The toxic effects of drugs and other xenobiotics arise not only from the compound itself but also from its metabolites (Baumann *et al.* 2010). Hence, during drug development process particular attention is paid to drug metabolite formation and identification of metabolic pathways. Progress in this research area depends critically on the improvement of methods involved in the generation and analysis of various types of drug metabolites, with special respect to the characterization of reactive metabolites (Prakash *et al.* 2008). In this work different approaches for the investigation of the 2-OH-AC oxidative metabolism, that may occurs in the living organism, were performed. The oxidative electrochemical behavior of 2-OH-AC was first investigated by the direct combination of electrochemistry and mass spectrometry. The applicability of the electrochemistry in drug metabolite synthesis was evaluated by conventional electrochemical (controlled-potential electrolysis) and enzymatic (microsomal incubation) approaches.

The electrochemical conversion of 2-OH-AC in an electrochemical thin-layer cell was successfully achieved (Figure 3). Table 2 consists of list of major products related to 2-OH-AC oxidative metabolism and m/z ratios of the protonated species $[M+H]^+$ used for mass spectra interpretation. The resulting molecular formulas were deduced by their accurate masses. The isotopic patterns correlated well with theoretical calculations. For all compounds, the deviations between the calculated and measured m/z values were less than 4 ppm.

The proposed chemical reactions that may occur in the electrochemical cell are summarized in Figure 7. Most of 2-OH-AC oxidation products seem to be species containing additional oxygen atom and can exist in at least two tautomeric forms. According to previous studies on the oxidative transformations of 2-OH-AC (Mazerska *et al.* 2002), the most likely oxidation site in the 2-OH-AC molecule is the position *ortho* to the hydroxyl group. This relates to the positions 1 or 3 of 2-OH-AC, wherein position 1 remains preferential. Thus, the mass ion at m/z 226 is proposed to correspond to 1,2-orthoquinone (P1a), while the signal at m/z 228 may include monohydroxylation product (P3). The molecular ion observed with little

1
2
3 signal intensity at m/z 421 was identified to be the dimer formed from two substrate
4 molecules of radical structure linked together at position 1 of 2-OH-AC (P8). The chemical
5 structures of these three products have been previously determined by means of MS and
6 NMR spectroscopy (Mazerska *et al.* 2002).
7
8
9

10 Other products of oxidative metabolism of 2-OH-AC have been identified for the first time
11 upon electrochemical oxidation. The P2 compound with the mass ion at m/z 227 may have
12 been formed via C- or N-hydroxylation because the mass corresponds to an addition of
13 15.99 Da to the parent compound with a lack of one hydrogen atom. As a result, the P2
14 molecule should contain a positive charge, probably located on the nitrogen atom of the
15 acridinone ring. Further, considering the mesomeric effects in electron density distribution,
16 the loss of one hydrogen atom in P2 compound would indicate the possibility of the formation
17 other than P1a structures for the mass ion at m/z 226, including P1b.
18
19
20
21
22
23
24
25

26 The products showing the mass ions at m/z 240 (P4), 242 (P5), 256 (P6), and 258 (P7)
27 have not been definitively identified. However, we propose here only their tentative
28 structures. The accurate mass of ion at m/z 242 (P5) differed from the accurate mass of ion
29 at m/z 226 (P1) by exactly 16.03 Da. This value may indicate the presence of the additional
30 one carbon and four hydrogen atoms, but not a single oxygen atom (15.99 Da), somewhere
31 within the P5 molecule. During the electrochemical and/or the ionization processes some
32 side reactions can take place, like the reaction of electrolyte components with
33 electrochemically generated intermediates and sometimes they are unexpected. Based on
34 the literature data (Sichilongo *et al.* 2011; Wang *et al.* 2011), electro-oxidation of methanol
35 from electrolyte may give a strongly electrophilic methylium (CH_3^+) or methyl radical ($\cdot\text{CH}_3$).
36 Thus, we can assume that it is quite likely that compound containing a methyl group in its
37 structure was formed. The probable methylation of 2-OH-AC derivative have not been
38 avoided even when acetonitrile was present in the electrolyte solution (supplemental
39 material). The possible location of attachment is speculated to be an electronegative nitrogen
40 atom due to valence considerations. Presumably further two-electron two-proton
41 dehydrogenation of P5 provided P4 (m/z 240), with molecular mass ion decreased by 2.01
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 Da. In turn, a product with the mass ion at m/z 258 (P7) could have been created by the
4 hydroxylation of the methyl group of P5, and its further two-electron two-proton
5 dehydrogenation gave a signal at m/z 256 (P6).
6
7

8
9 It should be pointed out that the signals at m/z 240 and m/z 256 may correspond to
10 aldehyde products as well. To test whether the hydroxymethyl group at the nitrogen atom in
11 P5 or in P7 products underwent further oxidation, trapping experiments were performed with
12 semicarbazide. It is a small molecule trapping agent that can form a Schiff base with
13 aldehydes in the process mimicking reactions between aldehyde metabolites with lysine
14 residues on proteins (Evans *et al.* 2004). Semicarbazide was added to the electrochemical
15 oxidation mixture of 2-OH-AC followed by LC/MS analysis. Trapping with semicarbazide did
16 not give any conjugates what excluded the presence of aldehyde products in oxidative
17 transformations of 2-OH-AC.
18
19
20
21
22
23
24
25

26 To simulate the phase II metabolism of 2-OH-AC, electrochemical oxidation of the
27 compound was carried out in the presence of an excess amount of nucleophilic trapping
28 agent. EC/MS successfully predicted the formation of chemically reactive metabolite that
29 spontaneously reacted with reduced glutathione (Figures 4a and 5) and/or *N*-acetylcysteine
30 (Figure 4b) to form conjugates (m/z 517 and m/z 373, respectively) via the thiol group.
31 General structures for probable GSH and NAC conjugates of 2-OH-AC are shown in Figure
32 8. We suspect that 2-OH-AC is likely to undergo a P450-catalyzed oxidative dehydrogenation
33 which results in the formation of a reactive intermediate in the form of a quinone imine
34 (Guengerich 2007), as, *e.g.*, it takes place in a metabolic pathway of the acetaminophen
35 (Jollow *et al.* 1973; Larson 2007). For this mechanism, a quinone imine of 2-OH-AC has to
36 be the precursor of the adduct as the corresponding quinone imine would have the m/z ratio
37 of 210 in the positive-ion mode detection. However, this intermediate product, presumably
38 due to its short life span and high reactivity, was not directly detected in the mass
39 voltammogram of pure 2-OH-AC. It is important to note that the same type of GSH S-
40 conjugate was detected after *in vitro* liver microsomal incubations in an NADPH-dependent
41 manner and in the reaction mixture from the electrolysis experiment (Figure 6). Quinone
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 imines are well known reactive intermediates which very often undergo adduct formation with
4 crucial cellular compounds such as GSH, structural proteins, enzymes and/or DNA (Bolton *et*
5 *al.* 2000; Zhou *et al.* 2005). As a consequence, these compounds can cause a variety of
6 hazardous effects *in vivo*, including acute cytotoxicity, immunotoxicity, and carcinogenesis,
7 especially in the case of depleted levels of cellular GSH (Lohmann *et al.* 2010). However, the
8 fact that reactive metabolite of 2-OH-AC may exist and, as it has been shown, may undergo
9 adduct formation does not consequently imply that it causes toxicity (Evans *et al.* 2004) so
10 further studies are needed to elucidate whether this metabolite contributes to the 2-OH-AC
11 toxicity *in vivo*.
12
13
14
15
16
17
18
19

20 The results of our studies clearly prove that EC-based approach has the potential to
21 simulate the majority of oxidative metabolism reaction, including the simulation of reactive
22 metabolite formation and its binding to biomolecules (e.g., GSH). The observed
23 dehydrogenation, hydroxylation and oxidation of acridinone molecule are in general
24 agreement with reports on the types of P450-catalyzed reactions being simulated by
25 electrochemistry (Lohmann & Karst 2008; Lohmann *et al.* 2010). Our observations on
26 product structures may be advantageous from the viewpoint of the structural factors that
27 influence the reactivity and functional group interactions of the compound, providing a good
28 approximation of what may occur *in vivo*.
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Conclusions

The present study demonstrated the significance of electrochemistry coupled on-line with mass spectrometry in drug metabolism studies. This combination appeared to be a suitable tool for a simulation of some types of oxidative drug metabolic reactions related to cytochromes P450, and for studying the formation of reactive metabolites. In this investigation, 2-hydroxyacridinone (2-OH-AC), the reference compound for antitumor-active imidazo- and triazoloacridinone derivatives, was easily oxidized in an electrochemical thin-layer cell to several different products. To trap potentially reactive metabolite(s) two types of nucleophilic trapping agents, reduced GSH and NAC, both with nucleophilic thiol group, were used. This allowed us to identify and characterize a novel GSH (NAC) S-conjugate of 2-OH-AC. The postulated reactive quinone imine metabolite of 2-OH-AC may potentially be involved in a number of biochemical transformations, and can be responsible for the antitumor activity of acridinone derivatives. The role of this reactive metabolite requires further investigation.

The electrochemical method we proposed here represented a good alternative for classical metabolic studies with the principal advantage which is the absence of proteins in the reaction medium. It will be very useful in further studies on metabolic transformations of antitumor imidazo- and triazoloacridinone drugs. Considering pharmacological aspect, our findings may provide important guidelines for the further modification of the acridinone compounds so they contribute to the design and the development of safer and more effective therapeutic agents.

1
2
3 **Acknowledgments**

4 This work was supported by the National Science Center (Poland) under Grant number
5
6 2012/07/D/NZ7/03395..
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

For Peer Review Only

1
2
3 **Declaration of interest statement**

4 The authors confirm that this article content has no conflicts of interest.
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

For Peer Review Only

References

Acheson RM. (1973). Acridines. In: Acheson RM, ed. The chemistry of heterocyclic compounds. New York, Interscience Publishers, Inc., p. 141.

Augustin E, Moś-Rompa A, Skwarska A, Witkowski JM, Konopa J. (2006). Induction of G2/M phase arrest and apoptosis of human leukemia cells by potent antitumor triazoloacridinone C-1305. *Biochem Pharmacol* 72: 1668-1679.

Baillie TA, Cayen MN, Fouda H, Gerson RJ, Green JD, Grossman SJ, *et al.* (2002). Drug metabolites in safety testing. *Toxicol Appl Pharmacol* 182: 188-196.

Baumann A, Lohmann W, Rose T, Ahn KC, Hammock BD, Karst U, *et al.* (2010). Electrochemistry-mass spectrometry unveils the formation of reactive triclocarban metabolites. *Drug Metab Dispos* 38: 2130-2138.

Bolton JL, Trush MA, Penning TM, Dryhurst G, Monks TJ. (2000). Role of quinones in toxicology. *Chem Res Toxicol* 13: 135-160.

Brandon EF, Raap CD, Meijerman I, Beijnen JH, Schellens JH. (2003). An update on in vitro test methods in human hepatic drug biotransformation research: pros and cons. *Toxicol Appl Pharmacol* 189: 233-246.

Bussy U, Boisseau R, Thobie-Gautier C, Boujtita M. (2015). Electrochemistry-mass spectrometry to study reactive drug metabolites and CYP450 simulations. *Trends Anal Chem* 70: 67-73.

Bussy U, Boujtita M. (2014). Advances in the electrochemical simulation of oxidation reactions mediated by cytochrome P450. *Chem Res Toxicol* 27: 1652-1668.

1
2
3 Cholody WM, Horowska B, Paradziej-Lukowicz J, Martelli S, Konopa J. 1996. Structure-
4 activity relationship for antineoplastic imidazoacridinones: synthesis and antileukemic activity
5 in vivo. *J Med Chem* 39: 1028-1032.
6
7

8
9
10 Cholody WM, Martelli S, Konopa J. (1990). 8-Substitued 5-[(aminoalkyl)amino]-6H-v-
11 triazolo[4,5,1-de]acridin-6-ones as potential antineoplastic agents. Synthesis and biological
12 activity. *J Med Chem* 33: 2852-2856.
13
14

15
16
17 Cholody WM, Martelli S, Lukowicz J, Konopa J. (1990). 5-[(Aminoalkyl)amino]imidazo[4,5,1-
18 de]acridin-6-ones as a novel class of antineoplastic agents. Synthesis and biological activity.
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

J Med Chem 33: 49-52.

Dziegielewski J, Konopa J. (1996). Interstrand crosslinking of DNA induced in tumor cells by
a new group of antitumor imidazoacridinones. *Proc Am Assoc Cancer Res* 37: 410.

Dziegielewski J, Konopa J. (1998). Characterisation of covalent binding to DNA of antitumor
imidazoacridinone C-1311, after metabolic activation. *Ann Oncol Suppl* 1: 137.

Dziegielewski J, Slusarski B, Konitz A, Skladanowski A, Konopa J. (2002). Intercalation of
imidazoacridinones to DNA and its relevance to cytotoxic and antitumor activity. *Biochem
Pharmacol* 63: 1653-1662.

Evans DC, Watt AP, Nicoll-Griffith DA, Baillie TA. (2004). Drug-protein adducts: An industry
perspective on minimizing the potential for drug bioactivation in drug discovery and
development. *Chem Res Toxicol* 17: 3-16.

1
2
3 Faber H, Jahn S, Kunнемeyer J, Simon H, Melles D, Vogel M, *et al.* (2011).
4 Electrochemistry/liquid chromatography/mass spectrometry as a tool in metabolism studies.
5 *Angew Chem* 50: A52-58.
6
7

8
9
10 Guengerich FP. (2007). Oxidative, reductive and hydrolytic metabolism of drugs. In: Zhang
11 D, Zhu M, Humphreys WG, eds. *Drug metabolism in drug design and development*. New
12 York, John Willey & Sons, Inc., p. 265.
13
14

15
16
17
18 Harding MM, Grummitt AR. (2003). 9-hydroxyellipticine and derivatives as chemotherapy
19 agents. *Mini Rev Med Chem* 3: 67-76.
20
21

22
23
24 Inoue K, Fukuda K, Yoshimura T, Kusano K. (2015). Comparison of the reactivity of trapping
25 reagents toward electrophiles: cysteine derivatives can be bifunctional trapping reagents.
26 *Chem Res Toxicol* 28: 1546-1555.
27
28

29
30
31
32 Jennings GS, Strauss M. (1999). Immortalization of hepatocytes through targeted
33 deregulation of the cell cycle. In: Al-Rubeai M, ed. *Cell engineering*. the Netherlands, Kluwer
34 Academic Publishers, p. 259.
35
36

37
38
39
40 Jollow DJ, Mitchell JR, Potter WZ, Davis DC, Gillette JR, Brodie BB. (1973). Acetaminophen-
41 induced hepatic necrosis. II. Role of covalent binding in vivo. *J Pharmacol Exp Ther* 187:
42 195-202.
43
44

45
46
47
48 Jurva U, Wikstrom HV, Weidolf L, Bruins AP. (2003). Comparison between
49 electrochemistry/mass spectrometry and cytochrome P450 catalyzed oxidation reactions.
50 *Rapid Commun Mass Spectrom* 17: 800-810.
51
52

1
2
3 Kalgutkar AS, Soglia JR. (2005). Minimising the potential for metabolic activation in drug
4 discovery. *Expert Opin Drug Metab Toxicol* 1: 91-142.

6
7
8
9 Koba M, Konopa J. (2007). Interactions of antitumor triazoloacridinones with DNA,. *Acta*
10 *Biochim Pol* 54: 297-306.

12
13
14 Kusnierczyk H, Cholody WM, Paradziej-Lukowicz J, Radzikowski C, Konopa J. (1994).
15 Experimental antitumor activity and toxicity of the selected triazolo- and imidazoacridinones.
16 *Arch Immunol Ther Exper (Warsz)* 42: 415-423.

18
19
20 Larson AM. (2007). Acetaminophen hepatotoxicity. *Clin Liver Dis* 11: 525-548.

22
23
24
25
26 Lemke K, Wojciechowski M, Laine W, Bailly C, Colson P, Baginski M, *et al.* (2005). Induction
27 of unique structural changes in guanine-rich DNA regions by the triazoloacridone C-1305, a
28 topoisomerase II inhibitor with antitumor activities. *Nucleic Acids Res* 33: 6034-6047.

30
31
32
33
34 Lohmann W, Karst U. (2008). Biomimetic modeling of oxidative drug metabolism: strategies,
35 advantages and limitations. *Anal Bioanal Chem* 391: 79-96.

36
37
38
39
40 Lohmann W, Baumann A, Karst U. (2010). Electrochemistry and LC-MS for metabolite
41 generation and identification: tools, technologies and trends. *LC•GC Europe* January: 1-7.

42
43
44
45
46 Marks TA, Venditti JM. (1976). Potentiation of actinomycin D or adriamycin antitumor activity
47 with DNA. *Cancer Research* 36: 496-504.

48
49
50
51
52 Martignoni M, Groothuis GM, de Kanter R. (2006). Species differences between mouse, rat,
53 dog, monkey and human CYP-mediated drug metabolism, inhibition and induction. *Expert*
54 *Opin Drug Metab Toxicol* 2: 875-894.

1
2
3 Mazerska Z, Dziegielewski J, Konopa J. (2001). Enzymatic activation of a new antitumour
4 drug 5-diethylaminoethylamino-8-hydroxyimidazo-acridinine, C-1311, observed after its
5 intercalation into DNA. *Biochem Pharmacol* 61: 685-694.
6
7

8
9
10 Mazerska Z, Zamponi S, Marassi R, Martelli S, Konopa J. (1997). Electrochemical oxidation
11 of antitumor imidazoacridinone derivatives and the reference 2-hydroxyacridinone. *J.*
12 *Electroanal Chem* 427: 71-78.
13
14

15
16
17
18 Mazerska Z, Zamponi S, Marassi R, Sowinski P, Konopa J. (2002). The products of electro-
19 and photochemical oxidation of 2-hydroxyacridinone, the reference compound of antitumor
20 imidazoacridinone derivatives. *J Electroanal Chem* 521: 144-154.
21
22
23

24
25
26 Nouri-Nigjeh E, Bischoff R, Bruins AP, Permentier HP. (2011). Electrochemistry in the
27 mimicry of oxidative drug metabolism by cytochrome P450s. *Curr Drug Metab* 12: 359-371.
28
29

30
31
32 Orhan H, Vermeulen NPE. (2011). Conventional and novel approaches in generating and
33 characterization of reactive intermediates from drugs/drug candidates. *Curr Drug Metab* 12:
34 383-394.
35
36
37

38
39
40 Park BK, Boobis A, Clarke S, Goldring CE, Jones D, Kenna JG, *et al.* (2011). Managing the
41 challenge of chemically reactive metabolites in drug development. *Nat Rev Drug Discov* 10:
42 292-306.
43
44
45

46
47
48 Permentier HP, Bruins AP, Bischoff R. (2008). Electrochemistry-mass spectrometry in drug
49 metabolism and protein research. *Mini Rev Med Chem* 8: 45-56.
50
51

52
53
54 Pinto N, Dolan ME. (2011). Clinically relevant genetic variations in drug metabolizing
55 enzymes. *Curr Drug Metab* 12: 487-497.
56
57

1
2
3 Prakash C, Sharma R, Gleave M, Nedderman A. (2008). In vitro screening techniques for
4 reactive metabolites for minimizing bioactivation potential in drug discovery. *Curr Drug Metab*
5 9: 952-964.
6
7

8
9
10 Sichilongo KF, Famuyiwa SO, R. Kibechu R. (2011). Pre-electrospray ionisation manifold
11 methylation and post-electrospray ionisation manifold cleavage/ion cluster formation
12 observed during electrospray ionisation of chloramphenicol in solutions of methanol and
13 acetonitrile for liquid chromatography-mass spectrometry employing a commercial
14 quadrupole ion trap mass analyser. *Eur J Mass Spectrom (Chichester)* 17: 255-264.
15
16
17
18

19
20
21
22 Skladanowski A, Plisov SY, Konopa J, Larsen AK. (1996). Inhibition of DNA topoisomerase II
23 by imidazoacridinones, new antineoplastic agents with strong activity against solid tumors.
24
25
26 *Mol Pharmacol* 49: 772-780.
27

28
29
30 Srivastava A, Maggs JL, Antoine DJ, Williams DP, Smith DA, Park BK. (2010). Role of
31 reactive metabolites in drug-induced hepatotoxicity. *Handb Exp Pharmacol* 196 (2010) 165-
32 194.
33
34
35

36
37
38 Volk KJ, Yost RA, Brajter-Toth A. (1992). Electrochemistry on line with mass spectrometry
39 insight into biological redox reactions. *Anal Chem* 64: 21A-26A.
40
41
42

43
44 Wang L, Chai Y, Tu P, Sun C, Pan Y. (2011). Formation of $[M + 15]^+$ ions from aromatic
45 aldehydes by use of methanol: in-source aldolization reaction in electrospray ionization mass
46 spectrometry. *J Mass Spectrom* 46: 1203-1210.
47
48
49

50
51
52 Zanger UM, Schwab M. (2013). Cytochrome P450 enzymes in drug metabolism: Regulation
53 of gene expression, enzyme activities, and impact of genetic variation. *Pharmacol Therapeut*
54 138: 103-141.
55
56
57

1
2
3
4 Zhou S, Chan E, Duan W, Huang M, Chen YZ. (2005). Drug bioactivation, covalent binding
5 to target proteins and toxicity relevance. Drug Metab Rev 37: 41-213.
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

For Peer Review Only

1
2
3 **Tables with captions**

4 **Table 1**

5 EC and ESI-Q-TOF-MS(/MS) parameters as applied in direct EC/MS(/MS) and in MS(/MS)
6
7 experiments for determination of accurate masses of product ions and getting ion
8
9 fragmentation.
10
11

	Parameter	Value or setting
EC settings	Flow rate	30 $\mu\text{L}\cdot\text{min}^{-1}$
	Potential	0 – 2.5 V (10 mV steps)
	EC operating mode	scan
	Cycle	continuous
MS(/MS) settings	The range of m/z	100 – 600
	Ion source	dual ESI
	Ion polarity	positive
	MS operating mode	scan
	Capillary voltage	3500 V
	Nebulizer gas (N_2) pressure	35 psig
	Drying gas (N_2) flow	10 $\text{L}\cdot\text{min}^{-1}$
	Drying gas temperature	325 $^{\circ}\text{C}$
	Fragmentor	175 V
	Skimmer	45 V
	OCT 1 RF V_{pp}	750 V
	Rate	1,5 $\text{spectra}\cdot\text{s}^{-1}$
	MS/MS method	targeted
- Slope	- 4 m/z	
- Offset	- 5 V	

Table 2

A summary of molecular formulas of 2-OH-AC and its products found after electrochemical oxidation on a GC working electrode in a potential range of 0 – 2.5 V versus Pd/H₂. Base

Product	Measured <i>m/z</i> / Da	Calculated <i>m/z</i> / Da ^a	Deviation / ppm	Molecular formula of [M+H] ⁺ ion	MS/MS fragment ions	Predicted modification of 2-OH-AC molecule
2-OH-AC	212.0712	212.0706	2.9	C ₁₃ H ₁₀ NO ₂	-	-
P1a, b	226.0501	226.0499	1.1	C ₁₃ H ₈ NO ₃	212	- 2H + O
P2	227.0578	227.0577	0.5	C ₁₃ H ₉ NO ₃	212	- H + O
P3	228.0648	228.0655	-3.1	C ₁₃ H ₁₀ NO ₃	212, 226, 227	+ O
P4	240.0658	240.0655	1.2	C ₁₄ H ₁₀ NO ₃	212, 227 , 228	+ CO
P5	242.0818	242.0812	2.6	C ₁₄ H ₁₂ NO ₃	212, 228 , 240	+ 2H + CO
P6	256.0614	256.0604	3.8	C ₁₄ H ₁₀ NO ₄	212, 226, 228 , 240, 242	+ CO ₂
P7	258.0758	258.0761	-1.2	C ₁₄ H ₁₂ NO ₄	212, 226, 228 , 240, 242, 256	+ 2H + CO ₂
P8	421.1176	421.1183	-1.6	C ₂₆ H ₁₇ N ₂ O ₄	212	2-OH-AC dimer

fragment ions are shown in bold.

^a Calculated using Molecular Mass Calculator freeware version v2.02.

Table 3

A summary of neutral losses and fragmentations for ions attributed to the respective glutathione S-conjugate of the 2-OH-AC.

MS/MS collision-induced dissociation of glutathione S-conjugate			2-OH-AC		
			Measured <i>m/z</i> / Da	Calculated <i>m/z</i> / Da ^a	Deviation / ppm
Neutral losses	Parent	<i>m/z</i> / Da	517.1382	517.1387	1.04
	Glycine	75.0320	None	442.1067	None
	Anhydroglutamic acid	129.0426	388.0963	388.0962	-0.39
	Glutamine	146.0691	None	371.0696	None
	γ-Glu-Ala-Gly	275.1117	244.0421 ^b	244.0427	2.34
	Glutathione (S-oxide)	322.0709	None	195.0679	None

^a Calculated using Molecular Mass Calculator freeware version v2.02.

^b Neutral loss of γ-Glu-Ala-Gly + 2H.

Figure captions**Figure 1**

Instrumental set-ups used for (a) the electrochemical simulation of the oxidative metabolism of 2-OH-AC and (b) the investigation of the reactivity of 2-OH-AC oxidation products towards GSH (NAC).

Legend: 1 – infusion syringe pump, 2 – potentiostat, 3 – electrochemical thin-layer cell (EC) – reactor cell, 4 – mass spectrometer (MS), 5 – T-piece and 100 μ L mixing coil

Chemical structures of the investigated 2-hydroxyacridinone (2-OH-AC) compound and glutathione (GSH)/*N*-acetylcysteine (NAC), used as nucleophilic trapping agents, are presented in the frame.

Figure 2

Cyclic voltammograms of 0.5 mM 2-OH-AC (a) alone and (b) in the mixture with 5 mM GSH in 0.02 M PBS buffer, pH 7.40, versus electrolysis progress. Experimental conditions: potential range -0.2 – 1.2 V; scan rate 100 $\text{mV}\cdot\text{s}^{-1}$; T = 21 $^{\circ}\text{C}$; ϕ GC 3 mm.

Figure 3

Two-dimensional plot (2-D mass voltammogram) of 10 μM 2-OH-AC oxidation at a GC working electrode (extracted ion intensity versus the progress of the electrochemical oxidation; positive-ion mode). The m/z ratios shown correspond to the protonated 2-OH-AC and its products (see legend). Experimental conditions: potential range 0 – 2.5 V; scan rate 10 $\text{mV}\cdot\text{s}^{-1}$, continuous; T = 21 $^{\circ}\text{C}$; ϕ GC 8 mm. The insets show the representative mass spectrum of 2-OH-AC without voltage applied to the electrochemical cell (cell off; top left corner) and after electrochemical oxidation (cell on; top right corner).

Figure 4

Two-dimensional plots (2-D mass voltammograms) of 10 μM 2-OH-AC oxidation at a GC working electrode in the presence of 100 μM (a) GSH and (b) NAC (extracted ion intensity versus the progress of the electrochemical oxidation; positive-ion mode). The m/z ratios shown correspond to the protonated 2-OH-AC and GSH or NAC S-conjugate (see legend).

1
2
3 Experimental conditions: potential range 0 – 2.5 V; scan rate 10 mV·s⁻¹, continuous; T = 21
4 °C; ϕ GC 8 mm.

6 7 **Figure 5**

8
9 The representative mass spectrum of a mixture of 2-OH-AC and GSH (a) without voltage
10 applied to the electrochemical cell (cell off) and (b) after electrochemical oxidation (cell on).
11
12 The insets show a zoom in the mass range from m/z 510 to 520 (positive-ion mode). (c)
13
14 Fragmentation spectrum of the conjugation product m/z 517.

16 17 **Figure 6**

18
19 (a) The representative high performance LC chromatograms of the reaction mixtures
20 obtained after metabolism simulation of 0.5 mM 2-OH-AC in the absence (black line) and the
21 presence (grey dashed line) of 5 mM GSH in RLM incubations (enzymatic oxidation) and
22 from controlled-potential electrolysis (oxidation by CPE). The inset shows UV-Vis spectra of
23 GSH S-conjugate and 2-OH-AC. (b) The representative extracted ion chromatograms for 2-
24 OH-AC oxidation in electrochemical cell.

30 31 **Figure 7**

32
33 Proposed oxidation reaction pathways of 2-OH-AC observed in the electrochemical cell.
34
35 Tentative structures were derived on the basis of accurate mass measurements and MS/MS
36 fragmentation patterns. Probable isomeric compounds are gathered in encircled blocks.

38 39 **Figure 8**

40
41 Schematic representation of the proposed mechanism of the 2-OH-AC electrochemical
42 oxidation and GSH (NAC) S-conjugate formation.

Figure 1

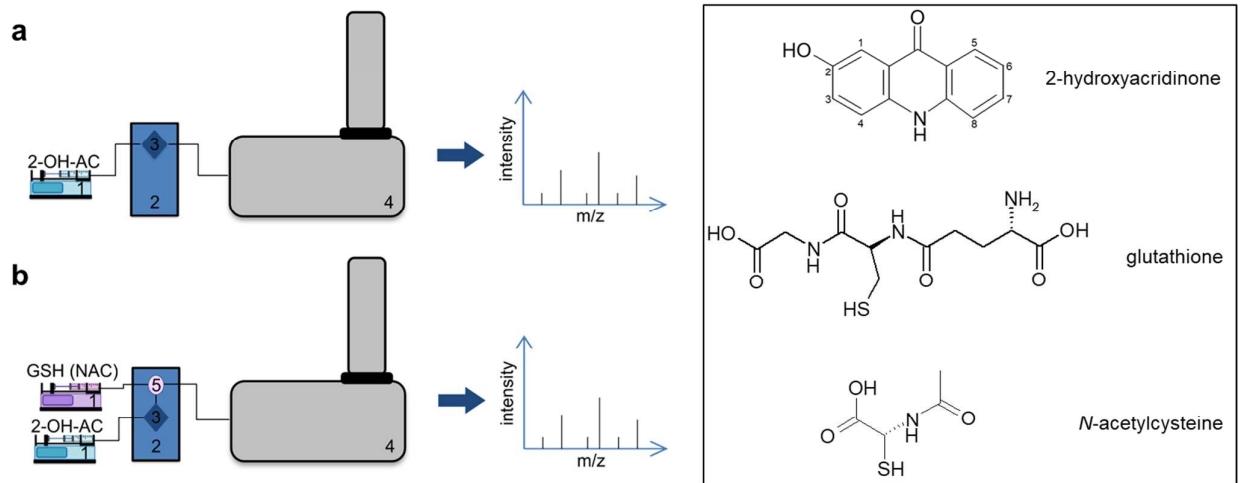


Figure 2

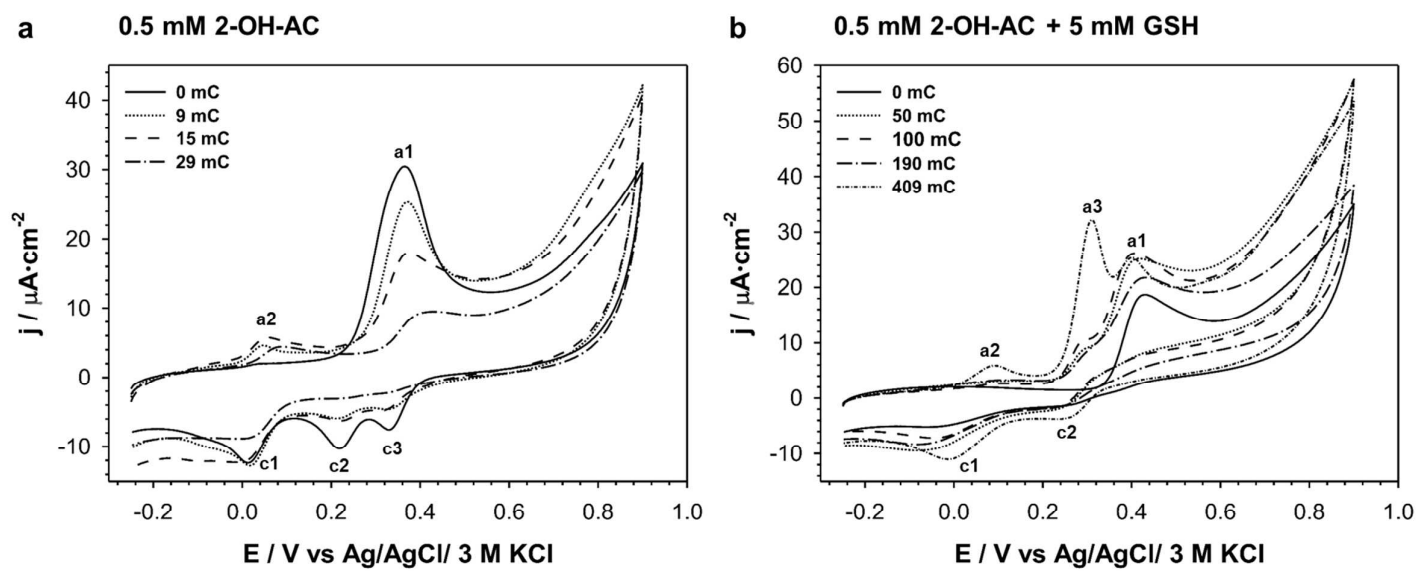


Figure 3

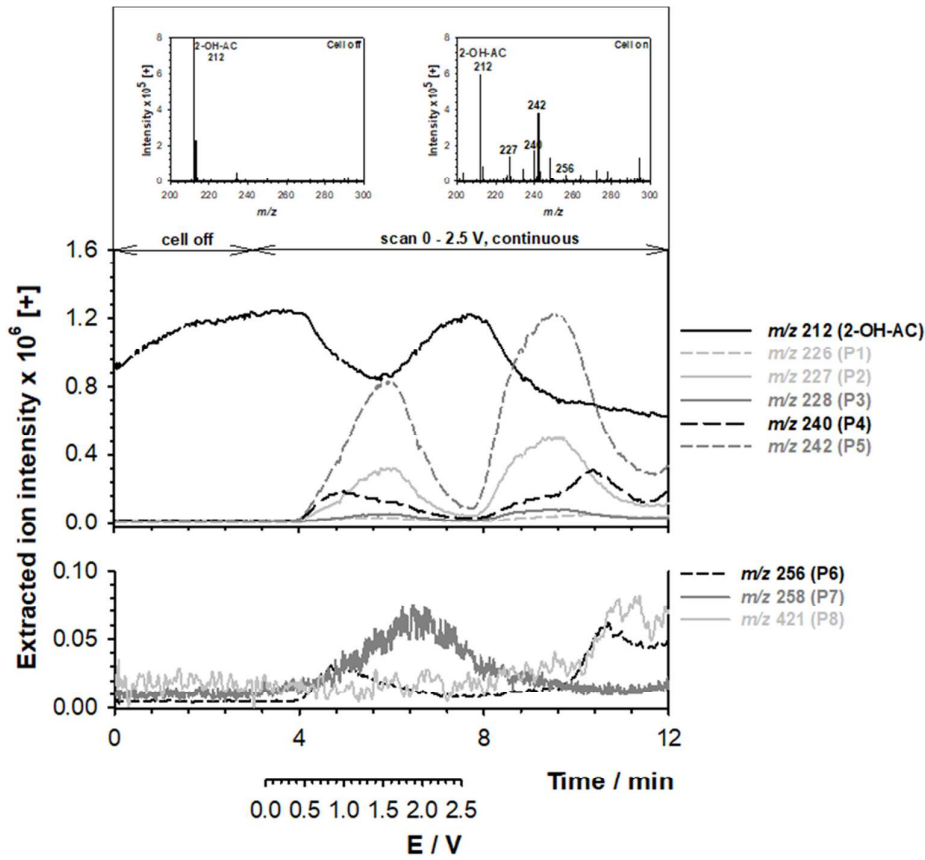


Figure 4

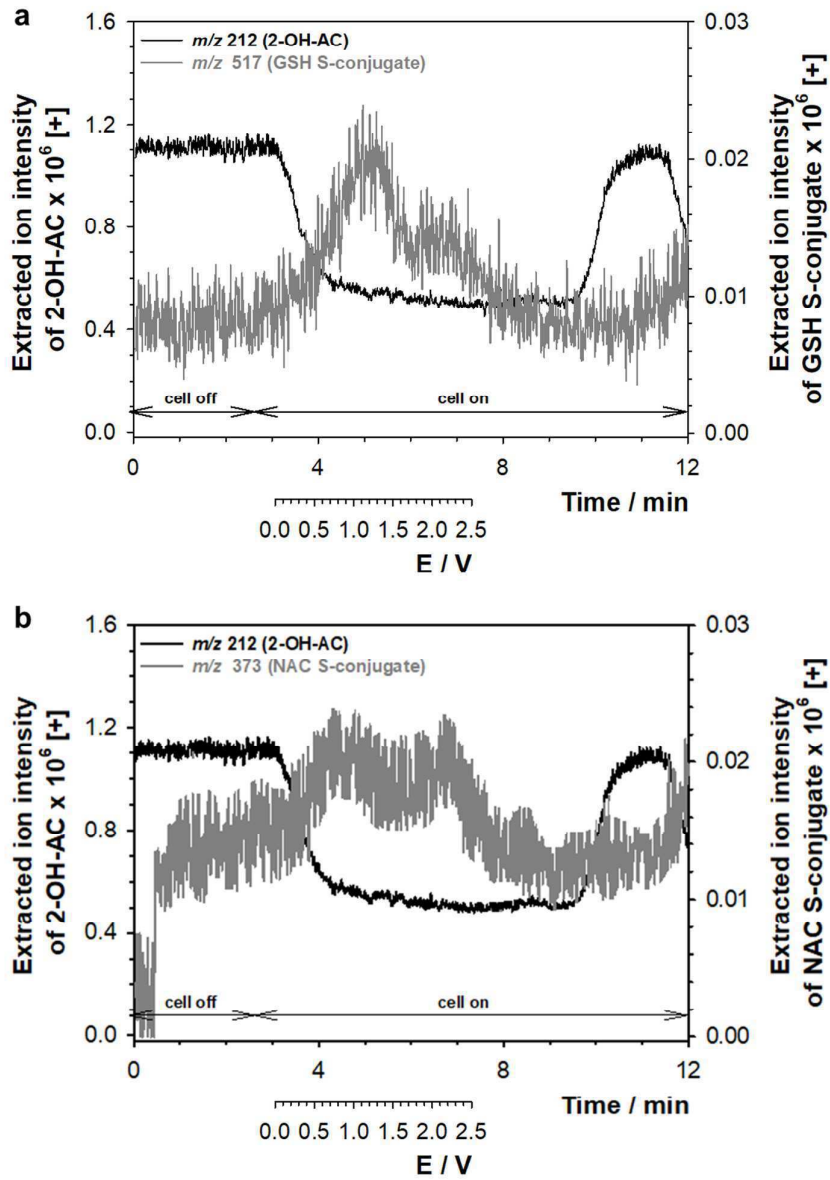


Figure 5

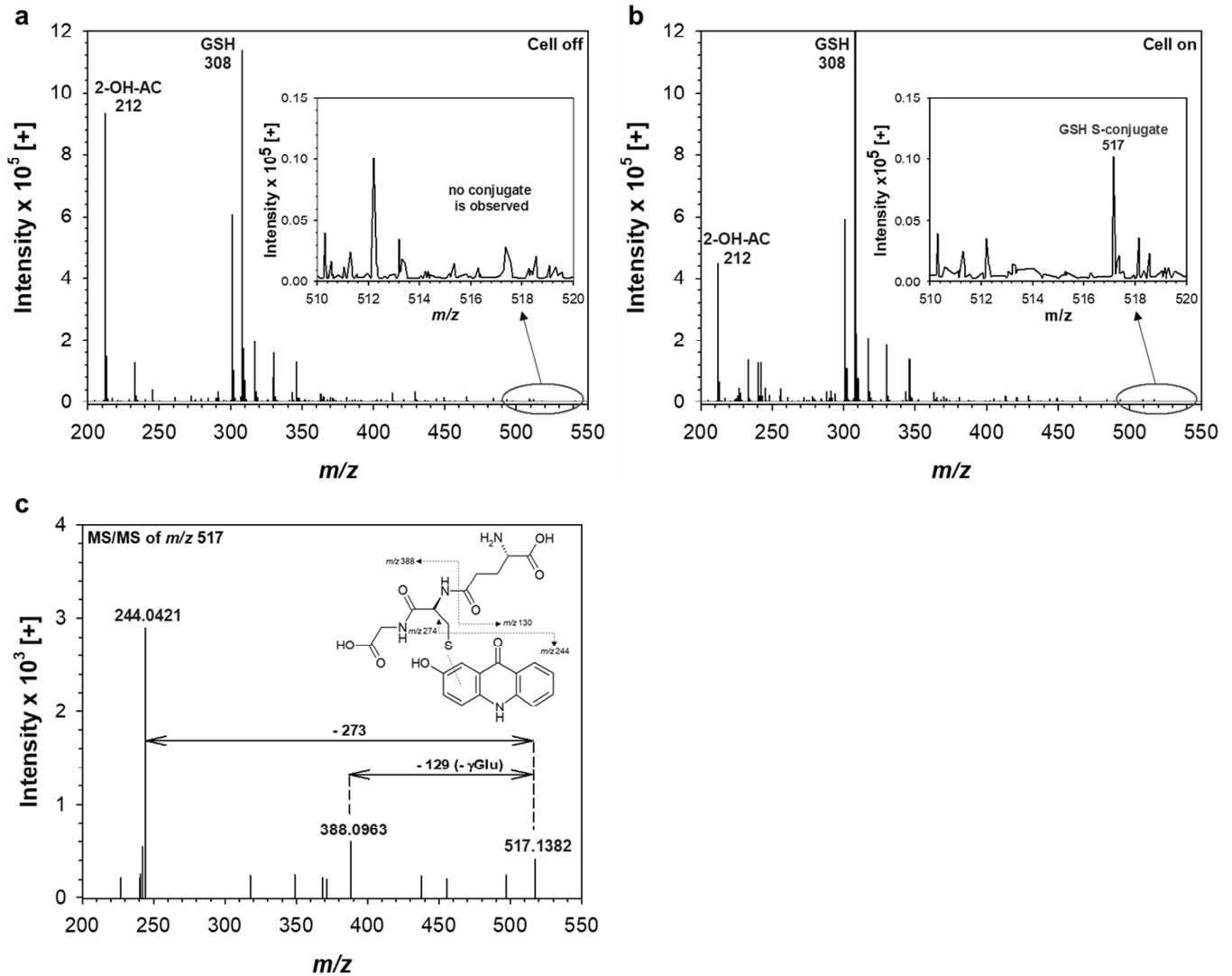


Figure 6

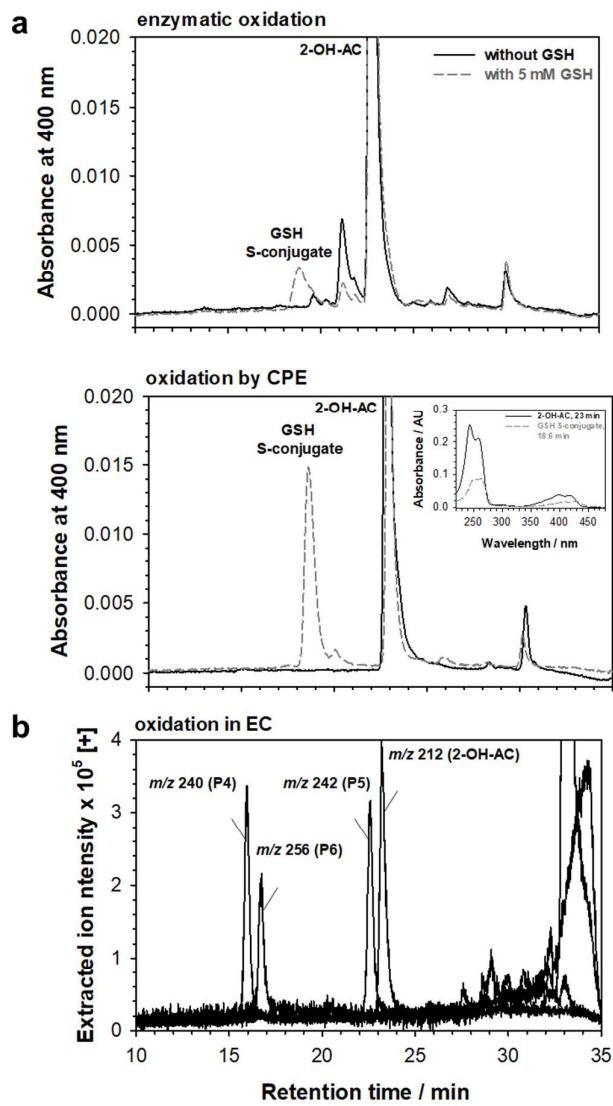


Figure 7

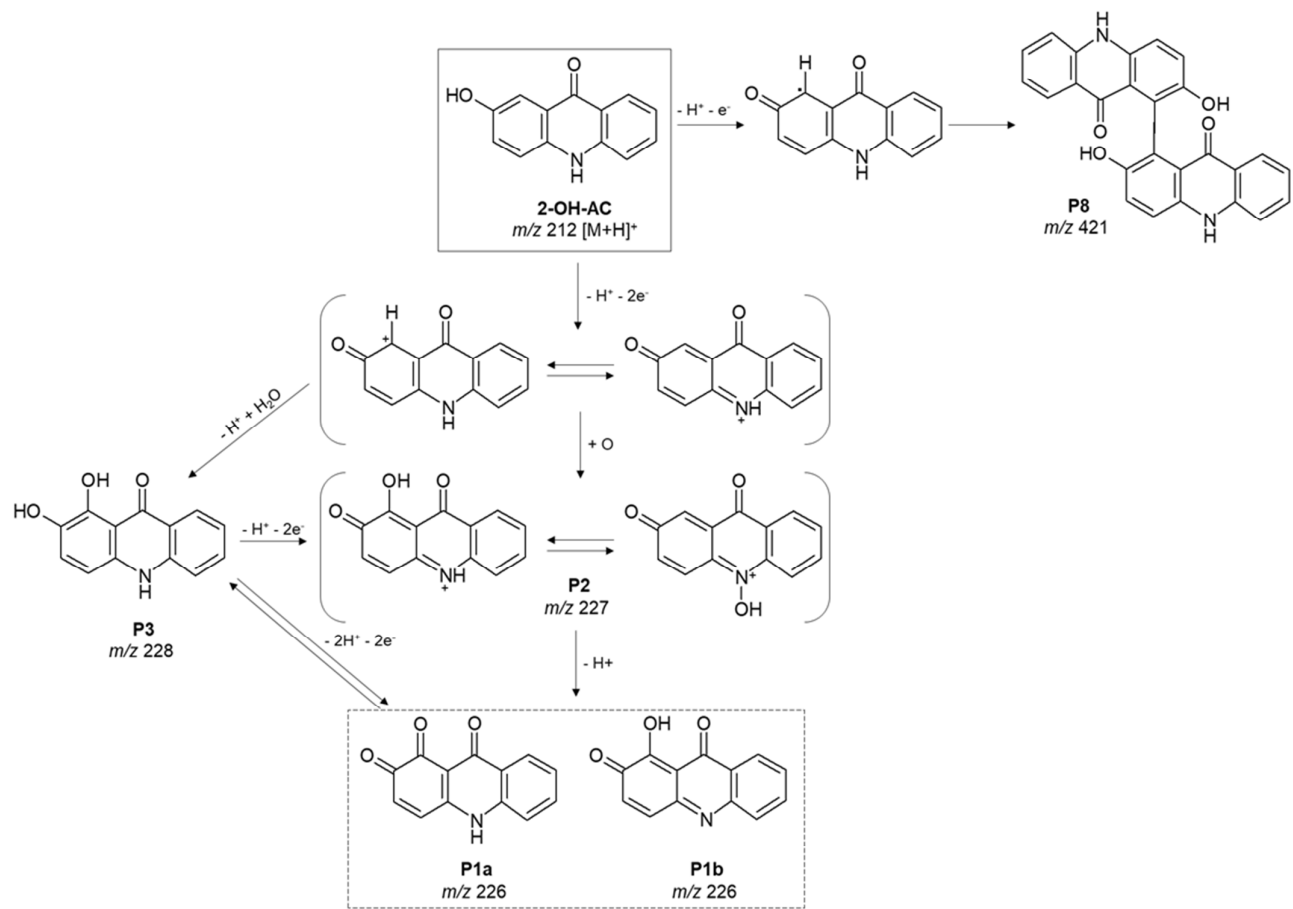


Figure 8

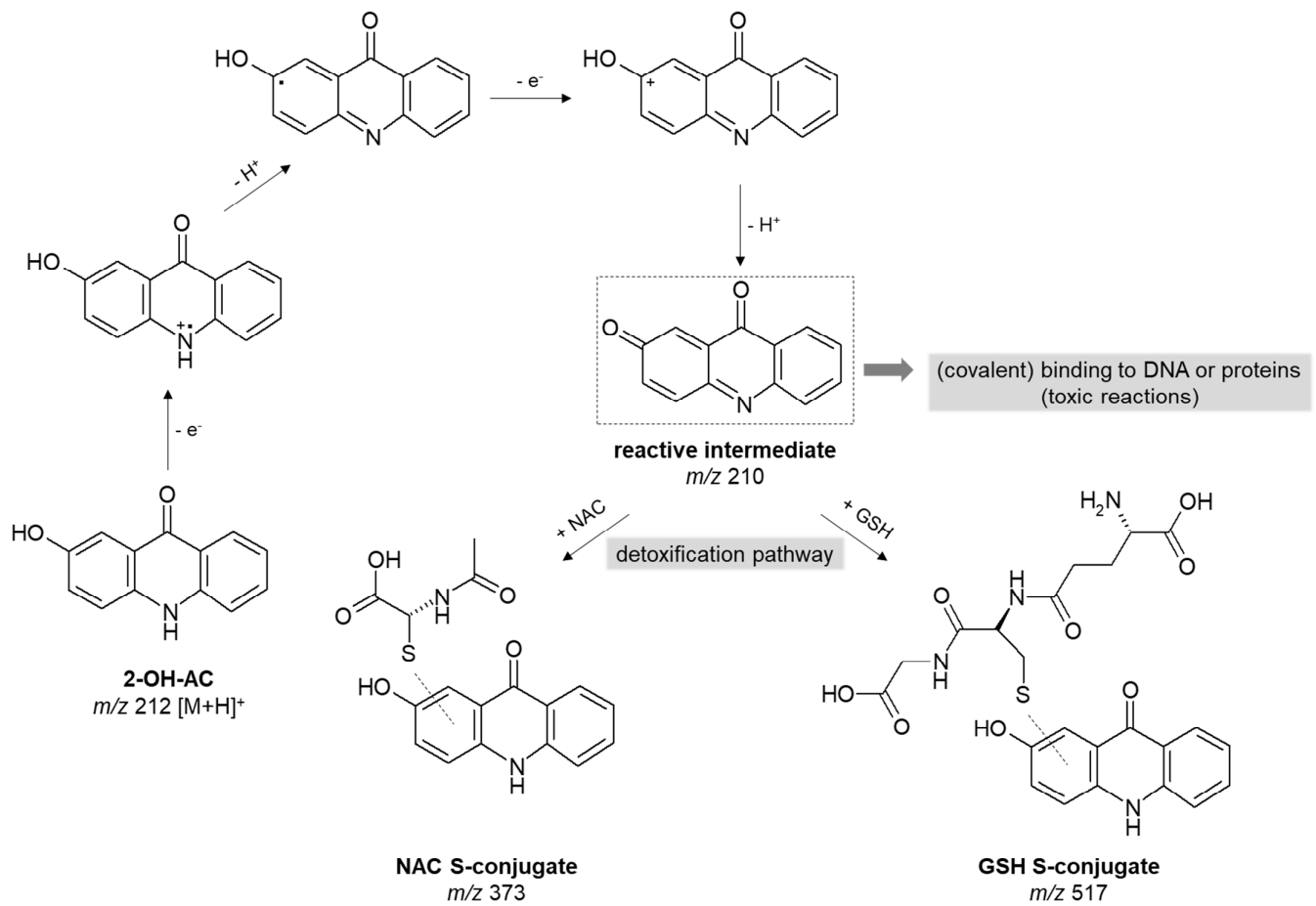


Table 1

	Parameter	Value or setting
EC settings	Flow rate	30 $\mu\text{L}\cdot\text{min}^{-1}$
	Potential	0 – 2.5 V (10 mV steps)
	EC operating mode	scan
	Cycle	continuous
MS(/MS) settings	The range of m/z	100 – 600
	Ion source	dual ESI
	Ion polarity	positive
	MS operating mode	scan
	Capillary voltage	3500 V
	Nebulizer gas (N_2) pressure	35 psig
	Drying gas (N_2) flow	10 $\text{L}\cdot\text{min}^{-1}$
	Drying gas temperature	325 $^{\circ}\text{C}$
	Fragmentor	175 V
	Skimmer	45 V
	OCT 1 RF Vpp	750 V
	Rate	1,5 $\text{spectra}\cdot\text{s}^{-1}$
	MS/MS method	targeted
- Slope	- 4 m/z	
- Offset	- 5 V	

Table 2

Product	Measured <i>m/z</i> / Da	Calculated <i>m/z</i> / Da ^a	Deviation / ppm	Molecular formula of [M+H] ⁺ ion	MS/MS fragment ions	Predicted modification of 2-OH-AC molecule
2-OH-AC	212.0712	212.0706	2.9	C ₁₃ H ₁₀ NO ₂	-	-
P1a, b	226.0501	226.0499	1.1	C ₁₃ H ₈ NO ₃	212	- 2H + O
P2	227.0578	227.0577	0.5	C ₁₃ H ₉ NO ₃	212	- H + O
P3	228.0648	228.0655	-3.1	C ₁₃ H ₁₀ NO ₃	212, 226, 227	+ O
P4	240.0658	240.0655	1.2	C ₁₄ H ₁₀ NO ₃	212, 227 , 228	+ CO
P5	242.0818	242.0812	2.6	C ₁₄ H ₁₂ NO ₃	212, 228 , 240	+ 2H + CO
P6	256.0614	256.0604	3.8	C ₁₄ H ₁₀ NO ₄	212, 226, 228 , 240, 242	+ CO ₂
P7	258.0758	258.0761	-1.2	C ₁₄ H ₁₂ NO ₄	212, 226, 228 , 240, 242, 256	+ 2H + CO ₂
P8	421.1176	421.1183	-1.6	C ₂₆ H ₁₇ N ₂ O ₄	212	2-OH-AC dimer

^a Calculated using Molecular Mass Calculator freeware version v2.02.

Table 3

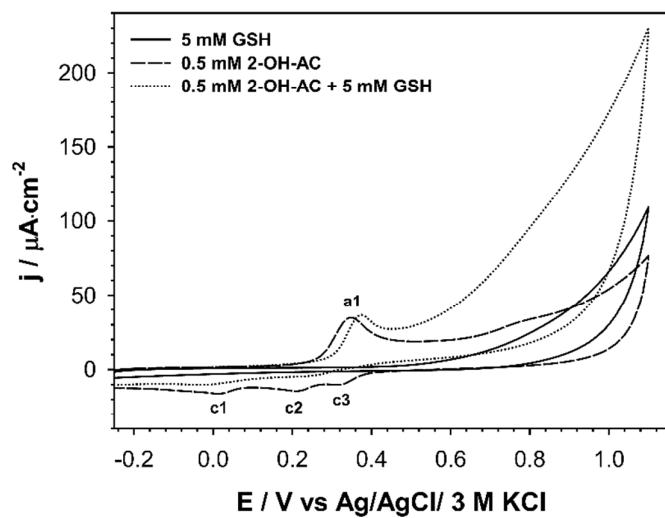
MS/MS collision-induced dissociation of glutathione S-conjugate			2-OH-AC		
			Measured <i>m/z</i> / Da	Calculated <i>m/z</i> / Da ^a	Deviation / ppm
Neutral losses	Parent	<i>m/z</i> / Da	517.1382	517.1387	1.04
	Glycine	75.0320	None	442.1067	None
	Anhydroglutamic acid	129.0426	388.0963	388.0962	-0.39
	Glutamine	146.0691	None	371.0696	None
	γ -Glu-Ala-Gly	275.1117	244.0421 ^b	244.0427	2.34
	Glutathione (S-oxide)	322.0709	None	195.0679	None

^a Calculated using Molecular Mass Calculator freeware version v2.02.

^b Neutral loss of γ -Glu-Ala-Gly + 2H.

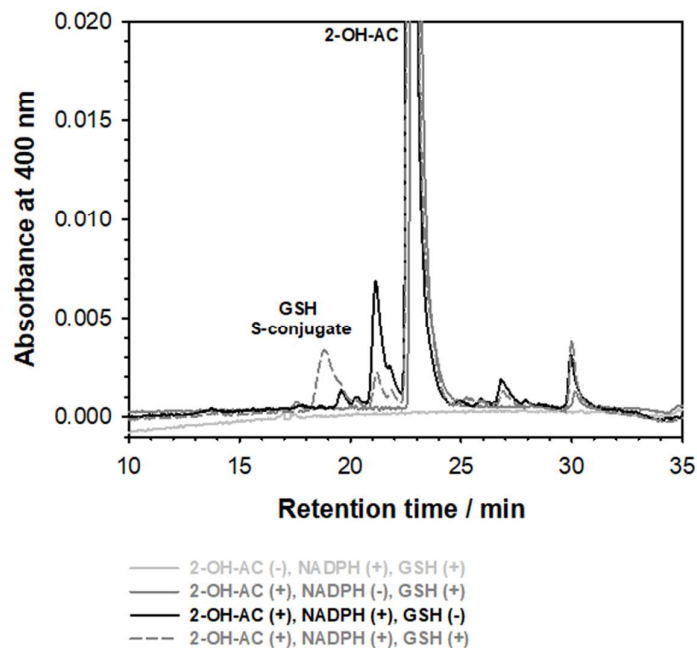
Figure

Cyclic voltammograms of 0.5 mM 2-OH-AC and 5 mM GSH alone and in the mixture in 0.02 M PBS buffer, pH 7.40. Experimental conditions: potential range -0.2 – 1.2 V; scan rate 100 $\text{mV}\cdot\text{s}^{-1}$; $T = 21\text{ }^\circ\text{C}$; ϕ GC 3 mm (supplemental data for Figure 2).



Figure

The representative high performance LC chromatograms of the control incubations prepared for enzymatic oxidation of 2-OH-AC in RLMs (supplemental data for Figure 6a).



Table

A summary of the electrochemical products observed under different electrolyte conditions.

Product / <i>m/z</i> / Da	Electrolyte composition		
	(1) 0.1% formic acid in water/methanol (50/50, v/v)	(2) 0.1% formic acid in water/acetonitrile (50/50, v/v)	(3) 20 mM ammonium formate (pH 3.40) in water/acetonitrile (50/50, v/v)
P1 / 226	+ ^a	+ ^a	++ ^b
P2 / 227	++ ^b	+ ^a	+ ^a
P3 / 228	+ ^a	++ ^b	++ ^b
P4 / 240	++ ^b	ND ^c	ND ^c
P5 / 242	++ ^b	+ ^a	+ ^a
P6 / 256	+ ^a	+ ^a	++ ^b
P7 / 258	+ ^a	+ ^a	+ ^a
P8 / 421	+ ^a	ND ^c	ND ^c

Ions showing the highest intensity in a given electrolyte are shown in bold. ^a +: low intensity, ^b ++: high intensity, ^c ND: not detected.